MOLECULAR GENETIC IMPROVEMENT OF PROTEIN QUALITY IN MAIZE

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Summary

Increasing the nutritional value of maize grain may contribute to improved human nutrition and health. Although maize is widely grown both for human consumption and for animal feed, maize is an inadequate source of essential amino acids in many diets. Maize protein is deficient in lysine and tryptophan, and in many poultry diets, maize protein is inadequate in the sulfur amino acids (methionine and cysteine). Many attempts have been made to change the amino acid composition of maize grain by changing the relative abundance of different seed proteins in the kernel. Considerable research has focused on the molecular genetic control of kernel protein accumulation. Genetic investigation of the opaque-2 mutant and the modifier genes which restore kernel vitreousness in Quality Protein Maize (QPM) has provided new marker-assisted and transgenic approaches to achieving improved protein quality in maize. Genetic studies of the high-methionine inbred line BSSS53 have identified genomic regions influencing whole-kernel methionine levels. The development of more efficient molecular marker systems will enhance traditional breeding programs aimed at improving the protein quality of elite maize varieties. Genetic engineering strategies to modify amino acid biosynthetic pathways and to over-express high-quality proteins will also yield varieties with improved protein quality.

1. Introduction

Maize (Zea mays L.) is one of the most productive and widely adapted crop species of

the world. Maize is an important staple food crop in Latin America and much of Sub-Saharan Africa. Additionally, maize grain is an important livestock feed in China, Southeast Asia and North America (see *Improving the Nutritional Quality of Maize and Wheat for Human Consumption*). Maize is also widely harvested as silage for dairy production. As maize is highly productive and widely grown, considerable research has focused on improving its nutritional value. Improvement of the nutritional quality of maize grain will increase its value as a food source and will improve the efficiency of animal production. Both of these outcomes can improve human health and nutrition and may have a broad impact, considering the global importance and widespread acceptance of maize.

Although maize grain is a valuable source of dietary energy, it is typically comprised of only 8-10% protein. Furthermore, maize is an inadequate source of essential amino acids in many diets. For monogastric animals, including humans, maize protein is deficient in lysine and tryptophan; and in many poultry diets, maize protein is inadequate in the sulfur amino acids (methionine and cysteine). The amino acid most often lacking in human diets is lysine. In swine and poultry rations balanced for lysine, tryptophan, and the sulfur amino acids, threonine can become limiting. Increasing the levels of lysine, tryptophan, threonine and methionine in maize grain would improve the nutritional value of maize for human nutrition and for livestock feed (see *Plant Based Sources of Proteins and Amino Acids in Relation to Human Health*).

In addition to improving the nutritional value of maize for poultry feed, increasing the methionine content of maize grain may also prove beneficial for reducing micronutrient deficiency in humans. There is increasing evidence that methionine may improve the bioavailability of zinc. Zinc deficiency is a widespread nutritional disorder that can lead to growth failure, pregnancy complications, low birth weight and impaired immune function (see *Global Importance of Zinc Deficiency in Humans: its Relation to Malnutrition and Strategies for its Prevention*). In a rat nutrition study comparing normal maize with conventionally selected high-methionine maize, high-methionine maize had higher levels of bioavailable zinc resulting in elevated dietary zinc intake. Increasing the methionine level of maize grain may alleviate the problems associated with zinc deficiency by increasing the bioavailability of dietary zinc.

Both traditional plant breeding and molecular genetic approaches have been utilized in order to improve the protein quality of maize. For the purposes of this article, traditional plant breeding will be assumed to include both the utilization of naturally occurring genetic variation through controlled crossing of maize lines or strains followed by artificial selection and the use of mutagenic agents to increase genetic variation prior to selection. Molecular genetic approaches will be assumed to include those methods for which molecular sequence information is required during development or implementation. These methods include the development of transgenic plants with novel phenotypic characteristics and the utilization of molecular markers to augment traditional breeding efforts.

The objective of this article is to highlight molecular genetic attempts to improve maize protein quality. To achieve this objective, the results of traditional breeding programs aimed at maize protein quality improvement will be referred to, since improved germplasm arising out of these programs has provided a starting point for molecular genetic efforts in many cases. Several attempts have been made to alter the amino acid composition of the maize kernel by changing the relative abundance of different seed proteins. In order to summarize this research, we first will briefly review the nomenclature of maize kernel proteins and the amino acid profiles of these proteins.

Considerable research has been performed to better understand the metabolic pathways involving lysine, threonine and methionine synthesis and degradation. Lysine, methionine and threonine are all produced via the aspartate biosynthetic pathway. Due to the nutritional importance of these amino acids, considerable effort has focused on understanding and manipulating this critical biosynthetic pathway. Elucidation of amino acid metabolic pathways and the key regulatory enzymes of these pathways have led to the design of molecular genetic approaches for improving the amino acid balance of maize grain.

2. Maize Seed Proteins

The relative proportions of different seed proteins in the kernel influence the nutritional value of maize protein. Maize kernel protein is divided into four major classes based on solubility properties (Table 1). The commonly used Osborne seed protein classification system separates seed storage proteins into albumins, globulins, glutelins and prolamins. The albumin proteins are soluble in water or low-salt aqueous solutions, and the globulins are soluble in high-salt aqueous solutions. Although the albumins and globulins are relatively high in lysine (5.0-7.0%), these two protein fractions combined represent only 6-12% of the total kernel protein. Glutelins are seed proteins soluble in dilute alkali solutions and they contain 3.0-3.4% lysine. Glutelins are the second most abundant class of maize seed proteins accounting for 35-45% of total kernel protein.

Protein class	Solubility properties	Relative abundance	Nutritional value
Albumins	Water or low-salt aqueous solutions	Low	High
Globulins	High-salt aqueous solutions	Low	High
Glutelins	Dilute alkali solutions	High	Intermediate
Prolamines (Zeins)	Alcohol	High	Low*

*Specific maize prolamines are high in methionine, however these tend to be lower in abundance

Table 1. Major classes of maize kernel proteins.

The major class of proteins in the maize kernel is the prolamine fraction. Maize prolamines, referred to as zeins, account for approximately 50-60% of the total kernel

protein. Considerable research has been directed at understanding and manipulating the various zein proteins. Zeins compose a large, heterogeneous group of storage proteins (Table 2).

The commonly used nomenclature system distinguishes four classes: α -, β -, γ -, and δ zeins based on solubility properties and structural similarities. Specific proteins within each class are further characterized by apparent molecular mass following SDS-PAGE. The α -zeins class consists of 19- and 22-kDa proteins and account for the majority of zein protein in common genetic backgrounds. Both the 19- and 22-kDa α -zeins are mixtures of related polypeptides encoded by large gene families. The α -zeins contain no lysine and low levels of methionine. The β -, γ -, and δ -zeins are soluble in alcohol in the presence of a reducing agent. The β -zein is a 14-kDa protein containing no lysine and approximately 11% methionine. The γ -zeins, including 16- and 27-kDa proteins, are also devoid of lysine and are less than 2.0% methionine. The δ -zein class is comprised of a 10-kDa protein encoded by a single structural gene and an 18-kDa protein encoded by the duplicate locus of the 10-kDa zein structural gene. The δ -zeins have a very high methionine content. The 10-kDa δ -zein is 23% methionine and the 18-kDa δ -zein is 27%. Additionally, the 18-kDa δ -zein contains one lysine and two tryptophan codons.

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Zein class	Molecular mass	Lysine and methionine content
α	19-kDa 22-kDa	Low levels of methionine; no lysine
β	14-kDa	Intermediate levels of methionine; no lysine
γ	16-kDa 27-kDa	Low levels of methionine; no lysine
δ	10-kDa 18-kDa	High levels of methionine; the 18-kDa zein structural gene also contains one lysine codon

Table 2. Major classes of zein proteins.

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Bibliography

Anthony, J., W. Brown, D. Buhr, G. Ronhovde, D. Genovesi, T. Lane, R. Yingling, K. Aves, M. Rosato and P. Anderson (1997). Transgenic maize with elevated 10 kD zein and methionine. In Cram, W.J., L.J. De Kok, I. Stulen, C. Brulen and H. Rennenberg (eds.) *Sulphur metabolism in higher plants: molecular, ecophysiological and nutritional aspects.* Backhuys Publishers, Leiden, The Netherlands. pp. 295-297. [Brief research paper discussing the transgenic approach to increasing methionine by overexpressing the 10-kDa δ -zein].

Azevedo, R.A., P. Arruda, W.L. Turner, and P.J. Lea (1997). The biosynthesis and metabolism of the aspartate derived amino acids in higher plants. *Phytochemistry* **46**, 395-419. [Review of aspartate pathway metabolism and transgenic manipulation of key regulatory enzymes].

Benner, M.S., R.L. Phillips, J.A. Kirihara and J.W. Messing (1989). Genetic analysis of methionine-rich storage protein accumulation in maize. *Theor. Appl. Genet.* **78**, 761-767. [Research paper reporting map position of *dzr1* and *dzs10*].

Bjarnason, M. and S.K. Vasal (1992). Breeding of Quality Protein Maize (QPM). In *Plant Breeding Reviews vol. 9.* J. Janick (ed.) Wiley: New York. pp. 181-216. [Comprehensive overview of maize breeding efforts aimed at improving protein quality].

Bryan, J.K. (1990). Advances in the biochemistry of amino acid biosynthesis. In B.J. Miflin and P.J. Lea (eds.) *The Biochemistry of Plants, Vol 16: Intermediary Nitrogen Metabolism.* Academic Press Inc. New York, NY pp. 161-195. [Overview of amino acid biosynthesis in plants].

Chaudhuri, S., J. Messing (1995). RFLP mapping of the maize *dzr1* locus, which regulates methioninerich 10 kDa zein accumulation. *Mol. Gen. Genet.* **246**, 707-715. [Research paper detailing the finemapping of *dzr1* on chromosome 4S].

Chung, K.O. and Y. Pomeranz (1985). Amino acids in cereal proteins and protein fractions. In Finley, J.W. and D.T. Hopkins (eds). *Digestibility and amino acid availability in cereals and oilseeds*. Amer. Assoc. of Cereal Chemists. St. Paul, MN. pp. 65-93. [Review article containing amino acid profiles of several major grains and their specific protein fractions].

Ciceri, P., E. Gianazza, B. Lazzari, G. Lippoli, A. Genga, G. Hoschek, R.J. Schmidt, and A. Viotti (1997). Phosphorylation of *Opaque-2* changes diurnally and impacts its DNA binding activity. *Plant Cell.* 9, 97-108. [Research paper reporting regulation of *Opaque-2* activity via phosphorylation of the *O2* protein].

Coleman, C.E., J.M. Dannenhoffer, and B.A. Larkins (1997). The Prolamin Proteins of Maize, Sorghum and *Coix*. In *Cellular and molecular biology of plant seed development*. B.A. Larkins and I.K. Vasil (eds). pp. 257-288. [Thorough review of zein protein properties and classification].

Cruz-Alvarez, M., J.A. Kirihara and J.W. Messing (1991). Post-transcriptional regulation of methionine content in maize kernels. *Mol. Gen. Genet.* **225**, 331-339. [Research article reporting regulatory control of 10-kDa δ-zein by *dzr1*].

Dannenhoffer, J.M., D.E. Bostwick, E. Or and B.A. Larkins (1995). *opaque-15*, a maize mutation with properties of a defective *opaque-2* modifier. *Proc. Natl. Acad. Sci. USA*. **92**, 1931-1935. [Research paper reporting the characteristics and map position of *opaque-15*].

Das, O.P., S.L. Minzi, M. Koury, M. Benner, J. Messing (1990). Asomatic gene rearrangement contributing to genetic diversity in maize. *Proc. Natl. Acad. Sci. USA*. **87**, 7909-7813. [Discussion of the AB and Ra alleles of the 27-kDa γ -zein structural gene].

Galili, G. (1995). Regulation of lysine and threonine synthesis. *Plant Cell* **7**, 899-906. [Review paper discussing regulation of the aspartate pathway].

Galili, G. and B.A. Larkins (1999). Enhancing the content of the essential amino acids lysine and threonine in plants. In B.K. Singh (ed.) *Plant Amino Acids*. Marcel Dekker. New York. pp. 487-507. [Review of biotechnological and traditional efforts aimed at increasing lysine and threonine in plants].

Gallusci, P., F. Salamini, and R.D. Thompson (1994). Differences in cell type-specific expression of the gene *Opaque-2* in maize and transgenic tobacco. *Mol. Gen. Genet.* **244**, 391-400. [Research paper describing endosperm-specific expression of the *Opaque-2* gene].

Gallusci, P., S. Varotto, M. Matsuoko, M. Maddaloni, and R. D. Thompson (1996). Regulation of cytosolic pyruvate, orthophosphate dikinease expression in developing maize endosperm. *Plant Mol. Biol.* **31**, 45-55. [Research paper reporting the effect of *Opaque-2* on the orthophosphate dikinase gene].

Geetha, K.B., C. R. Lending, M.A. Lopes, J.C. Wallace, and B.A. Larkins (1991). *opaque-2* modifiers increase gamma-zein synthesis and alter its spatial distribution in maize endosperm. *Plant Cell* **3**, 1201-1219. [Research paper reporting elevated levels of the 27-kDa γ -zein in QPM endosperm].

Gevers, H.O. and J. K. Lake (1992). Development of modified *opaque-2* maize in South Africa. In *Quality Protein Maize*. E. T. Mertz (ed). American Association of Cereal Chemists, St. Paul, MN. pp. 49-78. [Review of the development of QPM varieties in South Africa].

Gibson, R.S. (1994). Zinc Nutrition in Developing Countries. Paper prepared for presentation at the workshop "Food Based Strategies to Combat Micronutrient Malnutrition in Developing Countries." Washington, D.C. August 9, 1994. [Discussion of zinc deficiency symptoms in humans].

Graham, R., D. Senadhira, S. Beebe, C. Iglesias, and I. Monasterio (1999). Breeding for micronutrient density in edible portions of staple food crops: conventional approaches. *Field Crops Res.* **60**, 57-80. [Discussion of the rationale for breeding for increased levels of micronutrients with reference to the relationship between methionine and zinc bioavailability].

Habben, J.E., A.W. Kirleis and B.A. Larkins. 1993. The origin of lysine-containing proteins in *opaque-2* maize endosperm. *Plant Mol. Bio.* **23**, 825-838. [Research paper reporting elevated levels of Elongation Factor 1A in *opaque-2* mutant kernels].

Habben, J.E., G.L. Moro, B.G. Hunter, B.R. Hamaker and B.A. Larkins (1995). Elongation factor 1α concentration is highly correlated with the lysine content of maize endosperm. *Proc. Natl. Acad. Sci. USA* **92**, 8640-8644. [Research paper reporting correlation between level of Elongation Factor 1A and endosperm lysine content].

House, W.A., D.R. Van Campen, and R.M. Welch (1997). Dietary methionine status and its relation to the bioavailavity to rats of zinc in corn kernels with varying methionine content. *Nutr. Res.* **17**, 65-76. [Research paper reporting the association of increased methionine in corn grain used for feed with improved dietary zinc intake of rats].

Kata, S.R., B.H. Taylor, A.J. Bockholt, and J.D. Smith (1994). Identification of *opaque-2* genotypes in segregating populations of Quality Protein Maize by analysis of restriction fragment length polymorphism. *Theor. Appl. Genet.* **89**, 407-412. [Research paper describing the use of an RFLP probe from the *Opaque-2* gene to aid selection during backcrossing].

Kirihara, J.A., J.B. Petri and J.W. Messing (1988a). Isolation and sequence of a gene encoding a methionine-rich 10-kDa zein protein from maize. *Gene* **71**, 359-370. [Research paper reporting the cloning of *dzs10*].

Kleese, R.A., J.A. Kirihara and G.A. Sandahl (1991). Gene transfer to elevate methionine levels. Proc. 46th Annual Corn and Soybean Ind. Res. Conf. Amer. Seed Trade Assoc. Washington D.C. pp.124-129. [Research paper reporting attempts to overexpress the 10 kDa δ-zein].

Krebbers, E., R. Broglie, B. Hitz, T. Jones, and N. Hubbard (1997). Biotechnological approaches to altering seed composition. In B.A. Larkins and I.K. Vasil (eds.). *Cellular and Molecular Biology of Plant Seed Development*. Kluwer Academic Publishers. Netherlands. pp. 595-633. [Review of biotechnological efforts at seed quality modification including a section on improvement of amino acid profiles].

Krone, T.L. (1994). *Genetic analysis and breeding for kernel methionine content in maize (Zea mays* L.). University of Minnesota, Twin Cities. Ph.D. Thesis. [Thesis describing phenotypic selection for whole kernel methionine content and reporting the control of 10 kDa δ-zein levels by a region of chromosome 7S].

Lambert, R.J. and Chung, L.C. (1995). Phenotypic recurrent selection for increased endosperm hardness in two high-lysine maize synthetics. *Crop Sci.* **35**, 451-456. [Discussion of the development of QPM varieties at the University of Illinois].

Larkins, B.A., K. Pedersen, M.D. Marks, and D.R. Wilson (1984). The zein proteins of maize endosperm. *Trends Biochem. Sci.* **9**, 306-308. [Review of zein properties and nomenclature].

IMPACTS OF AGRICULTURE ON HUMAN HEALTH AND NUTRITION – Vol. II - Molecular Genetic Improvement of Protein Quality in Maize - Olsen, M.S. and Phillips, R.L.

Lin, K.R., A.J. Bockholt, C.W. Magill, and J.D. Smith (1997). Changes in soluble endosperm proteins associated with selection of quality protein maize lines. *Maydica*. **42**, 355-362. [Research paper comparing seed protein levels of wild-type, *o2* and QPM inbred lines].

Lohmer, S., M. Maddaloni, M. Motto, F. Salamini, and R. D. Thompson (1993). Translation of the mRNA of the maize transcriptional activator *Opaque-2* is inhibited by upstream open reading frames present in the leader sequence. *Plant Cell* **5**, 65:73. [Research paper describing regulation of *Opaque-2* expression].

Lopes, M.A. and B.A. Larkins (1991). Gamma-zein content is related to endosperm modification in Quality Protein Maize. *Crop Sci.* **31**, 1655-1662. [Research paper reporting elevated 27-kDa γ -zein levels in QPM and correlation between 27-kDa γ -zein level and kernel vitreousness].

Lopes, M.A. and B.A. Larkins (1995). Genetic analysis of *opaque-2* modifier gene activity in maize endosperm. *Theor. Appl. Genet* **19**, 274-281. [Research paper reporting the probable presence of two *opaque-2* modifier genes in QPM].

Lopes, M.A., K. Takasaki, D.E. Bostwick, T. Helentjaris and B.A. Larkins (1995). Identification of two *opaque-2* modifier loci in Quality Protein Maize. *Mol. Gen. Genet.* **247**, 603-613. [Research paper reporting map positions of two *opaque-2* modifier genes].

Maddaloni, M., G. Donini, C. Balconi, E. Rizzi, P. Gallusci, F. Forlani, S. Lohmer, R. Thompson, F. Salamini, M. Motto (1996). The transcriptional activator *Opaque-2* controls the expression of a cytosolic form of pyruvate orthophosphate dikinase-1 in maize endosperms. *Mol. Gen. Genet.* **250**, 647-654. [Research paper describing effect of *Opaque-2* on pyruvate orthophosphate dikinase].

Manzur, B., E. Krebbers, and S. Tingey (1999). Gene discovery and product development for grain quality traits. *Science* **285**, 372-375. [Review of genomics efforts aimed at developing improved grain quality traits].

Matthews, B.F. (1999). Lysine, threonine, and methionine biosynthesis. In B.K. Singh (ed.) *Plant Amino Acids*. Marcel Dekker. New York. pp. 205-225. [Review of the aspartate pathway].

Mertz, E.T., L.S. Bates, and O.E. Nelson (1964). Mutant gene that changes protein composition and increases lysine content of maize endosperm. *Science* **145**, 279-280. [Research paper reporting the discovery of increased lysine content caused by the *opaque-2* mutant allele].

Moro, G.L., M.A. Lopes, J.E. Habben, B.R. Hamaker, and B.A. Larkins. (1995).Phenotypic effects of *opaque-2* modifier genes in normal maize endosperm. *Cereal Chem.* **72**, 94-99. [Research paper reporting effects of *opaque-2* modifier genes in wild-type (non *o2*) maize endosperm].

Moro, G.L., J.E. Habben, B.R. Hamaker, and B.A. Larkins (1996). Characterization of the variability in lysine content for normal and *opaque-2* maize endosperm. *Crop Sci.* **36**, 1651-1659. [Research paper reporting the endosperm lysine content of several normal and *opaque-2* maize lines].

Motto, M., M. Maddaloni, G. Ponziani, M. Brembilla, R. Marotta, N. Di Fonzo, C. Soave, R. Thompson, and F. Salamini (1988). Molecular cloning of the *o2-m5* allele of *Zea mays* using transposon marking. *Mol. Gen. Genet.* **212**, 488-494. [Research paper reporting the cloning of the *opaque-2* allele using an *Ac* transposable element].

Motto, M., H. Hartings, M. Maddaloni, S. Lohmer, F. Salamini, and R. Thompson (1996). Genetic manipulations of protein quality in maize grain. *Field Crops Research* **45**, 37-48. [Review paper containing a summary of maize seed protein nomenclature and properties].

Muenchrath, D.A., and R.L. Phillips (1993). Relationship of maize seedling response to lysine-plusthreonine medium and whole kernel amino acid profile. *Crop Sci.* **33**, 1095-1099. [Research paper describing the correlation between whole kernel amino acid profile and root growth on media supplemented with lysine and threonine].

Muth, J.R., M. Muller, S. Lohmer, F. Salamini, and R.D. Thompson (1996). The role of multiple binding sites in the activation of zein gene expression by *Opaque-2. Mol. Gen. Genet.* **252**, 723-732. [Research paper describing the *Opaque-2* binding site within the 22-kDa α -zein promoter].

National Research Council (1988). *Quality-Protein Maize*. National Academy Press. Washington, D. C. [General overview of breeding efforts and applications of QPM].

Neto, G.C., J.A. Yunes, M.J. da Silva, A.L. Vettore, P. Arruda, and A. Leite (1995). The involvement of *Opaque-2* on β -prolamin gene regulation in maize and *Coix* suggests a more general role for this transcriptional activator. *Plant Mol. Bio.* **27**, 1015-1029. [Research paper describing regulation of the 14-kDa β -zein and 17-kDa β -coixins by *Opaque-2*].

Olsen, M.S. (1999). *Genetic analysis of whole-kernel methionine in maize*. University of Minnesota, Twin Cities. Ph.D. Thesis. [Thesis reporting results of phenotypic selection for increased whole-kernel methionine and genetic mapping of QTL controlling whole-kernel methionine levels].

Or, E., S.K. Boyer, and B.A. Larkins (1993). *opaque-2* modifiers act post-transcriptionally and in a polar manner on γ -zein gene expression in maize endosperm. *Plant Cell* **5**, 1599-1609. [Research paper describing influence of *opaque-2* modifiers on 27-kDa γ -zein transcript level].

Phillips, R.L., P.R. Morris, F. Wold, and B.G. Gengenbach (1981). Seedling screening for lysine-plusthreonine resistant maize. *Crop Sci.* **21**, 601-606. [Research paper reporting the identification of the line BSSS53 as having elevated whole-kernel methionine content].

Phillips, R.L., and B.A. McClure (1985). Elevated protein-bound methionine in seeds of a maize line resistant to lysine plus threonine. *Cereal Chem.* **62**, 213-218. [Research paper reporting the association of the 10-kDa δ -zein with the high methionine phenotype of the inbred line BSSS53].

Puckett, J.L. and A.L. Kriz (1991). Globulin gene expression in *opaque-2* and *floury-2* mutant embryos. *Maydica.* **36**, 161-167. [Research paper reporting an increase in globulin proteins in embryos of homozygous recessive *opaque-2* kernels].

Pysh, L.D., M.J. Auckerman, and R.J. Schmidt (1993). OHP1: a maize basic domain-leucine zipper protein that interacts with *opaque-2*. *Plant Cell* **5**, 227-236. [Research paper reporting the interaction of *opaque-2* with the basic domain-leucine zipper protein OHP1].

Ravanel, S., B. Gakiere, D. Job, and R. Douce (1998). The specific features of methionine biosynthesis and metabolism in plants. *Proc. Natl. Acad. Sci. USA* **95**, 7805-7812. [Review of methionine biosynthesis and metabolism].

Rossi, V., M. Motto, and L. Pellegrini (1997). Analysis of the methylation pattern of the maize *Opaque-2* (*O2*) promoter and *in vitro* binding studies indicate that the *O2* b-Zip protein and other endosperm factors can bind to methylated target sequences. *J. Biol. Chem.* **272**, 13758-13765. [Research paper reporting auto-regulation of the *Opaque-2* gene and the methylation status of the gene in varying maize tissues].

Schmidt, R.J., F.A. Burr, and B. Burr (1987). Transposon tagging and molecular analysis of the maize regulatory locus *opaque-2*. *Science* **238**, 960-963. [Research paper reporting the cloning of the *opaque-2* gene using an *Spm* transposable element].

Schmidt, R.J., F.A. Burr, M.J. Aukerman, and B. Burr (1990). Maize regulatory gene *opaque-2* encodes a protein with a "leucine-zipper" motif that binds to zein DNA. *Proc. Natl. Acad. Sci. USA* **87**, 46-50. [Research paper describing the protein encoded by the *opaque-2* gene].

Schmidt, R.J., M. Ketudat, M.J. Aukerman, and G. Hoschek (1992). *Opaque-2* is a transcriptional activator that recognizes a specific target site in 22-kDa zein genes. *Plant Cell* **4**, 689-700. [Research paper describing the *Opaque-2* binding site of the 22-kDa α -zein gene].

Sun, Y., N. Carneiro, A.M. Clore, G. L. Moro, J. E. Habben, and B.A. Larkins (1997). Characterization of Elongation Factor 1A and its relationship to protein quality in the endosperm. *Plant Physiol.* **115**, 1101-1107. [Research paper proposing a stoichiometric realtionship between Elongation Factor 1A and other non-zein high-lysine proteins].

Swarup, S., M.C. Timmermans, S. Chaudhuri, and J. Messing (1995). Determinants of the highmethionine trait in wild and exotic germplasm may have escaped selection during early cultivation of maize. *The Plant J.* **8**, 359-368. [Research paper describing the 18-kDa δ -zein protein encoded by the duplicate locus of the 10-kDa zein structural gene].

Unger, E., R.L. Parsons, R.J. Schmidt, B. Bowen, and B.A. Roth (1993). Dominant negative mutants of *Opaque-2* suppress transactivation of a 22-kD zein promoter by *Opaque-2* in maize endosperm cells. *Plant Cell.* **5**, 831-841. [Research paper describing efforts to create a dominant transgenic allele of *Opaque-2*].

Vicente-Carbajosa, J., S.P. Moose, R.L. Parsons, and R.J. Schmidt (1997). A maize zinc-finger protein binds the prolamin box in zein gene promoters and interacts with the basic leucine zipper transcroptional activator *Opaque-2. Proc. Natl. Acad. Sci. USA.* **94**, 7685-7690. [Research paper describing interaction of the *Opaque-2* protein and a prolamine box binding factor].

Villegas, E., S.K. Vasal, and M. Bjarnason. (1992). Quality Protein Maize – what is it and how was it developed. In *Quality Protein Maize*. E. T. Mertz (ed). American Association of Cereal Chemists, St. Paul, MN. pp 27-48. [Review of the development of QPM].

Wilson, C.M. (1983). Seed protein fractions of maize, sorghum, and related cereals. In *Seed Proteins*. pp 271-307. W. Gottschalk and H.P. Muller (eds.). Martinus Nijhoff, Dr. W. Hunk Pub., The Hague. [Review of maize seed protein properties and nomenclature].

Young V.R., and P.L. Pellett. (1990). Current concepts concerning indispensable amino acid needs in adults and their implications for international nutritional planning. *Food and Nutrition Bulletin* **12**, 289-300. The United Nations University. [Discussion of amino acid requirements in humans].

Biographical Sketches



Michael S. Olsen is a corn breeder for Monsanto Company. His current research effort is aimed at developing maize hybrids with enhanced grain quality traits for animal nutrition. He completed a B.A. degree from Bethel College in St. Paul, Minnesota before earning the M.S. and Ph.D. degrees from the University of Minnesota. His dissertation focused on the genetics of whole-kernel methionine in maize. Prior to joining Monsanto, he worked for three years as a specialty traits corn breeder for Wilson Genetics, L.L.C., a joint venture between Syngenta Seeds and Land O' Lakes. His research focus at Wilson Genetics was the development of maize hybrids with enhanced grain quality traits.

Ronald L. Phillips is Regents' Professor, University of Minnesota. He earned the B.S. and M.S. degrees from Purdue University and a Ph.D. from the University of Minnesota; his postdoctoral training was at Cornell University. Throughout his career, Dr. Phillips has coupled the techniques of classical cytogenetics with research advances in tissue culture and molecular biology to enhance our understanding of basic biology of cereal crops and to improve these species by innovative methods. His research program at the University of Minnesota was one of the early programs in modern plant biotechnology related to agriculture. He is a founding member and former Director of the Plant Molecular Genetics Institute of the University of Minnesota. He has served on numerous editorial boards, has edited six books, and has published over 50 chapters, 100 refereed journal articles, and 200 abstracts. Dr. Phillips teaches a world renowned course in cytogenetics and is invited to teach the course or present the results of his research at numerous university, governmental, and industrial institutions in the U.S. and abroad. He served as Chief Scientist of the USDA (1996-1998) in charge of the National Research Initiative Competitive Grants Program. Awards include Fellow of ASA, CSSA, and AAAS, the Purdue University Agriculture Distinguished Alumni Award, an Honorary Degree from Purdue University, and the Dekalb Genetics Crop Science Distinguished Career Award. In 1991, he was elected a member of the National Academy of Sciences. Dr. Phillips has also served as the President of the Crop Science Society of America.