

KINGDOM FUNGI

Merje Toome

*Department of Plant Pathology and Crop Physiology, Louisiana State University
Agricultural Center, Baton Rouge, Louisiana 70803, U.S.A.*

*Institute of Agricultural and Environmental Sciences, Estonian University of Life
Sciences, Tartu 51014, Estonia*

M. Catherine Aime

*Department of Plant Pathology and Crop Physiology, Louisiana State University
Agricultural Center, Baton Rouge, Louisiana 70803, U.S.A.*

Keywords: Ascomycota, Basidiomycota, chytrid, endophytes, fungal diversity, fungal genomics, fungal parasites, fungal systematics, Glomeromycota, lichen, Microsporidia, mushroom, mycorrhiza, saprobe, use of fungi, zygomycetes

Contents

1. Introduction
 - 1.1. Importance of Fungi
 - 1.1.1. Human Use of Fungi
 - 1.1.2. Role in Nature
 2. Fungal Biology
 - 2.1. Somatic Growth
 - 2.2. Asexual Reproduction
 - 2.3. Sexual Reproduction
 - 2.4. Fungal Metabolites
 3. Fungal Ecology
 - 3.1. Saprobic Fungi
 - 3.2. Parasitic Fungi
 - 3.2.1. Plant Parasites
 - 3.2.2. Animal Parasites
 - 3.2.3. Parasites of Other Fungi
 - 3.3. Mutualistic Fungi
 - 3.3.1. Mycorrhiza
 - 3.3.2. Endophytes
 - 3.3.3. Lichens
 4. Fungal Taxonomy and Systematics
 5. Fungal Genomics
 6. Introduction to Major Fungal Groups
 - 6.1. Chytrids
 - 6.1.1. Neocallimastigomycota
 - 6.1.2. Chytridiomycota
 - 6.1.3. Blastocladiomycota
 - 6.2. Zygomycetes
 - 6.2.1. Mucoromycotina
 - 6.2.2. Kickxellomycotina + Zoopagomycotina + Entomophthoromycotina
 - 6.3. Microsporidia

- 6.4. Glomeromycota
- 6.5. Ascomycota
 - 6.5.1. Taphrinomycotina
 - 6.5.2. Saccharomycotina
 - 6.5.3. Pezizomycotina
- 6.6. Basidiomycota
 - 6.6.1. Pucciniomycotina
 - 6.6.2. Ustilaginomycotina
 - 6.6.3. Agaricomycotina
- 7. Future Outlook
- Glossary
- Bibliography
- Biographical Sketches

Summary

Kingdom Fungi contains a diverse group of organisms. The common characters among all true Fungi are the presence of chitin in the cell wall and in most species, the presence of zygotic meiosis (meiosis that occurs in the zygote). The best-known fungi include mushrooms, molds and yeasts. However, fungal growth exists in an incredible range of sizes, shapes and colors. Additionally, fungi have adapted to diverse ecological niches that have enabled these organisms to thrive in almost every habitat on the planet. The majority of fungi are saprobes, facilitating the break-down of organic matter and enabling the recycling of carbon and other nutrients essential for the development and growth of all known living organisms.

However, other species are parasitic on plants, animals or other fungi, or they form various symbiotic relationships with plants and animals. Due to their microscopic size, fungi are often overlooked and their role in nature and human life underestimated. Most estimates of fungal diversity indicate that there are at least 1.4 million species still to be discovered; therefore the systematics and classification of fungi will continue to change as scientists continue the important work of documenting fungal diversity. Today with the aid of molecular data, the main taxonomic units of Fungi are better resolved than ever before, and are currently divided into eight phyla.

The largest and the most morphologically variable phyla are Ascomycota and Basidiomycota, which form the crown radiation of Fungi known as the Dikarya. Other phyla include arbuscular mycorrhizal fungi (Glomeromycota); chytrids (Blastocladiomycota, Chytridiomycota, Neocallimastigomycota), which possess posteriorly inserted whiplash flagella; and a potentially polyphyletic group, Zygomycota. Another group, Microsporidia, a group of intracellular animal pathogens, has also been shown to belong to Fungi, but its position in relation to other members of the kingdom has yet to be resolved.

1. Introduction

There are more than 110000 species known today in the kingdom Fungi. Taking into consideration their “celebrity,” we may say that the “king” could be the well-known

button mushroom and the “queen” may be the chanterelle. However, every proper kingdom needs to have a large enough court and enough citizens for it to function properly. Various predictions have been made for the number of species in the fungal kingdom and at least 1.4 million but perhaps as many as five million fungal species remain undescribed. Thus, by any estimate less than 10% of fungal species have been discovered and described by science, and an even smaller percentage of these have been studied extensively. Thus, there is a vast need for both additional research and greater numbers of scientists trained to study these organisms.

The branch of biology devoted to the study of fungi is known as mycology, derived from the Greek word for “mushroom” – *Mykes*. Although the number of mycologists and fungal biologists today is greater than ever, the majority of these specialize on a small number of taxa, often “model fungi,” leaving the majority of fungal groups understudied.

The best known fungi are probably the mushrooms that decorate parks and forest floors or our dinner plates. But mushroom-forming species are only a small part of the fungal kingdom, and most species are actually microscopic, growing for instance inside the plants in the garden or on the hair of the cat sitting on the street corner. Fungi surround us all on a daily basis and directly facilitate life on Earth by recycling important nutrients, serving as a food source for various organisms and facilitating the growth of plants. Of course, not all fungi are beneficial and many of them cause diseases in plants and animals and produce harmful toxins. Therefore, mycological research is very important for discovering new ways to benefit from fungi as well as to protect our food and health from fungal disease.

In this chapter we provide an overview of the general characteristics of fungi and their importance, review the current state of fungal systematics and genomics, and, finally, discuss each of the major groups of fungi. While classifications for these organisms have, and will continue to change, the importance of the members of this kingdom to humans and their impact on the world around us will not.

1.1. Importance of Fungi

1.1.1. Human Use of Fungi

Humans have used fungi on a daily basis since recorded time and beyond. Mushrooms and other fungal fruiting bodies have been used as a food source, in medicine, or as an aid in contacting spirits. For instance, in 1991 the well-preserved body of a bronze-age man, nicknamed the Iceman, was discovered in the Alps. Also preserved were his clothing, gear, and two species of fungi that he carried with him. One of these, *Piptoporus betulinus*, is believed to have been used for medical purposes, while the other, *Fomes fomentarius*, was used as fire tinder. The fact that these fungi were carefully tended and stored by the Iceman shows that they must have served an important role in the life of humans more than 5000 years ago. In fact fungi are still used in folk medicine in the Alps region today. Even earlier uses of fungi include the production of wine, beer and bread by ancient Egyptians and other cultures. The fermentative processes needed for the production of these foods are facilitated by

microscopic fungi that were skillfully, although unknowingly, cultured at least 8000 years ago. Ancient writings contain descriptions of various fungal crop diseases that have plagued man since the beginning of agriculture, and gods, such as Robigus, the Roman god of rust and mildew, were invoked for protection against devastating epidemics.

In many ways, the use of fungi by modern man is similar to that in ancient times. The main use is in food production, where various foods are produced through fermentation (e.g. bread, alcoholic beverages, soy sauce, miso). Several edible mushrooms including the button mushroom (*Agaricus bisporus*) and shiitake (*Lentinula edodes*) are grown commercially and used on a daily basis, especially in Asian cuisine. Mycophagy of wild mushrooms, however, is probably practiced by fewer cultures than in the past, perhaps due to less knowledge about fungal species and their edibility by modern people. Fungi are also used in industry to produce various biochemical compounds, including antibiotics and other medically important products.

Some fungi are able to break down toxic compounds such as diesel fuel and phenol, and they contribute to bioremediation efforts to treat toxic waste or purify contaminated soils. Fungi can also serve as bio-indicators to detect changes in the environment. For example, some lichen species are especially sensitive to air pollution, while others can grow well in urban environments even with rather high pollution rates. Monitoring the occurrence of these lichens therefore provides information about the air quality. Additionally, fungi are utilized as natural fertilizers and as biological control agents to provide environmentally friendly pest or weed control in modern agricultural systems. Increasing issues with the use of artificial and chemically produced products and awareness about the need for sustainable systems will most likely result in the discovery of new uses of fungi.

1.1.2. Role in Nature

It is hard to over-emphasize the importance of fungi in the ecosystem functioning of the Earth. Fossils and molecular evidence indicate that fungi accompanied the first photosynthesizing organisms on land to help them cope with the harsh environment they met. Fungi were the first organisms to break down plant and other organic material and therefore facilitate the formation of soils.

Moreover, it has been speculated that fungi have greatly contributed to animal evolution by binding carbon and raising the atmospheric oxygen content, which enabled the development of large eukaryotic organisms in the oceans and led to the evolution of all animals we know today.

Even the extinction of dinosaurs and the subsequent rise of mammals have been proposed to have resulted from fungal infections that killed most of the cold-blooded animals. In this possible scenario, mammals survived because fungi were not able to grow in their higher body temperatures. Of monumental but often overlooked importance is the role that fungi play as the major decomposers of plant and animal material on the planet. Were the saprobic action of fungi to cease, the planet would

become covered with debris, and nutrient recycling would be too slow for most of the life forms we know today to flourish or survive.

In modern ecosystems fungi play an important role in almost every nutrient cycle, either as sources of food, decomposers of organic materials, or essential symbionts of plants and animals, providing necessary nutrients or protecting their hosts. Although fungi are thought to be primarily terrestrial, recent studies have emphasized that a diversity of fungi are found in freshwater, seas and ocean bottoms, ranging from tropical to Arctic and Antarctic regions. Although the precise roles of fungi in these ecosystems are still poorly understood, it is clear that they are essential.

Therefore, studies of fungal diversity and conservation are important because as is the case with other organisms, fungi are becoming extinct due to the changes in the environment and as a result of human activities. Several countries have taken steps to protect fungi as is also done for animals, insects and plants, but only public awareness will make a real difference in the conservation and preservation of these organisms that helped to shape this planet and its ecosystems long before the advent of humans.

2. Fungal Biology

2.1. Somatic Growth

Fungi are eukaryotic (e.g., contain a nucleus) organisms, most of which grow as yeasts or as characteristic elongated cells called hyphae (Figure 1a, b). Yeasts are a unicellular growth form with a spherical to ovoid shape. They may be either colorless or pigmented and primarily grow in nutrient-rich and humid environments. Yeasts occur among zygomycetes (e.g., *Mucor*), ascomycetes (e.g. *Saccharomyces* spp.) and basidiomycetes (e.g. *Rhodotorula* spp.). Some yeasts, including several vertebrate pathogens (e.g., *Blastomyces*), also form hyphae at some stage of their growth.

Hyphae are long filamentous cells that are surrounded by a rigid cell wall. Fungal thalli (the fungal “body”) and a great variety of fruiting structures (e.g., mushrooms) are all produced by hyphae, which, when aggregated into groups are known collectively as mycelium (Figure 1c, l, m). Fungal cell walls contain chitin (more complex forms of chitin are also known from arthropods), glucans and mannose-containing glycoproteins. Hyphae can be septate, meaning that they are divided into numerous compartments or cells by internal cross-walls called septa (Fig 1a), or non-septate wherein the hyphae are multinucleate. The septa of most fungi have central pores that allow cytoplasm and organelles, sometimes including nuclei, to move from one cell to another, securing the transport of nutrients and necessary cellular components.

The structure of these septal pores is often an important character used to differentiate between fungal phyla and subphyla. An important characteristic of hyphae is that they grow apically in one direction only and often branch during growth. The rigid cell walls enable the build-up of turgor pressure inside the hyphae, allowing fungi to penetrate very dense tissues like plant epidermis or insect cuticles. The majority of fungi have no motile organs so they reach their food source by hyphal growth. Hyphal growth is also one method of dispersal and some fungi (e.g. *Armillaria ostoyae*) have reached several

kilometers from their point of origin by spreading mostly in the soil as mycelium. Hyphae of some fungi may produce very dense structures called sclerotia (Figure 1c) that serve in overwintering as well as dissemination, showing that hyphae may serve various functions in the life cycle of fungi.

2.2. Asexual Reproduction

Most fungi produce asexual structures (Table 1). This reproductive strategy does not usually contribute to genetic diversity since all the spores produced are the result of mitosis (rather than meiosis, as in sexual spores) and are therefore genetically identical. However, asexual reproduction is advantageous in allowing the production of tremendous numbers of mitospores (spores produced by mitosis) that can be the main units of dissemination for a majority of species. There is an amazing diversity in form and mode of production of these spores in fungi.

The most commonly produced mitospores are called conidia (produced mainly by ascomycetes and some basidiomycetes), sporangiospores (produced by zygosporic fungi; Figure 1f), and zoospores (flagellated spores of chytrids; Figure 1e). Conidia and sporangiospores are usually formed on specialized hyphae called conidiophores or sporangiophores, respectively, that may be uni- or multicellular and vary greatly in size, shape and color. Asexually produced spores tend to be lighter and smaller than sexually produced spores and are therefore easily dispersed by wind, water or animals.

Fungi have developed many adaptations for mitospore dispersal in specialized niches such as the production of appendices and air pockets that help spores of non-motile aquatic species to ride on currents or float on foam; and the production of spores within a sticky sugary matrix, which attracts insects for dispersal. In addition, the flagellated zoospores of chytrids allow these propagules to swim to new substrates. Although very commonly found, not all fungi have known asexual stages, and especially in Agaricomycotina, reliance on this type of reproduction has been greatly reduced.

2.3. Sexual Reproduction

Sexual reproduction in fungi tends to be more infrequent than asexual reproduction and there is some evidence that, while long distance dispersal can occur, sexually produced spores are not as well adapted for this as asexually produced spores. Rather, meiospores (spores formed by meiosis) are increasing the genetic diversity of the populations by sexual recombination, and often are adapted for surviving harsh conditions (such as overwintering). One key feature that distinguishes Fungi from plants and animals is that the diploid condition in most species of Fungi is fleeting and often immediately followed by meiosis.

Thus, fungi are said to have zygotic meiosis. Sexual reproduction is typically initiated when haploid hyphae (or in some cases, haploid spores) from two compatible mating types fuse. What happens next varies greatly by fungal group, with the chytrids being especially variable, and will be discussed in more detail under the relevant sections in the following. In general, at some stage after plasmogamy, karyogamy and meiosis will occur within a specialized cell or structure that in ascomycetes is called an ascus

(Figure 1g), in basidiomycetes, a basidium (Figure 1h), and in zygomycetes, a zygosporangium. Meiospores are formed on or within these cells and are termed ascospores, basidiospores, and zygospores, respectively (Table 1).

Sexual reproduction in the chytrids is often, but not always, accomplished by the production of a resting sporangium (or resting spore) within which meiosis occurs. Not all fungi have known sexual reproductive stages.

In some cases, this stage may simply be rare and undiscovered, but in other cases the fungi may have lost their ability to reproduce sexually and are able to inhabit their niches successfully without the need for sexual recombination. Some fungi, especially within the Dikarya, produce elaborate fruiting bodies during this stage of development.

2.4. Metabolites

In addition to primary metabolites many fungi produce low-molecular-weight, often biologically active compounds known as secondary metabolites. Although these compounds may be chemically diverse, they are usually produced via common biosynthetic pathways, often related to morphological development.

Some secondary metabolites may have potent physiological activities that provide fungi with fitness benefits in nature, for instance, for competing with other microorganisms, for protection from consumption, for attracting a mate, or by facilitating their distribution by attracting vectors.

One example of vector attraction involves some gasteroid fungi (e.g. truffles) that form their fruiting bodies underground and have therefore no traditional means for spore dispersal.

However, their secondary metabolites are detected by various animals like pigs and squirrels, which dig out the fruiting bodies, eat them, and via their digestive system transport the fungal spores to new locations.

The best-known and most studied fungal metabolites are various toxins that harm vertebrates, including humans. For example the toxins of some *Amanita* species are lethal to humans within days after consumption, whereas other toxins, like those produced by *Aspergillus flavus* and ergot alkaloids produced by *Claviceps purpurea*, cause effects in humans after a longer period of consumption, sometimes months or even years.

The metabolites of other fungi such as certain species of *Psilocybe* may cause hallucinations when consumed and have been used not only recreationally but also for spiritual experiences. Other fungi have been shown to produce beneficial compounds such as antibiotics (e.g. *Penicillium* spp.), inhibitors of virus reproduction, and inhibitors of cancer cell growth (e.g. metabolites of *Ganoderma lucidum*). At present, numerous studies are aimed at uncovering new fungal secondary compounds of benefit to humans.

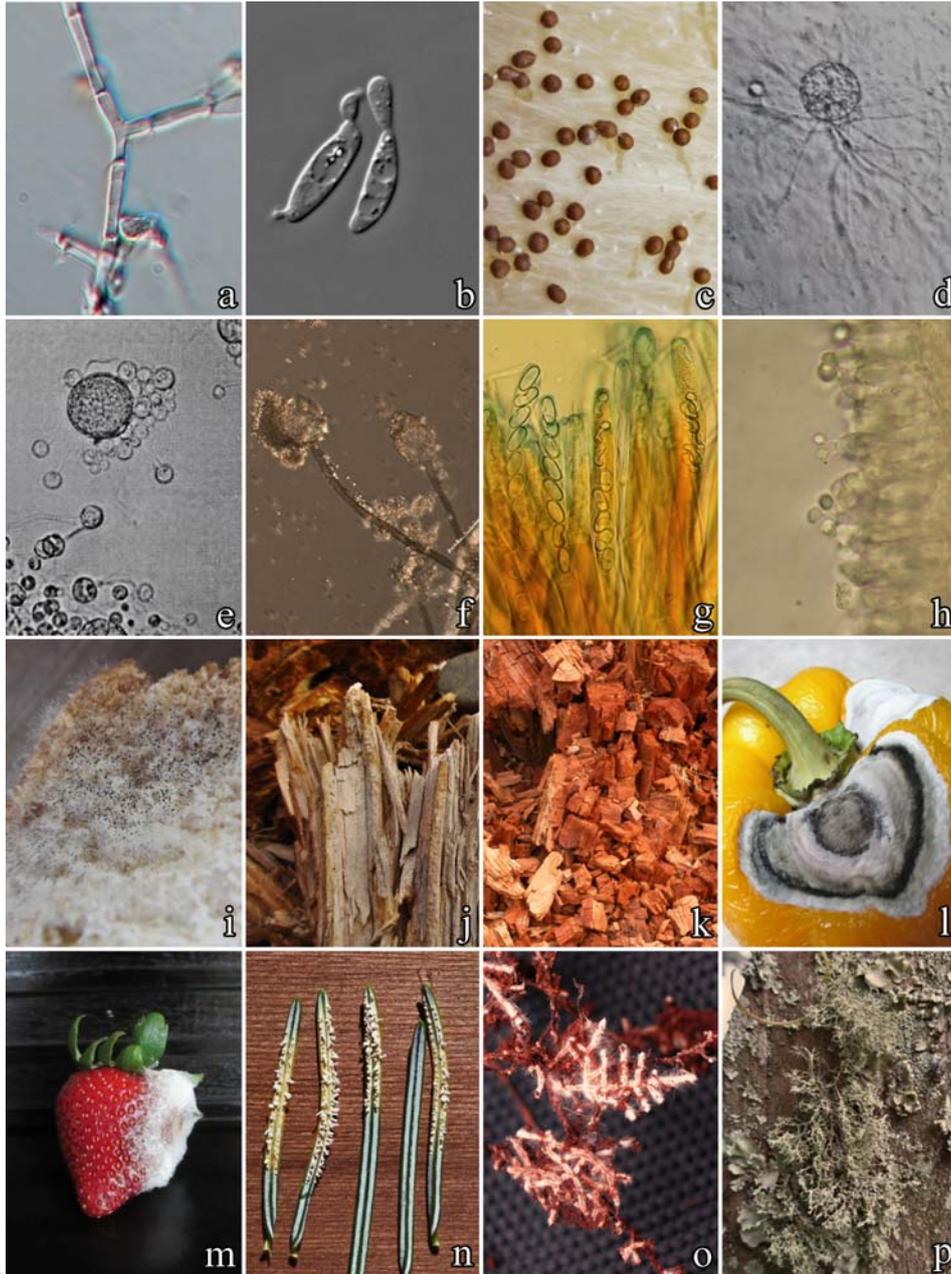


Figure 1. General features of Fungi.

TO ACCESS ALL THE 47 PAGES OF THIS CHAPTER,
Visit: <http://www.eolss.net/Eolss-sampleAllChapter.aspx>

Bibliography

- Agrios G.N. (2005). *Plant Pathology* (5th ed.). Elsevier Inc. 922 pp. [This book gives a thorough overview of plant pathogenic fungi]
- Aime M.C., Matheny P.B., Henk D.A., Frieders E.M., Nilsson R.H., Piepenbring M., McLaughlin D.J., Szabo L.J., Begerow D., Sampaio J.P., Bauer R., Weiss M., Oberwinkler F., Hibbett D.S. (2006). An overview of the higher-level classification of Pucciniomycotina based on combined analyses of nuclear large and small subunit rDNA sequences. *Mycologia* 98: 896–905. [This study presents the first multi-locus phylogenetic analyses of Pucciniomycotina]
- Aime M.C., Toome M., McLaughlin D.J. (2012). Pucciniomycotina. In: McLaughlin D.J., Spatafora J.W. (eds), *Mycota Vol. VII: Systematics and Evolution* (2nd ed.). In press. [This book chapter reviews the most up-to-date knowledge of the biology, ecology and systematics of Pucciniomycotina]
- Alexopoulos C.J., Mims C.W. Blackwell M. (1996). *Introductory Mycology* (4th ed). John Wiley & Sons, Inc. 880 pp. [This book covers basic and advanced concepts of mycology in thorough detail]
- Arnold A.E., Maynard Z., Gilbert G.S., Coley P.D., Kursar T.A. (2000). Are tropical fungal endophytes hyperdiverse? *Ecology Letters* 3, 267–274. [This pioneering study shows that there is a tremendous amount of undiscovered fungal diversity inside plants]
- Begerow D., Stoll M., Bauer R. (2006). A phylogenetic hypothesis of Ustilaginomycotina based on multiple gene analyses and morphological data. *Mycologia* 98: 906–916. [This study provides a phylogenetic overview of Ustilaginomycotina]
- Blackwell M., Hibbett D.S., Taylor J.W., Spatafora J.W. (2006). Research coordination networks: a phylogeny for kingdom Fungi (Feep Hypha). *Mycologia* 98, 829–837. [This paper provides an overview of past and current fungal phylogenetic studies]
- Blackwell M. (2010). Fungal evolution and taxonomy. *BioControl* 55, 7–16. [This review covers various aspects of fungal evolution and taxonomy, with an emphasis on insect-associated fungi]
- Blackwell M. (2011). The Fungi: 1, 2, 3 ... 5.1 million species? *American Journal of Botany* 98, 426–438. [This article summarizes the studies estimating the number of fungal species]
- Blehert D.S., Hicks A.C., Behr M., Meteyer C.U., Berlowski-Zier B.M., Buckles E.L., Coleman J.T.H., Darling S.R., Gargas A., Niver R., Okoniewski J.C., Rudd R.J., Stonem W.B. (2009). Bat white-nose syndrome: an emerging fungal pathogen? *Science* 323, 227. [This is an article reporting the spreading occurrence of white-nose syndrome of bats and describing the growth conditions of the fungus associated with the disease]
- Butt T.M., Jackson C.V., Magan N. (eds.) (2001). *Fungi as biocontrol agents: progress, problems and potential*. CABI Publishing, USA. 385 pp. [This book describes the use of fungi in biocontrol]
- Cairney J.W.G., Meharg A.A. (2003). Ericoid mycorrhiza: a partnership that exploits harsh edaphic conditions. *European Journal of Soil Science* 54, 735–740. [This review covers the role of ericoid mycorrhiza in the survival of plants in stressful environments]
- Casadevall A. (2005). Fungal virulence, vertebrate endothermy, and dinosaur extinction: is there a connection? *Fungal Genetics and Biology* 42, 98–106. [This paper discusses the possible role of fungal pathogens in the extinction of dinosaurs and demonstrates the role of body temperature in fungal infection development]
- Conti M.E., Cecchetti G. (2001). Biological monitoring: lichens as bioindicators of air pollution assessment – a review. *Environmental Pollution* 114, 471–492. [This review focuses on the use of lichens as bioindicators and provides a list of lichens that have been used in such studies]
- Corradi N., Keeling P.J. (2009). Microsporidia: a journey through radical taxonomical revisions. *Fungal Biology Reviews* 23, 1–8. [This paper reviews how the position of Microsporidia on the Tree of Life has changed over time]
- Cummins G.B., Hiratsuka Y. (2003). *Genera of Rust Fungi*. APS Press, St. Paul, Minnesota. 225 pp. [This monograph of rusts provides general information about the biology of Pucciniales and describes the accepted genera]

- De Priest P.T. (2004). Early molecular investigations of lichen-forming symbionts: 1986–2001. *Annual Review of Microbiology* 58: 273–301. [This review covers the main aspects of lichen formation and recent advances in lichenology]
- De Roman M. (2010). The contribution of wild fungi to diet, income and health: a world review. In: Rai, M., Kövocs, G. (eds), *Progress in Mycology*. Scientific Publishers and Springer, 327–348. [This book chapter is an interesting summary about the consumption of fungi among various cultures around the world and reviews the nutritional value of fungi in human diets]
- Divon H.H., Fluhr R. (2007). Nutrition acquisition strategies during fungal infection of plants. *FEMS Microbiology Letters* 226, 65–74. [This review provides information about how plant pathogenic fungi obtain nutrients from their hosts and discusses the signaling involved in the development of an infection]
- Ferguson B.A., Dreisbach T.A., Parks C.G., Filip G.M., Schmitt C.L. (2003). Coarse-scale population structure of pathogenic *Armillaria* species in a mixed-conifer forest in the Blue Mountains of northeast Oregon. *Canadian Journal of Forest Research* 33, 612–623. [This paper compares the size and age of various *Armillaria* species, among which is the largest fungal individual known to date]
- Fisher M.C., Graner T.W.J., Walker S.F. (2009). Global emergence of *Batrochochytrium dendrobatidis* and amphibian chytridiomycosis in space, time, and host. *Annual Reviews of Microbiology* 63, 291–310. [This review provides an up-to-date summary about amphibian chytridiomycosis]
- Fuller M.S. (1978). *Lower fungi in the laboratory*. University of Georgia, USA. 213 pp. [This book covers techniques for growing and studying the morphology of basal fungi]
- Gams W., Diederich P., Pöldmaa K. (2004). Fungicolous Fungi. In: Mueller, G.M., Bills, G.F., Foster, M.S. (eds) *Biodiversity of Fungi: inventory and monitoring methods*. Elsevier, 343–392. [This book chapter provides an overview of mycoparasitic fungi]
- Gleason F.H., Marano A.V., Johnson P., Martin W.W. (2010). Blastocladian parasites of invertebrates. *Fungal Biology Reviews* 24, 56–67. [This is a review about the Blastocladiomycota species causing diseases of invertebrates]
- Goffeau A., Barrell B.G., Bussey H., Davis R.W., Dujon B., Feldmann H., Galibert F., Hoheisel J.D., Jacq C., Johnston M., Louis E.J., Mewes H.W., Murakami Y., Philippsen P., Tettelin H., Oliver S.G. (1996). Life with 6000 genes. *Science* 274, 546–567. [This paper presents the genome of the baker's yeast – *Saccharomyces cerevisiae*]
- Harman G.E. (2000). Myths and dogmas of biocontrol changes in perceptions derived from research on *Trichoderma harzianum* T-22. *Plant Disease* 84: 377–393. [This paper reviews the use of fungi as biocontrol agents]
- Heckman D.S., Geiser D.M., Eidell B.R., Stauffer R.L., Kardos N., Hedges S.B. (2001). Molecular evidence for the early colonization of land by fungi and plants. *Science* 293, 1129–1133. [This paper provides evidence that major fungal lineages existed long before the occurrence of land plants]
- Heitman J. (2011). Microbial pathogens in the fungal kingdom. *Fungal Biology Reviews* 25, 48–60. [This review is a good reference for various human and animal pathogens in Fungi]
- Hibbett D.S. (2006). A phylogenetic overview of the Agaricomycotina. *Mycologia* 98, 917–925. [This study presents a phylogenetic overview of Agaricomycotina]
- Hibbett D.S., Binder M., Bischoff J.F., Blackwell M., Cannon P.F., Eriksson O.E., Huhndorf S., James T., Kirk P.M., Lücking R., Lumbsch T., Lutzoni F., Matheny P.B., Mclaughlin D.J., Powell M.J., Redhead S., Schoch C.L., Spatafora J.W., Stalpers J.A., Vilgalys R., Aime M.C., Aptroot A., Bauer R., Begerow D., Benny G.L., Castlebury L.A., Crous P.W., Dai Y.-C., Gams W., Geiser D.M., Griffith G.W., Gueidan C., Hawksworth D.L., Hestmark G., Hosaka K., Humber R.A., Hyde K., Ironside J.E., Kõljalg U., Kurtzman C.P., Larsson K.-H., Lichtwardt R., Longcore J., Miądlikowska J., Miller A., Moncalvo J.-M., Mozley-Standridge S., Oberwinkler F., Parmasto E., Reeb V., Rogers J.D., Roux C., Ryvarden L., Sampaio J.P., Schüßler A., Sugiyama J., Thorn R.G., Tibell L., Untereiner W.A., Walker C., Wang Z., Weir A., Weiß M., White M.M., Winka K., Yao Y.-J., Zhang N. (2007). A higher-level phylogenetic classification of the Fungi. *Mycological Research* 111: 509–547. [This paper proposes a single classification for Fungi down to the ordinal level]

James T.Y., Porter D., Leander C.A., Vilgalys R., Longcore J.E. (2000). Molecular phylogenetics of the Chytridiomycota support the utility of ultrastructural data in chytrid systematics. *Canadian Journal of Botany* 78, 336–350. [This study demonstrates polyphyly of Chytridiomycota and shows connections between molecular results and the morphology of zoospores]

James T.Y., Letcher P.M., Longcore J.E., Mozley-Standridge S., Porter D., Powell M.J., Griffith G.W., Vilgalys R. (2006). A molecular phylogeny of the flagellated fungi (Chytridiomycota) and description of a new phylum (Blastocladiomycota). *Mycologia* 98, 860–871. [This study describes Blastocladiomycota and resolves the relationships between the orders of zoosporic fungi]

James T.Y., Kauff F., Schoch C., Matheny P.B., Hofstetter V., Cox C., Celio G., Gueidan C., Fraker E., Miadlikowska J., Lumbsch H.T., Rauhut A., Reeb V., Arnold A.E., Amtoft A., Stajich J.E., Hosaka K., Sung G.-H., Johnson D., O'Rourke B., Crockett M., Binder M., Curtis J.M., Slot J.C., Wang Z., Wilson A.W., Schüßler A., Longcore J.E., O'Donnell K., Mozley-Standridge S., Porter D., Letcher P.M., Powell M.J., Taylor J.W., White M.M., Griffith G.W., Davies D.R., Humber R.A., Morton J.B., Sugiyama J., Rossman A.Y., Rogers J.D., Pfister D.H., Hewitt D., Hansen K., Hambleton S., Shoemaker R.A., Kohlmeyer J., Volkmann-Kohlmeyer B., Spotts R.A., Serdani M., Crous P.W., Hughes K.W., Matsuura K., Langer E., Langer G., Untereiner W.A., Lücking R., Büdel B., Geiser D.M., Aptroot A., Diederich P., Schmitt I., Schultz M., Yahr R., Hibbett D., Lutzoni F., McLaughlin D., Spatafora J., Vilgalys R. (2006). Reconstructing the early evolution of the fungi using a six gene phylogeny. *Nature* 443:818–822. [This study presents a multi-gene phylogeny of Fungi, with emphasis on resolving the basal lineages]

Keeling P.J., Corradi N. (2011). Shrink it or lose it. Balancing loss of function with shrinking genomes in the microsporidia. *Virulence* 2, 67–70. [This is a good review about the genomes of Microsporidia, demonstrating the decrease in gene length and other reductions]

Keeling P.J., Fast, N.M. (2002). Microsporidia: biology and evolution of highly reduced intracellular parasites. *Annual Review of Microbiology* 56, 93–116. [This is a thorough review of microsporidian research]

Keeling P.J., Luker M.A., Palmer J.D. (2000). Evidence from beta-tubulin phylogeny that Microsporidia evolved from within the Fungi. *Molecular Biology and Evolution* 17, 23–31. [This study demonstrates that Microsporidia is a member of kingdom Fungi, although the positioning remains unresolved]

Keeling P. (2009). Five questions about Microsporidia. *PLoS Pathogens* 5, e1000489. [A key paper summarizing the main aspects of Microsporidia]

Keller N.P., Turner G., Bennett J.W. (2005). Fungal secondary metabolism – from biochemistry to genomics. *Nature Reviews Microbiology* 3, 937–947. [This review covers the recent research on fungal metabolites and their use in biotechnology]

Kirk P.M., Cannon P.F., Minter D.W., Stalpers J.A. (2008). *Dictionary of the Fungi* (10th ed). Wallingford, UK: CABI. 771 pp. [The most current edition of the indispensable classic that provides definitions of mycological terms and all known taxa, including relevant references]

Krüger M., Krüger C., Walker C., Stockinger H., Schüssler A. (2012). Phylogenetic reference data for systematics and phylotaxonomy of arbuscular mycorrhizal fungi from phylum to species level. *New Phytologist* 193, 970–984. [This paper provides the basis for molecular studies of Glomeromycota]

Lehmkuhl J.F., Gould L.E., Cázares E., Hosford D.R. (2004). Truffle abundance and mycophagy by northern flying squirrels in eastern Washington forests. *Forest Ecology and Management* 200, 49–65. [This paper demonstrates the importance of truffles in the diet of squirrels and their role in truffle life cycle]

Liggenstoffer A.S., Youssef N.H., Couger M.B., Elshahed M.S. (2010). Phylogenetic diversity and community structure of anaerobic gut fungi (phylum Neocallimastigomycota) in ruminant and non-ruminant herbivores. *The ISME Journal*, 1–12. [This study of rumen fungi indicates a great diversity in Neocallimastigomycota]

Liu Y., Leigh J.W., Brinkmann H., Cushion M.T., Rodriguez-Ezpeleta N., Philippe H., Lang B.F. (2009). Phylogenomic analyses support the monophyly of Taphrinomycotina, including *Schizosaccharomyces* fission yeasts. *Molecular Biology and Evolution* 26, 27–34. [This article places the *Schizosaccharomyces* in the Taphrinomycotina]

Longcore J.E., Pessier A.P., Nichols D.K. (1999). *Batrachochytrium dendrobatidis* gen. et sp. nov., a chytrid pathogenic to amphibians. *Mycologia* 91, 219–227. [This article describes the causal agent of amphibian decline]

Louis E. (2011). *Saccharomyces cerevisiae*: gene annotation and genome variability, state of the art through comparative genomics. *Methods in Molecular Biology* 759, 31–40. [This paper summarizes the genomic studies of the baker's yeast]

Lutzoni F., Kauff F., Cox C.J., McLaughlin D., Celio G., Dentinger B., Padamsee M., Hibbett D., James T.Y., Baloch E., Grube M., Reeb V., Hofstetter V., Schoch C., Arnold A.E., Miadlikowska J., Spatafora J., Johnson D., Hambleton S., Crockett M., Shoemaker R., Sung G.-H., Lücking R., Lumbsch T., O'Donnell K., Binder M., Diederich P., Ertz D., Gueidan C., Hansen K., Harris R.C., Hosaka K., Lim Y.-W., Matheny B., Nishida H., Pfister D., Rogers J., Rossman A., Schmitt I., Sipman H., Stone J., Sugiyama J., Yahr R., Vilgalys R. (2004). Assembling the fungal tree of life: progress, classification, and evolution of subcellular traits. *American Journal of Botany* 91, 1446–1480. [This paper provides a phylogenetic synthesis of Fungi and discusses the evolution of septal features]

Malloch D. (1995). Fungi with heteroxenous life histories. *Canadian Journal of Botany* 73, S1334–S1342. [This review discusses the heteroecious life styles in Fungi]

Matheny P.B., Gossmann J.A., Zalar P., Arun Kumar T.K., Hibbett D.S. (2006). Resolving the phylogenetic position of the Wallemiomycetes: an enigmatic major lineage of Basidiomycota. *Canadian Journal of Botany* 84, 1794–1805. [This phylogenetic study of Wallemiomycetes confirms their placement in Basidiomycota *incertae sedis*]

Martin F., Kohler A., Murat C., Balestrini R., Coutinho P.M., Jaillon O., Montanini B., Morin E., Noel B., Percudani R., Porcel B., Rubini A., Amicucci A., Amselem J., Anthouard V., Arcioni S., Artiguenave F., Aury J.-M., Ballario P., Bolchi A., Brenna A., Brun A., Buée M., Cantarel B., Chevalier G., Couloux A., Da Silva C., Denoeud F., Duplessis S., Ghignone S., Hilselberger B., Iotti M., Marçais B., Mello A., Miranda M., Pacioni G., Quesneville H., Riccioni C., Ruotolo R., Splivallo R., Stocchi V., Tisserant E., Viscomi A.R., Zambonelli A., Zampieri E., Henrissat B., Lebrun M.-H., Paolocci F., Bonfante P., Ottonello S., Wincker P. (2010). Périgord black truffle genome uncovers evolutionary origins and mechanisms of symbiosis. *Nature* 464, 1033–1038. [This paper presents the genome of *Tuber melanosporum*]

McDonald B.A., Linde C. (2002). Pathogen population genetics, evolutionary potential, and durable resistance. *Annual Review of Phytopathology* 40, 349–379. [This review provides an overview of the evolution of plant pathogens, including fungi]

McGovern P.E., Fleming S.J., Katz S.H. (eds) (2005). *The origins and ancient history of wine*. Taylor & Francis. 403 pp. [This book details the history of wine making and the yeast involved in it]

Minter D.W. (2010). International society for fungal conservation. *IMA Fungus* 1, 27–29. [This paper emphasizes the importance of fungal conservation]

Nagano Y., Nagahama T. (2012). Fungal diversity in deep-sea extreme environments. *Fungal Biology* doi: 10.1016/j.funeco.2012.01.004. [This is a recent mini-review of fungal biodiversity in marine environments]

O'Brien B.L., Parrent J.L., Jackson J.A., Moncalvo J.M., Vilgalys R. (2005). Fungal community analysis by large-scale sequencing of environmental samples. *Applied and Environmental Microbiology* 71, 5544–5550. [This paper presents a study of fungi from forest soil and demonstrates a great diversity of fungi in these habitats]

O'Donnell K. (1979). *Zygomycetes in culture*. University of Georgia, USA. 257 pp. [This classic provides techniques for growing and identifying zygomycetes]

Orpin C.G. (1975). Studies on the rumen flagellate *Neocallimastix frontalis*. *Microbiology* 91, 249–262. [The description of the first anaerobic fungus]

Padamsee M., Kumar A., Riley R., Binder M., Boyd A., Calvo A.M., Furukawa K., Hesse C., Hohmann S., James T.Y., Labutti K., Lapidus A., Lindquist E., Lucas E., Lucas S., Miller K., Shantappa S., Grigorjev I., Hibbett D.S., McLaughlin D.J., Spatafora J.W., Aime M.C. (2012). The genome of the xerotolerant mold *Wallemia sebi* reveals adaptations to osmotic stress and suggests cryptic sexual

reproduction. *Fungal Genetics and Biology* 49: 217–226. [This study combines genomic and ultrastructural data to resolve the phylogenetic placement of Wallemiomycetes as well as make predictions about its reproductive biology and ability to colonize osmotically stressful environments]

Peintner U., Pöder R., Pümpel T. (1998). The iceman's fungi. *Mycological Research* 102, 1153–1162. [This paper describes the fungi found with the body of the Iceman and the ways in which they were probably used]

Porter T.M., Martin W., James T.Y., Longcore J.E., Gleason F.H., Adler P.H., Letcher P.M., Vilgalys R. (2011). Molecular phylogeny of the *Blastocladiomycota* (Fungi) based on nuclear ribosomal DNA. *Fungal Biology* 115, 381–392. [This paper presents the most recent phylogenetic study of Blastocladiomycota]

Porter T.M., Schadt C.W., Rizvi L., Martin A.P., Schmidt S.K., Scott-Denton L., Vilgalys R., Moncalvo J.M. (2008). Widespread occurrence and phylogenetic placement of a soil clone group adds a prominent new branch to the fungal tree of life. *Molecular Phylogenetics and Evolution* 46: 635–644. [This landmark paper uncovered a previously unknown lineage of fungi termed Soil Clone Group 1 from environmental sequence data]

Redecker D., Raab P. (2006). Phylogeny of the Glomeromycota (arbuscular mycorrhizal fungi): recent developments and new gene markers. *Mycologia* 98, 885–895. [This study presents a reassessment of Glomeromycota phylogenetics]

Remy W., Taylor T.N., Hass H., Kerp H., (1994). Four hundred-million-year-old vesicular arbuscular mycorrhizae. *Proceedings of the National Academy of Sciences of the United States of America* 91, 11841–11843. [This article presents the fossil findings that demonstrate the presence of arbuscular mycorrhiza-like associations in the Early Devonian]

Rosendahl S. (2008). Communities, populations and individuals of arbuscular mycorrhizal fungi. *New Phytologist* 178, 253–266. [This review examines species concepts and population biology of arbuscular mycorrhizal fungi]

Rosling A., Cox F., Cruz-Martinez K., Ihrmark K., Grelet G.-A., Lindahl B.D., Menkins A., James T.Y. (2011). Archaeorhizomycetes: unearthing an ancient class of ubiquitous soil fungi. *Science* 333, 876–879. [This article describes a new class of Taphrinomycotina that accommodates the group of fungi previously known as Soil Clone Group 1]

Schell W.A., Lee A.G., Aime M.C. (2011). A new lineage in Pucciniomycotina: class Tritirachiomycetes, order Tritirachiales, family Tritirachiaceae. *Mycologia* 103, 1331–1340. [This study describes a new basidiomycete class of anamorphic molds]

Schoch C.L., Seifert K.A., Huhndorf S., Spouge J.L., Robert V., Bolchacova E., Voigt K., Chen W., Crous P.W., Miller A.N., Wingfield M.J., Okada G., Suzuki M., Hambleton S., Levesque A., Otte J., Schmitt I., Boonyuen N., Jones E.B.G., Suetrong S., Tretter E., White M.M., Hognabba F., Stenroos S., Hagen F., Eberhardt U., Quaedvlieg W., Boekhout T., Damm U., De Hoog S., Groenewald J.Z., Groenewald M., Walther G., Duong V., Schubler A., Qing C., Yang Z.L., Bogale M., Untereiner W.A., Maganti H., Xu J.P., Leavitt S.D., Lumbsch H.T., Hansen K., Olariaga I., Duong T.A., De Beer Z.W., Nilsson R.H., Cardinali G., Burgaz A.R., Crespo A., Del-Prado R., Divakar P.K., Ruibal C., Sotome K., Huhtinen S., Fliegerova K., Douglas B., Griffith G.W., An K.D., Johnston P.R., Park D., Weir B.S., Blackwell M., Urbina H., Aime M.C., Heller G., McTaggart A.R., Hyde K.D., Kurtzman C.P., Luangsa-Ard J.J., Mongkols Amrit S., Hosaka K., Tedersoo L., Bergeron M.J., Hamelin R.C., Vialle A., Okane I., Liimatainen K., Niskanen T., Dieguez-Urbeondo J., Duenas M., Garcia M.A., Martin M.P., Pino-Bodas R., Sarmiento-Ramirez J.M., Telleria M.T., Zamora J.C., Coppins B.J., Harrold P., Hollingsworth P., Kelly L.J., Yahr R., Griffiths K., May T., Stefani F.O.P., Yurkov A., Begerow D., Bai F.Y., Cai L., Guo L.D., Raja H.A., Redecker D., Stockinger H., Shearer C., Nagy L.G., Nyilasi I., Papp T., Petkovits T., Vagvolgyi C., Koljalg U., Barreto R., Buyck B., Chaverri P., Dentinger B., Elshahed M.S., Ge Z.W., Gryzenhout M., Ho H.M., Hofstetter V., Hong S.B., Houbraken J., Hughes K., James T., Johnson E., Kirk P., Kovacs G.M., Landvik S., Liggenstoffer A.S., Lombard L., Meyer W., Moncalvo J.M., Rintoul T., Suh S.O., Tanaka K., Vu D., Wang Y., Weiss M., Zhang N., Zhuang W.Y., Schindel D. (2012). The internal transcribed spacer as a universal DNA barcode marker for Fungi. *Proceedings of the National Academy of Sciences* 109, 6241–6246. [The study that provides a default barcode locus for kingdom Fungi]

Schoch C.L., Sung G.H., López-Giráldez F., Townsend J.P., Miadlikowska J., Hofstetter V., Robbertse B., Matheny P.B., Kauff F., Wang Z., Gueidan C., Andrie R.M., Trippe K., Ciufetti L.M., Wynns A., Fraker E., Hodkinson B.P., Bonito G., Groenewald J.Z., Arzanlou M., de Hoog G.S., Crous P.W., Hewitt D., Pfister D.H., Peterson K., Gryzenhout M., Wingfield M.J., Aptroot A., Suh S.O., Blackwell M., Hillis D.M., Griffith G.W., Castlebury L.A., Rossman A.Y., Lumbsch H.T., Lücking R., Büdel B., Rauhut A., Diederich P., Ertz D., Geiser D.M., Hosaka K., Inderbitzin P., Kohlmeyer J., Volkmann-Kohlmeyer B., Mostert L., O'Donnell K., Sipman H., Rogers J.D., Shoemaker R.A., Sugiyama J., Summerbell R.C., Untereiner W., Johnston P.R., Stenroos S., Zuccaro A., Dyer P.S., Crittenden P.D., Cole M.S., Hansen K., Trappe J.M., Yahr R., Lutzoni F., Spatafora J.W. (2009). The Ascomycota tree of life: A phylum-wide phylogeny clarifies the origin and evolution of fundamental reproductive and ecological traits. *Systematic Biology* 58, 224–239. [This paper presents one of the most up to date phylogenies of Ascomycota]

Smith S.E., Read D.J. (2008). *Mycorrhizal Symbiosis*, 3rd edition. Elsevier, 769 pp. [This book gives a complete overview of the mycorrhiza types, their biology and role in natural and agricultural systems]

Spatafora J.W., Johnson D., Sung G.-H., Hosaka K., O'Rourke B., Serdani M., Spotts R., Lutzoni F., Hofstetter V., Fraker E., Gueidan C., Miadlikowska J., Reeb V., Lumbsch T., Lücking R., Schmitt I., Aptroot A., Roux C., Miller A., Geiser D., Hafellner J., Hestmark G., Arnold A.E., Büdel B., Rauhut A., Hewitt D., Untereiner W., Cole M.S., Scheidegger C., Schultz M., Sipman H., Schoch C. (2006). A five-gene phylogenetic analysis of the Pezizomycotina. *Mycologia* 98, 1020–1030. [This paper gives an overview of Pezizomycotina phylogeny and general characteristics]

Spatafora J.W., Sung G.-H., Sung J.-M., Hywel-Jones N.L., White Jr J.F. (2007). Phylogenetic evidence for an animal pathogen origin of ergot and the grass endophytes. *Molecular Ecology* 16, 1701–1711. [This study examines the evolutionary origins of the fungi that cause ergot of grains]

Stajich J.E., Berbee M.L., Blackwell M., Hibbett D.S., James T.Y., Spatafora J.W., Taylor J.W. (2009). The Fungi. *Current Biology* 19, R840–R845. [A brief review of the importance and evolution of Fungi]

Sugiyama J., Hosaka K., Suh S.-O. (2006). Early diverging Ascomycota: phylogenetic divergence and related evolutionary enigmas. *Mycologia* 98, 996–1005. [This study presents phylogenetic studies of Taphrinomycotina]

Suh S.-O., Blackwell M., Kurtzman C.P., Lachance M.-A. (2006). Phylogenetics of Saccharomycetales, the ascomycete yeasts. *Mycologia* 98, 1006–1017. [This study presents phylogenetic studies of Saccharomycotina]

Taylor J.W., Jacobson D.J., Fisher M.C. (1999). The evolution of asexual fungi: reproduction, speciation and classification. *Annual Reviews of Phytopathology* 37, 197–246. [This is a thorough review of asexual reproduction in Fungi]

Turchetti B., Thomas Hall S.R., Connell L.B., Branda E., Buzzini P., Theelen B., Müller W.H., Boekhout T. (2011). Psychrophilic yeasts from Antarctica and European glaciers: description of *Glaciozyma* gen. nov., *Glaciozyma martini* sp. nov. and *Glaciozyma watsonii* sp. nov. *Extremophiles* 15: 573–586. [This article describes new basidiomycete yeasts isolated from extreme cold environments]

Van Kan J.A.L. (2008). Licensed to kill: the lifestyle of a necrotrophic plant pathogen. *Trends in Plant Science* 11, 247–253. [This article is a review on necrotrophs, their modes of infection and life cycles]

Vánky K. 2012. *Smut Fungi of the World*. APS Press. 1458 pp. [This book is a complete and up to date monograph of smut fungi]

van Peer A.F., Wang F., van Driel K.G., de Jong J.F., van Donselaar E.G., Müller W.H., Boekhout T., Lugones L.G., Wösten H.A. (2010). The septal pore cap is an organelle that functions in vegetative growth and mushroom formation of the wood-rot fungus *Schizophyllum commune*. *Environmental Microbiology* 12: 833–844. [This study examines the function of the septal pore cap in Agaricomycotina]

Walsh T.J., Groll A.H. (1999). Emerging fungal pathogens: evolving challenges to immunocompromised patients for the twenty-first century. *Transplant Infectious Disease* 1, 247–261. [This review discusses the role of fungal pathogens in modern medicine]

Wasser S.P. (2011). Current findings, future trends, and unsolved problems in studies of medicinal mushrooms. *Applied Microbiology and Biotechnology* 89, 1323–1332. [This is a review of the use of medicinal mushrooms]

Whisler H.C., Zebold S.L., Shemanchuk J.A. (1975). Life history of *Coelomomyces psorophorae*. *Proceedings of the National Academy of Sciences of the United States of America* 72, 693–696. [This study demonstrates the heteroecious life cycle of *Coelomomyces*]

White M.M., James T.Y., O'Donnell K., Cafaro M.J., Tanabe Y., Sugiyama J. (2006). Phylogeny of the Zygomycota based on nuclear ribosomal sequence data. *Mycologia* 98, 872–884. [This study presents a phylogenetic overview of zygomycetes]

Woodward S., Stenlid J., Karjalainen R., Hüttermann A. (eds) (1998). *Heterobasidion annosum: biology, ecology, impact and control*. CABI Publishing, UK. 589 pp. [This is a compilation of studies on all important aspects of the root rot fungus that causes major economical losses in temperate forests]

Zugmaier W., Oberwinkler F. (1995). Tremelloid haustorial cells with haustorial filaments and potential host range of *Tremella mesenterica*. *Nordic Journal of Botany* 15, 207–213. [This paper describes tremelloid haustoria, the characteristic feeding structures of mycoparasitic fungi]

Biographical Sketches

Merje Toome received her BSc in Biology from University of Tartu (Estonia) in 2004 and her PhD in Plant Pathology from the Estonian University of Life Sciences in 2010. She is a member of Mycological Society of America, Estonian Naturalists' Society, and American Phytopathological Society. At present she is a postdoctoral fellow at the Louisiana State University Agricultural Center. Her main research activities include the phylogeny and genomics of Pucciniomycotina and the taxonomy and ecology of some rust species and their relatives.

Mary Catherine Aime received her B.S. in 1996 and Ph.D. in 2001 in Biology from Virginia Polytechnic Institute and State University. She is a Fellow of the Mycological Society of America, the Linnaean Society of London, and the Explorer's Club. Her research primarily focuses on the systematics of Basidiomycota, especially of rust fungi and Pucciniomycotina, and the biodiversity of tropical fungi.