

## ENZYMES FROM DEEP-SEA MICROORGANISMS

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**Keywords:** Mariana Trench, Challenger Deep, Deep-sea environments, Microbial flora, Bacteria, Actinomycetes, Yeast, Fungi, Extremophiles, Halophiles, 16S rDNA, Phylogenetic tree, Psychrophiles, Thermophiles, Alkaliphiles, Amylase,  $\alpha$ -maltotetraohydase (G4-amylase), Protease, *Pseudomonas* strain MS300, Hydrostatic pressure, *Shinkai 2000*, *Shinkai 6500*, *Kaiko*

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### Summary

In an attempt to characterize the microbial flora on the deep-sea floor, we isolated thousands of microbes from samples collected at various deep-sea (1 050–10 897 m) sites located in the Mariana Trench and off southern Japan. Various types of bacteria, such as alkaliphiles, thermophiles, psychrophiles, and halophiles were recovered on agar plates at atmospheric pressure at a frequency of  $0.8 \times 10^2$ – $2.3 \times 10^4$ /g of dry sea mud. No acidophiles were recovered. Similarly, non extremophilic bacteria were recovered at a frequency of  $8.1 \times 10^2$ – $2.3 \times 10^5$ . These deep-sea isolates were widely distributed and detected at each deep-sea site, and the frequency of isolation of microbes from the deep-sea mud was not directly influenced by the depth of the sampling site. Phylogenetic analysis of deep-sea isolates based on 16S rDNA sequences revealed that a wide range of taxa were represented in the deep-sea environment. Among these deep-sea isolates, we have isolated a *Pseudomonas*-like amylase producer, strain MS300, which displayed a large halo on starch medium. Strain MS300 produced two major and two minor  $\alpha$ -maltotetraohydrolases (G4-amylase). The two major G4-amylases share the same molecular weight of 55 000 but had different pI values of 5.0 and 4.7, respectively. The optimum temperature for activity of both major G4-amylases is 40°C,

and the optimum pH is 6.8 for one and 8.9 for the other. MS300 produced more amylase under higher hydrostatic pressure than under atmospheric pressure.

## 1. Introduction

The major environment of the Earth is the deep-sea environment, at an average depth of 3 800 m, because the ocean occupies 70% of the earth. The deep sea is an extreme environment in terms of land plants and animals, including human beings, as it is characterized by high hydrostatic pressure, low temperature, and lack of light. However, the Earth's surface, exposed to strong UV light, desiccation, and atmospheric pressure, would be an extreme environment for organisms living in the deep sea. Microorganisms living in the deep sea have developed specific characteristics that allow them to thrive in that environment. Bacteria have been isolated from deep-sea mud and from benthic organisms, such as amphipods and sea cucumbers in the bathypelagic zone. However, little information is available on the bacterial diversity and the distribution of useful microbes for industrial application in sediments of the deep-sea floor.

On March 2, 1996, the 3 m long unmanned submersible *Kaiko* touched the bottom of the Challenger Deep in the Mariana Trench and successfully scooped out a mud sample, the first obtained at a depth of 10 897 m. We isolated thousands of microbes from this deep-sea mud sample and found that the microbial flora contained was composed of actinomycetes, yeasts, and a range of bacteria including various extremophilic genera.

To explore in greater detail the microbial diversity in various shallower deep-sea environments, we attempted to isolate and characterize a number of bacteria from deep-sea mud collected by the manned submersibles *Shinkai 2000* and *Shinkai 6500*. We recorded considerable bacterial diversity and the occurrence of extremophilic bacteria at several deep-sea sites located off the southern part of Japan. It is well known that some microbes produce different types of enzyme depending on the growth environment. Thus, it is expected that the deep sea will be a good source for new types of enzyme producers, because the deep-sea ecosystem differs from that on land. We therefore attempted to explore unique enzyme-producing bacteria from deep-sea isolates and focused on new types of amylase producers, as the first step in industrial application of deep-sea microbes.

## 2. Collection of Deep-Sea Mud

Samples of deep-sea sediment were collected from the Iheya Ridge, Nankai Islands (1 050 m depth, 27°47.18'N, 126°54.15'E), using the manned submersible *Shinkai 2000* and from the Izu-Ogasawara Trench (2 759 m, 30°07.05'N, 139°58.42'E; and 3 400 m depth, 29°04.2'N, 140°43.3'E) using the manned submersible *Shinkai 6500*, employing cylinder mud samplers (Figure 1). The 3 m long submersible *Kaiko* touched the bottom of the Challenger Deep and successfully scooped out a mud sample, the first in the world obtained at the great depth of 10,897 m, as shown in Figure 2B, C, and D. A sterile 50 ml-Falcon tube filled with a very fine mud of grayish-brownish particles was inserted tightly in a tube holder and brought to the sea surface without being contaminated by upper-ocean bacteria (Figure 2A and D).

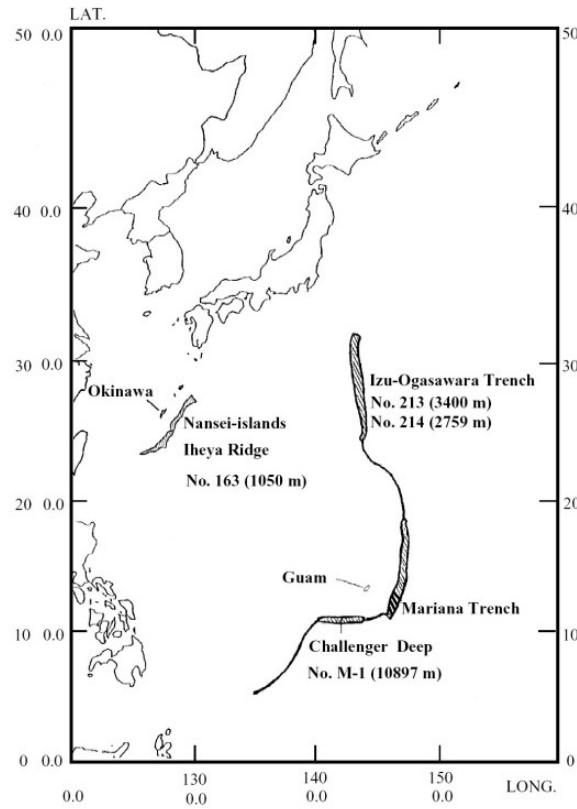


Figure. 1. Deep-sea sites for collection of mud samples.

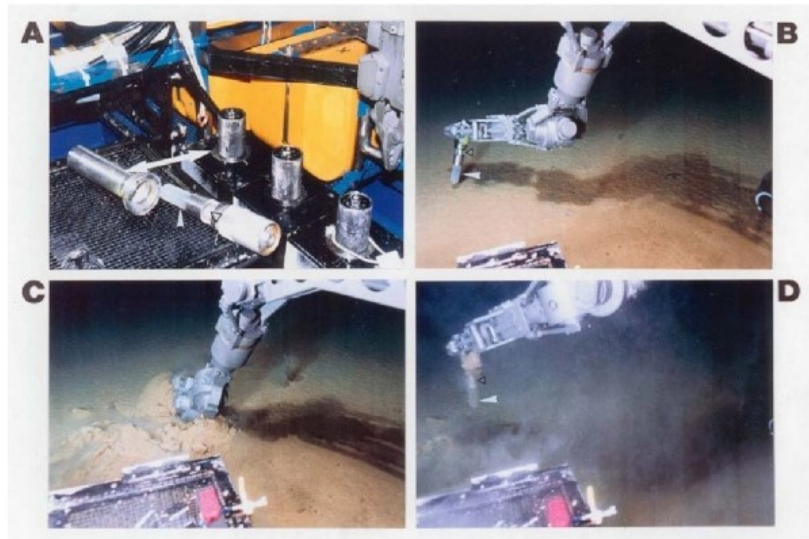


Figure 2. A, B, C, and D. Collection of a mud sample from a depth of 10,897 m in the Challenger Deep.

### 3. Isolation of Microorganisms From Deep-Sea Mud

The ChallengerDeep mud sample was diluted two-fold with Marine broth 2216, and 100 to 200  $\mu\text{l}$  (5 to 10 mg as dry weight) of the suspension was spread on Marine agar or

half-strength nutrient agar plates used as a basal medium. In addition, modified Marine agar plates supplemented with 1% skimmed milk or 1% potato starch, which had varied pH values (pH 3, 7, or 10) and concentrations of sodium chloride (0%, 2%, or 15%), were used for isolation. The alkaline and acidic media contained 1% sodium carbonate and 50 mM citric acid, respectively. The alkaline or acidic stock solution concentrated 10-fold was autoclaved separately and then added to the plain or modified Marine agar medium. The agar plates were incubated at 4° to 75°C at atmospheric pressure (0.1 MPa) or at 100 MPa for 1 to 4 weeks. Cultivation techniques, at high pressure, were as described previously by authors from our laboratory.

### 3.1. Bacteria From The Mariana Trench

Various non-extremophilic bacteria were isolated from the mud sample with a frequency of  $2.2 \times 10^4$  to  $2.3 \times 10^5$  colonies per gram of dry sea mud under the conditions described in Table 1. This is the first report of the isolation of viable bacteria from the bottom of the Challenger Deep and is significant to our understanding of the nature of microbial flora in the deepest-sea mud. Previously, by means of microscopic observations, ZoBell confirmed the existence of viable bacterial cells in mud collected at a depth of 10 400 m from the Philippine Trench, but none of those bacteria were cultivated. We have obtained thousands of isolates from the Challenger Deep. Some were found to be producers of enzymes, such as proteases and amylases (Figure 3A and B). Although barophilic, halophilic, and acidophilic bacteria have not yet been detected in mud samples from the Challenger Deep, various other types of extremophilic bacteria were isolated at 0.1 MPa (Table 1). Among the extremophiles found were numerous alkaliphiles and thermophiles that could be expected to thrive in an extreme environment different from the *in situ* conditions of high hydrostatic pressure and low temperature in the Challenger Deep. The population of psychrophilic isolates tended to be smaller than those of alkaliphiles and thermophiles. In addition, we successfully isolated filamentous fungi and actinomycetes at 0.1 MPa at almost the same frequency ( $2.0 \times 10^2$  per gram of dry sea mud) as that of psychrophilic bacteria.



Figure 3. A and B. Deep-sea enzyme-producing isolates grown on modified Marine agar plates.

Category	Isolation conditions	Origin (Depth)	No.	Bacterial recovered (colonies g <sup>-1</sup> dry sea mud)
<b>Alkaliphile</b>	<b>pH 9.7 ± 0.3</b> 25°C, 0.1 MPa	163 (1050m) 214 (2759m) 213 (3400m) MI (10897m)		3.0-6.1 x 10 <sup>2</sup> 0.2-2.3 x 10 <sup>4</sup> 0.9 x 10 <sup>2</sup> 0.4-1.2 x 10 <sup>3</sup>
<b>Thermophile</b>	<b>55°C</b> pH 7.3 ± 0.2, 0.1 MPa	163 (1050m) 214 (2759m) 213 (3400m) MI (10897m)		0.8-2.3 x 10 <sup>2</sup> 1.1-7.8 x 10 <sup>2</sup> 1.0-6.0 x 10 <sup>2</sup> 0.6-3.5 x 10 <sup>3</sup>
<b>Psychrophile</b>	<b>4°C</b> pH 7.3 ± 0.2, 0.1 Mpa	163 (1050m) 214 (2759m) 213 (3400m) MI (10897m)		0.8-5.3 x 10 <sup>2</sup> 1.4-7.8 x 10 <sup>2</sup> 1.0 x 10 <sup>2</sup> 2.0 x 10 <sup>2</sup>
<b>Halophile</b>	<b>15%-NaCl, 25°C</b> pH 7.3 ± 0.2, 0.1 Mpa	163 (1050m) 214 (2759m) 213 (3400m) MI (10897m)		4.6 x 10 <sup>2</sup> 3.6 x 10 <sup>3</sup> 0.9 x 10 <sup>2</sup> —————
<b>Acidophile</b>	<b>pH 3.7 ± 0.2</b> 25°C, 0.1 MPa	163 (1050m) 214 (2759m) 213 (3400m) MI (10897m)		————— ————— ————— —————
<b>Non-extremophile</b>	25°C, 0.1 Mpa pH 7.3 ± 0.2	163 (1050m) 214 (2759m) 213 (3400m) MI (10897m)		0.5-6.6 x 10 <sup>3</sup> 0.2-1.1 x 10 <sup>5</sup> 8.1-9.4 x 10 <sup>2</sup> 0.2-2.3 x 10 <sup>5</sup>

—————: no growth obtained

Table 1. Isolation of extremophilic bacteria from several deep-sea sites.

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### Bibliography

Abe F, Horikoshi K (1995) Hydrostatic pressure promotes the acidification of vacuoles in *Saccharomyces*

*cerevisiae*. *FEMS Microbiol Lett* **130**, 307-312 [This paper demonstrates the cultivation method for microbes under high hydrostatic pressure].

DeLong EF, Yayanos AA (1985) Adaptation of the membrane lipids of a deep-sea bacterium to changes in hydrostatic pressure. *Science* **228(4703)**, 1101-1103 [This paper discusses the flexibility of membrane lipids under various hydrostatic pressures].

Fujita M, Torigoe K, Tsusaki K, Kubota M, Sakai S, Tsujisaka Y (1989) Cloning and nucleotide sequence of the gene (amyP) for maltotetraose-forming amylase from *Pseudomonas stutzeri* MO-19. *J Bacteriol* **171(3)**, 1333-1339 [This paper is a good example of maltotetraose-forming amylase produced by the microbe isolated from the land].

Higgins DG, Sharp PM (1988) Clustal: a package for performing multiple alignment on a microcomputer. *Gene* **73**, 237-244 [This paper describes a method for multiple alignment of the sequence to construct phylogenetic tree].

Lu J, Nogi Y, Takami H (2001) *Oceanobacillus iheyensis* gen. nov. sp. nov., a deep-sea extremely halotolerant and alkaliphilic species isolated from 1,050 m depth of Iheya Ridge. *FEMS Microbiol Lett* **205**, 291-297 [This paper presents information on the taxonomy of a newly isolated bacterium from a depth of 1 050 m on the Iheya Ridge].

Jannash HW, Taylor CD (1984) Deep-sea Microbiology. *Ann. Rev. Microbiol.* **38**, 487-514 [This review addresses important concepts in understanding microbial life in deep-sea environments].

Kato C, Smorawinska M, Li L, Horikoshi K (1997) Comparison of the gene expression of aspartate  $\beta$ -D-semialdehyde dehydrogenase at elevated hydrostatic pressure in deep-sea bacteria. *J Biochem* **121**, 717-723 [This paper presents a good example of an enzyme activated by pressurization].

Kaneshiro SM and Clark DS (1995) Pressure effects on the composition and thermal behavior of lipids from the deep-sea thermophile *Methanococcus jannaschii*. *J Bacteriol* **177(13)**, 3668-3672. [This paper demonstrates the change of core lipid composition and temperature dependence of fluidity by pressure].

Kobayashi H, Takaki Y, Kobata K, Takami H, Inoue A (1998) Characterization of  $\alpha$ -maltotetraohydrolase produced by *Pseudomonas* sp. MS300 isolated from the deepest site of the Mariana Trench. *Extremophiles* **2**, 401-407 [This paper is the first report an amylase-producing bacterium isolated from the deepest mud of Mariana Trench].

Nagahama T, Hamamoto M, Nakase T, Takami H, Horikoshi, K (2001) Distribution and identification of red yeasts in deep-sea environments around the northwest Pacific Ocean. *Antonie van Leeuwenhoek* **80**, 101-110 [This paper demonstrates the biodiversity of yeasts isolated from deep-sea environments].

O'Sullivan J, McCullough J, Johnson JH, Bonner DP, Clark JC, Dean L, Trejo WH (1990) Janthinocins A, B and C, novel peptide lactone antibiotics produced by *Janthiobacterium lividum*. I. Taxonomy, fermentation, isolation, physicochemical and biological characterization. *J Antibiot* **43**, 913-919 [This paper presents information on the taxonomy of novel peptide lactone antibiotics producer, *Janthiobacterium lividum*].

Roby J, Ackerman R (1971) Isolation, purification and characterization of  $\alpha$  maltotetraose-producing amylase from *Pseudomonas stutzeri*. *Arch Biochem Biophys* **145**, 105-109 [This paper describes the unique properties of maltotetraose-producing amylase from *Pseudomonas stutzeri*].

Saitou N, Nei M (1987) A neighbor-joining method: a new method for reconstructing phylogenetic trees. *Mol. Biol. Evol.* **44**, 406-425 [This paper demonstrates a new method for constructing phylogenetic trees].

Sakano Y, Kashiyama E, Kobayashi T (1983) Purification of  $\alpha$ -maltotetraose-forming exo-amylase of *Pseudomonas stutzeri*: two-forms of the amylase and their enzymatic properties. *Agric Biol Chem* **47(8)**, 1761-1768 [This paper presents different type of  $\alpha$ -maltotetraose-forming exo-amylase from *Pseudomonas stutzeri*].

Schmidt J, John M (1979) Starch metabolism in *Pseudomonas stutzeri*: I. Studies on maltotetraose-forming amylase. *Biochem Biophys Acta* **566**, 88-89 [This paper describes the first isolation of maltotetraose-producing amylase from *Pseudomonas stutzeri*].

Takagawa S, Takahashi K, Sano T, Mori Y, Nakanishi T, Kyo M (1989) 6500 m deep manned research submersible *Shinkai 6500* system. *Oceans* **89**, 741-746 [This paper illustrates the performances of manned research submersible *Shinkai 6500* system].

Takami H, Inoue A, Fuji F, Horikoshi K (1997) Microbial flora in the deepest sea mud of the Mariana trench. *FEMS Microbiol Lett* **152**, 279-285. [This paper is the first report dealing with the isolation of various bacterial species from the deepest point of the Mariana Trench].

Takami H, Kobata K, Nagahama T, Kobayashi H, Inoue A, Horikoshi K (1999) Biodiversity in deep-sea sites near the south part of Japan. *Extremophiles* **3**, 97-102 [This paper provides an overview of the biodiversity of deep-sea isolates from different sampling points near the south part of Japan].

Yayanos AA, (1979) Isolation of a Deep-Sea Barophilic Bacterium and Some of Its Growth Characteristics. *Science* **205**, 808-810 [This paper describes the first isolation of a barophilic bacterial species from the deep-sea environment].

Yayanos AA (1981) Obligately barophilic bacterium from the Mariana Trench. *Proc Natl Acad Sci USA* **78**, 5212-5215 [This paper describes the first isolation of an obligately barophilic bacterial species from the Mariana Trench].

Yayanos AA (1995) Microbiology to 10,500 meters in the deep sea. *Ann. Rev. Microbiol.* **49**, 777-805 [This review provides a large amount of background information on deep-sea microbiology].

ZoBell, C. E. (1941) Studies on marine bacteria. I. The cultural requirements of hetero-trophic aerobes. *J. Mar. Res.* **4**, 42-75. [This paper demonstrates the effect of nutrients and trace elements on the growth of marine bacteria, and optimum growth conditions for general marine bacteria].

ZoBell, C. E. (1952) Bacterial life at the bottom of the Philippine Trench. *Science* **115**, 507-508. [This paper deals with the microscopic observation of viable bacteria in the deep-sea mud collected at a depth of 10 400 m from Philippine Trench].

### Biographical Sketch

**Hideto Takami** received his Bachelors degree in agricultural chemistry and education from Tokyo University of Agriculture in 1984, and his M.S. degree in 1986 from Yamanashi University. He then worked for one and a half year at Kurita Water Industry Co. Ltd. and for three and a half years at RIKEN Institute as a visiting research associate. He received a Ph.D. degree in 1993 from Tokyo Institute of Technology, Department of Bioengineering. He next went to Michigan State University at East Lansing, where he carried out research as a postdoctoral fellow (1993-1995) on the distribution of 2,4-D degrading bacteria in the pristine soil and 2,4-D metabolic pathways. In May 1995, he joined the department of Frontier Research System for Extremophiles at Japan Marine Science and Technology Center (JAMSTEC), where he is currently senior scientist and group leader of Microbial Genome Research Group. In 1997, he and his colleagues published the first paper on the microbial flora of the deepest mud of Mariana Trench, Challenger Deep. In 1998, he initiated the whole genome sequencing project of alkaliphilic *Bacillus halodurans* which produces various kinds of enzymes. His group completed the sequencing project of *B. halodurans* genome in 2000. His research interests include the adaptation mechanisms of extremophilic bacteria and the evolutionary process of the genomes of microorganisms. He lives with his wife, Yumiko, in Yokohama city.