# PHYSIOLOGICAL REGULATION OF GENE ACTIVITY BY OXYGEN (O<sub>2</sub>)

### Juha-Pekka Pursiheimo

Turku Centre for Biotechnology, Turku, Finland

**Keywords:** Genes, Hypoxia, Hypoxia inducible factor, Erythropoietin, Heart failure, Tumors, Hyperoxia, Hyperbaric oxygen therapy, Reactive oxygen species, Antioxidants

#### Contents

- 1. Introduction
- 2. HIF: transcriptional regulator of hypoxic responses
- 3. Oxygen-dependent regulation of HIF
- 3.1. Oxygen-dependent regulation of HIF $\alpha$  stability
- 3.2. Oxygen-dependent regulation of HIF transcriptional activity
- 4. Reactive Oxygen Species (ROS) and cellular responses
- 5. Oxygen and disease progression
- 5.1. Solid tumors
- 5.2. Myocardial Ischemia
- 5.3. Alveolar cell death
- 5.4. Reperfusion injury
- Glossary
- Bibliography

Biographical Sketch

#### Summary

All eukaryotic cells sense the decrease in oxygen level and respond to hypoxic-stress by regulating transcription of a wide variety of genes by the heterodimeric Hypoxia-Inducible Factor (HIF). Perhaps 1–4% of all human genes are expressed in response to hypoxia in a HIF-dependent manner. Both subunits of HIF, HIF $\alpha$  and HIF $\beta$  are constitutively expressed, but the HIF $\alpha$  is quickly degraded in normally oxygenated cells and the activation of HIF target genes is blocked. During hypoxia degradation attenuates, HIF $\alpha$  is stabilized and binds to DNA with HIF $\beta$  leading to an activation of array of genes involved in hypoxic adaptation. The Von Hippel-Lindau protein (pVHL) is required for the ubiquitination of HIF $\alpha$  once these proteins have physically interacted and this targets HIF $\alpha$  to proteasomal degradation. The mechanism of cellular oxygen sensing is bound to this interaction, since the recognition and binding of pVHL to HIF $\alpha$  requires oxygen-dependent hydroxylation of specific prolyl residues on HIF $\alpha$ . Three human HIF prolyl hydroxylases (PHD1 to 3) able to hydroxylate HIF $\alpha$  have been identified. The activity of HIF prolyl hydroxylases is directly regulated by oxygen availability indicating that these enzymes function as cellular oxygen sensors.

Aerobically growing cells are exposed to reactive oxygen species (ROS) which damage proteins, lipids and DNA (see *Metabolism of Oxygen*). Oxidative stress occurs when cellular defense mechanisms are unable to cope with existing ROS, and it has been associated with a number of pathologies including cancer and cardiovascular disease.

Since aerobic life is restricted within a very narrow range of oxygen concentrations, cells have been forced to develop highly sophisticated mechanisms to respond to fluctuations in oxygen tension. In this review, cellular mechanisms to cope with both hypoxic and hyperoxic stresses are discussed.

## 1. Introduction

For all aerobic life molecular oxygen  $(O_2)$  is essential for survival, since the energy production of aerobic organisms fundamentally depends on the oxygen bound cell respiration. In the process of oxidative phosphorylation, where the oxidoreductive energy of mitochondrial electron transport is converted into the chemical energy of ATP, oxygen functions as the ultimate electron acceptor. At the final step of the electron transport chain, cytochrome c reduces oxygen into water; this completes the process by which glucose is oxidized to carbon dioxide and water.

Decrease in oxygen availability directly impairs energy production and hence, cell viability. On the other hand excessive oxygen level is highly toxic through the generation of reactive oxygen species (ROS) which directly damage proteins and lipids in cell membranes as well as carbohydrates and genetic material. Thus aerobic life is restricted within a very narrow range of oxygen concentration. Because of its crucial role in sustaining aerobic life elaborated strategies have evolved to respond to decreased (hypoxia) or increased (hyperoxia) oxygen tensions. Cells have developed highly sophisticated systems to respond to variations in oxygen tension, leading to the activation of specific intracellular programs by induction of defined transcription factors and expression of appropriate target genes.

Fluctuations in oxygen levels in the blood stream are sensed by the carotic body (CB), the small organ located at the bifurcation of the common carotid artery close to the heart. Freshly oxygenated blood returns from the lung circulation pumped by the heart to the brain via the carotid artery. Even a small decrease in the blood oxygen level is sensed instantaneously, involving only rapid modifications of pre-existing proteins. Signals launched initiate systemic responses including respiratory and cardiovascular reflexes to ensure proper oxygenation in the vital organs. Prolonged decrease in oxygen availability activates chronic responses and initiates metabolic alterations to ensure cell and organ viability and functionality under hypoxic stress. These responses include stimulation of red blood cell production (erythropoiesis) as well as formation of new blood vessels (angiogenesis) into the hypoxic tissues, improving the efficiency of oxygen delivery.

The gene expression profile of hypoxic cells differs distinctively from normoxic cells. Cell specific sets of hypoxia-responsive genes are activated and production of proteins required to cope with lowered oxygen concentration is induced. Almost all genes controlled by low oxygen tension harbour the Hypoxia Response Element; (HRE) in their regulatory regions. In hypoxia the heterodimeric transcription factor, Hypoxia-Inducible Factor (HIF) binds to this conserved consensus sequence, leading to transcriptional activation of the particular gene.

Normally tissues and cells are not challenged by excess oxygen (hyperoxic stress). Administration of high concentration of pressurised oxygen (hyperbaric oxygen therapy,  $HBO_2$ ) is used as a supportive treatment for patients suffering from severe respiratory failure or chronic ischemic wounds. Despite the benefits of  $HBO_2$  it does have its drawbacks, since prolonged  $HBO_2$  treatment increases tissue damage because of high generation of ROS. Rapid reperfusion of ischemic tissue is extremely critical for restoring normal organ functions. However, a sudden burst of oxygenated blood may produce a progressive destruction of reversibly damaged cells leading to sustained tissue dysfunction. This reperfusion injury has an extremely important role in pathology of the brain during stroke and the heart during myocardial infarction.

## 2. HIF: transcriptional regulator of hypoxic responses

HIF is the most important and best-described factor controlling cellular responses to low oxygen. HIF was originally discovered as being responsible for the regulation of erythropoietin (EPO) expression in hypoxia. Since its discovery, over 100 genes have been identified, which are regulated by HIF and efficiently responding to hypoxia.

Many of the hypoxia-regulated genes play a central roles in acute or chronic adaptation to hypoxia. These genes are implicated in a vast variety of different cellular functions such as cell survival, apoptosis, cell motility, cytoskeletal structure, cell adhesion, angiogenesis, erythropoiesis, vasculature tone, transcriptional regulation, epithelial homeostasis, drug resistance, nucleotide-, amino acid-, iron-, glucose-, energy metabolism and pH regulation.

HIF is ubiquitously expressed and its activity is induced by hypoxia in almost all cell types. The expression of the majority of HIF target genes are induced tightly in a cell-type-specific manner. It is evident that HIF do not alone account for this cell type restricted gene expression. The functional interactions of HIF with other transcription factors determine the subgroup of HIF target genes which is activated in any particular cell type. HIF should be viewed as a messenger that is sent to the nucleus to activate transcriptional responses to hypoxia. The fine details of this response are determined by the developmental and physiological programming of each cell. As a result, the total number of HIF target genes cannot be ascertained by analysis of one or a few cell types. Perhaps 1–4% of all human genes are expressed in response to hypoxia in one or more cell types in a HIF-dependent manner.

HIF is transcriptionally active only as a heterodimeric protein complex composed of two subunits: HIF $\alpha$  and HIF $\beta$  (also known as ARNT which stands for Aryl hydrocarbon Receptor Nuclear Translocator). HIF $\alpha$  members exclusively dimerize with HIF $\beta$  family members, whereas the  $\beta$  subunits can dimerize with various non-HIF transcription factors. Both subunits contain a basic helix-loop-helix (bHLH) domain and two Per-ARNT-Sim (PAS) domains, designated PAS-A and PAS-B. The intact bHLH and PAS domains are required for protein dimerization and DNA binding. There are three HIF $\alpha$  family members HIF-1 $\alpha$ , HIF-2 $\alpha$  and HIF-3 $\alpha$  and three  $\beta$  family members (HIF1 $\beta$ /ARNT1, HIF2 $\beta$ /ARNT2, HIF3 $\beta$ /ARNT3). HIF-1 $\alpha$  and HIF-2 $\alpha$  are expressed in a variety of human cell lines. There is concordance between the HIF-1 $\alpha$  and HIF-2 $\alpha$  responses as well as their regulation. HIF-1 $\alpha$  is the predominant protein in epithelial cells, whereas HIF-2 $\alpha$  is the main form in endothelial and fibroblast cells. Recently it has become evident that HIF-1 $\alpha$  and HIF-2 $\alpha$  have somewhat different and specialized

functions. HIF-3 $\alpha$  has high similarity with HIF-1 $\alpha$  and HIF-2 $\alpha$  in the bHLH and PAS domains. One of the HIF-3 $\alpha$  isoforms (HIF-3 $\alpha$ 2) referred as IPAS lacks structures required for transcriptional activation. Its expression is up-regulated in hypoxia, it dimerizes with HIF $\beta$ /ARNT and binds to the HRE core sequence, but it suppresses hypoxia-inducible HIF-mediated gene expression and appears to be the negative regulator of hypoxia-inducible gene expression. The overall importance of HIF-3 $\alpha$  as a regulator of hypoxia-induced transcription remains to elucidated.

HIF $\alpha$ /HIF $\beta$  dimer binds to consensus DNA sequence (A/G)CGTG (HRE: Hypoxia Response Element) common for almost all genes induced by low oxygen tension. Transcriptional activation and interaction with co-activators such as p300 and CBP are mediated by two transactivation domains in the C-terminal half of HIF $\alpha$ , termed the N-terminal (N-TAD) and C-terminal (C-TAD) transactivation domains. HIF $\beta$  also contains a TAD but it is not required for transcriptional activity.

Both *hif-1a* and *hif-1β* genes are constitutively expressed but only HIF-1β protein is found in cells under all oxygen conditions, whereas HIF-1a is undetectable in normal oxygen conditions. Negative regulation of HIF-1a under normoxic conditions occurs via the oxygen-dependent degradation domain (ODDD). The ODDD domain contains prolyl residues that are hydroxylated in normoxia. This results in the binding of the Von Hippel–Lindau protein (pVHL), which subsequently targets the HIF-1a protein for rapid degradation by the ubiquitin-proteasome system. HIF-2a, as well as some of the splice variants of HIF-3a proteins contains the conserved VHL-binding domain and they are substrates for pVHL and proteasomal degradation (see Figure 1).



Figure 1. Protein domains of HIF $\alpha$  and HIF $\beta$ .

Functional domains are shown. Hydroxylation sites mentioned in the text are indicated. ODDD, oxygen-dependent degradation domain; N-TAD and C-TAD, N- and Cterminal transactivation domain; NLS, nuclear localization signal; PAS, Per-ARNT-Sim.

# 3. Oxygen-dependent regulation of HIF

Being the most crucial regulator of hypoxic responses, the activity of HIF is tightly controlled at multiple levels. The regulation is mostly targeted to the  $\alpha$  subunits and the

PHYSIOLOGY AND MAINTENANCE – Vol. II – Physiological Regulation of Gene Activity by Oxygen  $(O_2)$  - Juha-Pekka Pursiheimo

main regulatory mechanisms involve oxygen-dependent protein stabilization and transcriptional activation. Moreover, activity is further fine-tuned via post-translational modifications, nuclear translocation, dimerization and interactions with other regulatory proteins. In normoxic conditions, when HIF $\alpha$  and HIF $\beta$  are constitutively expressed, the abrogation of HIF activity results mainly from constitutive HIF $\alpha$  degradation.

-

-

-

# TO ACCESS ALL THE **12 PAGES** OF THIS CHAPTER, Visit: http://www.eolss.net/Eolss-sampleAllChapter.aspx

#### Bibliography

Barry R.E. and Krek W. (2004). The von Hippel–Lindau tumour suppressor: a multi-faceted inhibitor of tumourigenesis. *Trends in Molecular Medicine* 10, 466-472. [A very good and well written introduction to the function and the role of VHL in malignant progression.]

Giordano F. (2005). Oxygen, oxidative stress, hypoxia, and heart failure. *The Journal of Clinical Investigation* 115:500-508. [A thorough introduction to the essential role of oxygen in heart physiology.]

Ikeda E. (2005). Cellular response to tissue hypoxia and its involvement in disease progression. *Pathology International*, 55: 603–610. [A good quality paper discussing the hypoxic mechanisms of several disease conditions and showing nice examples of various roles of oxygen in disease progression.]

Khalil A.A., Aziz F.A. and Hall, J.C. (2006). Reperfusion injury. *Plastic and Reconstructive Surgery* 117, 1024-1033. [Discusses the interesting features of hyperoxia and reperfusion in various situations crucial to the success of plastic surgery.]

Lahiri S., Roy A., Baby S., Hoshi T., Semenza G., and Prabhakar N. (2006). Oxygen sensing in the body. Progress in Biophysics and Molecular Biology. 91, 249–286. [A high-class review article discussing thoroughly the various physiological mechanisms participating in oxygen sensing processes in the context of the whole body.]

Lee P. and Choi A. (2003). Pathways of cell signaling in hyperoxia. *Free Radical Biology & Medicine* 35, 341–350. [A detailed discussion of the role and the meaning of excess oxygen to the cell viability.]

Lopez-Neblina, F., Toledo, A., and Toledo-Pereyra, L. (2005). Molecular biology of apoptosis in ischemia and reperfusion. *Journal of Investigative Surgery* 18, 335–350. [A detailed discussion about the apoptotic mechanisms induced by the excess oxygen.]

Michiels C., Minet E., Mottet D., and Raes M. (2002). Regulation of gene expression by ogygen: NF-κB and HIF-1, two extremes. *Free Radical Biology & Medicine*, 3, 1231–1242. [Good paper introducing response mechanisms to both extremes of oxygen-derived stress.]

Schofield C.J., and Ratcliffe, P. (2004). Oxygen sensing by HIF hydroxylases. *Nature Reviews Molecular Cell Biology* 5, 343-354. [The best review article available about the role of hydroxylases in sensing oxygen and guiding the cellular functions in response to fluctuations in oxygen levels.]

Vaupel P. (2004). The role of Hypoxia-Induced Factors in tumor progression. *The Oncologist* 9, 10-17. [Very clear and well written article giving the nice introduction to the central role of HIF in carcinogenesis.]

Willam C., Nicholls L.G., Ratcliffe P.J., Pugh C.W. and Maxwell P.H. (2004). The prolyl hydroxylase enzymes that act as oxygen sensors regulating destruction of hypoxia-inducible factor alpha. *Advances in* 

PHYSIOLOGY AND MAINTENANCE – Vol. II – Physiological Regulation of Gene Activity by Oxygen  $(O_2)$  - Juha-Pekka Pursiheimo

*Enzyme Regulation* 44, 75–92. [A well written and balanced paper about the oxygen sensing hydroxylases, with introduction to the in vivo expression patterns.]

#### **Biographical Sketch**

**Juha-Pekka Pursiheimo,** PhD, was born in 1970 in the town of Pori located in the Western part of Finland. He studied genetics and biochemistry in the University of Turku and received his Master of Science degree in 1995. In 1997 he joined in the Proteoglycan research team of Professor Markku Jalkanen located in the Turku Centre for Biotechnology (TCB) and in 2002 he finished his Doctoral thesis entitled "*Protein Kinase A: Regulator of Growth Factor-Induced Responses*". During the year 2002 he worked as a senior scientist in the drug development company Juvantia Pharma oy. He returned to the Academic research community in 2003 when he joined the Docent Panu Jaakkolas' research group (currently located in TCB) studying the cellular response mechanisms to hypoxia. In 2005 he received the postdoctoral fellowship of Academy of Finland and currently he is continuing the groundbreaking hypoxia research in the Turku Centre for Biotechnology. His special interests in research are the identification of a new and novel hydroxylation targets and the role of PHDs controlling the function of these putative target proteins and hence the vital cellular functions. He has contributed several papers in refereed journals and more are to come in the near future.