

TP53 GENE AND P53 PROTEIN AS TARGETS IN CANCER MANAGEMENT AND THERAPY

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Contents

1. Introduction
 2. TP53 Mutations and Human Cancer
 3. The p53 Protein—A Sensor of Genotoxic Stress
 4. Gene Therapy Using TP53
 - 4.1. Replacement Gene Therapy
 - 4.2. E1B-defective Adenoviruses
 5. The p53 Pathway as a Target for Pharmacological Intervention
 - 5.1. Restoration of Wild-type p53 Functions
 - 5.2. Modulation of Wild-type p53
 - 5.3. Amifostine—A Drug that Targets the p53 Pathway
 6. TP53 in Cancer Detection and Monitoring
 - 6.1. Auto Antibodies to p53
 - 6.2. Plasmatic Free TP53 DNA
 7. Perspectives and Future Challenges
- Glossary
Bibliography
Biographical Sketches

Summary

The TP53 gene is commonly altered by mutation in many types of human cancers. The p53 protein is an inducible transcription factor that plays multiple, anti-proliferative roles in response to exposure to many forms of stress, including in particular various classes of DNA-damaging agents. Thus, p53 function is essential for the genetic homeostasis of cells exposed to mutagens. In a physiological context, the status of p53 controls the sensitivity of cells to environmental mutagens. In a pathological context, the status of p53 is considered as a key factor in the response to cancer cells to cytotoxic therapies. Thus, control of p53 functions is a very promising target for cancer management.

Given the central importance of TP53 in carcinogenesis, there have already been many attempts at restoring or modulating p53 protein functions through genetic or pharmacological approaches. First, gene therapy experiments using either retroviral or adenoviral vectors have given a proof of principle for the restoration of wild-type TP53 function in cancer cells containing mutant alleles. However, the therapeutic efficiency of such approaches is currently limited by difficulties in targeting cancer cells, in

obtaining high expression levels of the transgene, and in maximizing so-called by-stander effects. Alternative approaches to “replacement” gene therapies are also being developed. For example, the ONYX-015 viral vector is a defective adenovirus that can selectively kill cancer cells with deficient TP53 functions. This vector is currently under clinical evaluation.

The rapid accumulation of knowledge on p53 protein functions has led to the development of a number of pharmacological approaches. One of the most promising makes use of small peptides that target specific regulatory domains of the protein to either activate wild-type p53 or restore function of mutant p53. Other methods address specific biochemical properties of p53, such as sensitivity to redox signals. For example, we have shown that the chemoprotective drug amifostine could activate wild-type p53 and induce preferentially cell cycle arrest in cells containing wild-type alleles. This specific response may contribute to the protective effects of the drug.

It should be kept in mind that the TP53 gene and its product can be used as biomarker for cancer detection, diagnosis, and prognosis. This can be achieved by the analysis of gene or protein status in cancer or pre-cancer lesions, as well as by detection of gene fragments or antibodies in plasma or serum. All these aspects are currently the topic of intense research efforts. However, their outcome on the clinical management of cancer will not be available for several years.

1. Introduction

The TP53 gene was discovered twenty years ago, but it took about ten years before a clear idea of its function eventually emerged in the early 1990s. Experimental studies then demonstrated that this gene encodes a tumor suppressor, and molecular pathological approaches showed that this suppressor was inactive in a majority of human cancers. Since then, the TP53 gene and its product, the p53 protein, have occupied the center stage of the molecular biology of cancer, and have raised great expectations for applications leading to better cancer management or therapy.

TP53 is special among cancer genes in at least three respects. First, most of its alterations in cancers are missense mutations. This is uncommon for suppressor genes, which are classically inactivated by deletions or non-sense mutations. Second, it is altered at a significant frequency (between 20 and 80%) in almost every human cancer, irrespective of the organ site or the histological type. This observation stresses the central role of p53 as one of the basic elements of the cellular growth control machinery. Third, the protein itself is apparently essential for many aspects of normal life. This also contrasts with many tumor suppressors, which encode “vital” proteins. Strikingly, mice deficient in TP53 by homologous recombination show essentially normal development and behavior. However, when they reach 20 or 30 weeks of age, most of them die from multiple, early cancers. Thus, TP53 may be considered as, in the words of M. Oren, the “ultimate tumor suppressor gene”, the function of which is essentially to protect cells against the occurrence and development of cancer.

This very special position of p53 in the control of cell proliferation is due to two biological characteristics. First, p53 is an inducible protein at the post-

transcriptional level. It is almost absent, or “latent,” in most normal cells and tissues, but becomes stabilized and activated in response to many forms of cellular stress, in particular stress inducing the formation of DNA-damage. Moreover, p53 is capable of regulating many overlapping pathways. P53 is a transcription factor with more than 30 known target genes in pathways such as cell cycle control, apoptosis, DNA repair, differentiation, and senescence. The protein also acts through direct, complex formation with other cellular components, further increasing the range of responses elicited by p53 activation. Overall, p53 appears to sit at the center of a network of signals that connect stress response (in particular to DNA damage) with growth regulation. This special function has earned p53 the nickname of “guardian of the genome”. Loss of p53 function thus eliminates a protection system by which cells normally regulate their capacity to proliferate in stressful conditions, and increases the likelihood that such cells may acquire other genetic changes during cancer progression.

Therefore, p53 represents an interesting target for genetic or pharmacological intervention in cancer treatment. Below, we briefly review the implication of TP53 in human cancer, and we describe current approaches for cancer gene therapy, pharmacological modulation of p53 protein, and exploitation of TP53 in cancer detection and monitoring.

2. TP53 Mutations and Human Cancer

The human TP53 gene is located in 20 kb of chromosome band 17p13.1. The gene is composed of 11 exons, the first of which is non-coding. The product of the gene is a 53kD nuclear phosphoprotein, composed of 393 amino acids. The functional molecule is a tetramer and acts as a transcriptional factor. It is involved in cell cycle checkpoints, apoptosis, genomic instability, and DNA repair (Figure 1).

The p53 protein is activated in response to genotoxic (DNA-damaging) and non-genotoxic stresses. The stability of the protein is controlled by Mdm2 and by JNK. After activation, p53 regulates genes and proteins involved in cell cycle arrest (in G1, G1/S and G2/M) in replication, transcription, repair, and apoptosis. The p53 protein is constitutively expressed in almost all cell types but has a very rapid turnover and appears to be latent under normal conditions. However, p53 is rapidly converted to an active form in response to a number of physical or chemical DNA-damaging agents such as gamma irradiation, UV rays, oxidizing agents, cytotoxic drugs, and cancer-causing chemicals. Induction of p53 implies nuclear retention, accumulation of the protein as a result of post-translational stabilization, and allosteric conversion to a form with high sequence-specific DNA-binding capacity. This has led to the concept that p53 is specifically activated in response to DNA-damage thus acting as a “guardian” against genotoxic stress.

The p53 protein is a sequence specific transcription factor which binds DNA sequences corresponding to repeats of the consensus motif RRRC(A/T)(T/A)GYYY (where R is a purine and Y pyrimidine). The protein has five structural and functional domains: a N-terminal, transcriptional activation domain, a proline-rich regulatory domain, a sequence-specific DNA-binding domain, an oligomerization domain, and a C-terminal domain involved in the regulation of DNA binding (Figure 2A). In terms of three-

dimensional structure (Figure 2B), the protein is made of a scaffold of beta-sheets that support flexible loops and helices which are in direct contact with DNA. The position of these loops and helices is stabilized by the binding of an atom of zinc.

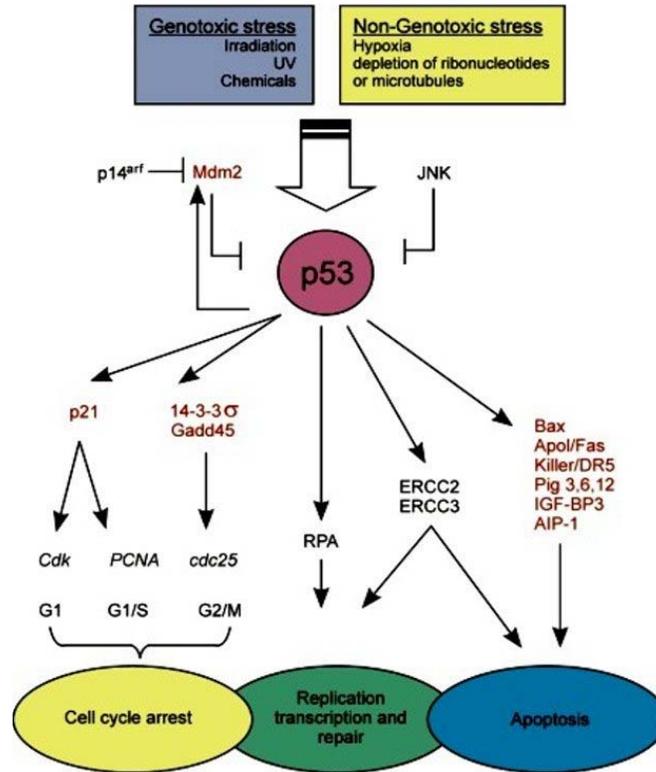


Figure 1: The p53 signaling pathway.

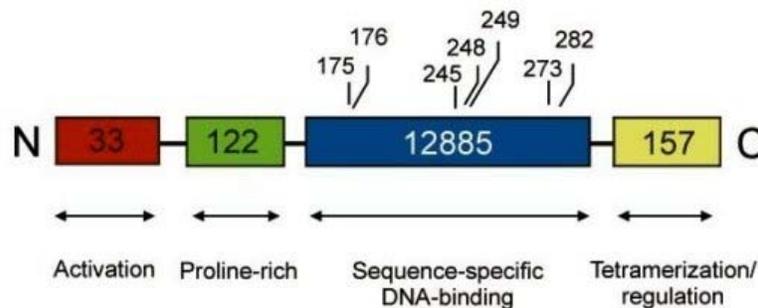


Figure 2: Structure of the p53 protein.

The protein in Figure 2 contains several functional domains, as indicated. The number of mutations detected in the human cancer that falls within each of these domains is given. The most frequently mutated portion is the sequence-specific DNA-binding domain. Within this domain, several residues are “hotspots” for mutation. The three most frequently mutated residues in human cancers are represented using a space-fill

model in which each atom is pictured as a small sphere. The target DNA that p53 binds to is outlined.

In cancer, inactivation of p53 occurs through various mechanisms, including genetic alteration (mutation, deletion), inactivation of the protein by binding to viral or cellular oncoprotein, and sequestration in the cytoplasm. The DNA binding domain contains 93% of all mutations identified to date. This high frequency may be overestimated, since after initial reports that mutations tended to cluster in the central portion of the coding sequence (DNA binding domain), most investigators have limited their analysis to exon 5 to 8. A database of all published mutations is maintained at the International Agency for Research on Cancer. The most frequently mutated residues (Figures 2A and B) are conserved among species and play an important, direct or indirect, role in the contacts between the protein and target DNA. All these mutations result in impaired DNA-binding and loss of transcriptional activity.

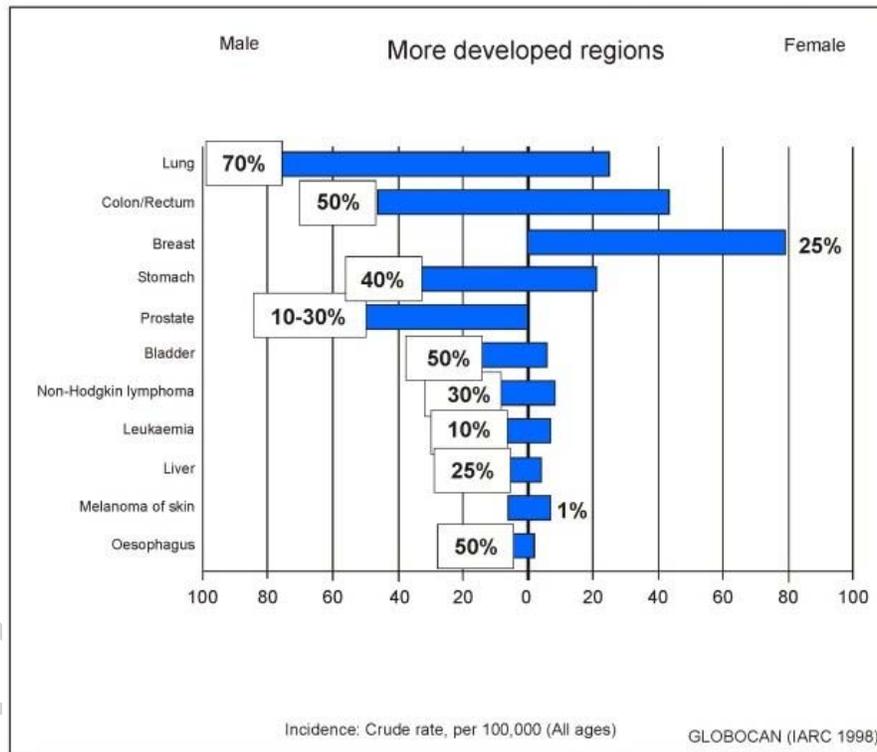


Figure 3: Incidence of cancers in developed countries.

Mutations in the TP53 are found in almost every kind of human tumor. Malignancies in which the mutation prevalence is higher than 50% include skin cancer (except melanoma), late stage cancer of bladder cancers, and carcinomas of the aero-digestive tract. Lymphomas and tumors of the brain, breast, prostate, and liver show an intermediate mutation frequency (15 to 35%). Malignancies with low mutation frequency include leukemia (10%), testicular cancer, and malignant melanoma (both less than 5%). In cancers such as breast and colon, TP53 mutations seem to occur late in

tumorigenesis. In several other cancers (head and neck, lung, skin), mutations occur very early and may even precede tumor development. The nature and type of mutations is often informative of the mutagenic mechanisms that have caused them, making TP53 an interesting gene to study in molecular epidemiology.

Mutations in the p53 protein can have at least three phenotypic effects: loss of function, in which a missense mutation abrogates p53's ability to block cell division or reverse a transformed phenotype; gain of function (or dominant-positive effect), where mutant p53 acquires novel functions as demonstrated with the introduction of a mutant p53 gene into cells lacking wild-type p53 allele, which induces a tumorigenic phenotype; trans-dominant mutation (dominant-negative effect), seen when a mutant p53 allele is introduced into cells bearing a wtp53 allele, resulting in the ability of mutant p53 to drive wtp53 to a mutant conformation overriding of the normal inhibitory function of p53.

3. The p53 Protein—A Sensor of Genotoxic Stress

In most cells, p53 is almost undetectable because it is rapidly degraded by the proteasome. Upon activation, the protein escapes degradation and accumulates in the nucleus. At the same time, it is turned from a latent to an active form by conformational changes which activate its capacity to transactivate target genes. The main factor controlling p53 accumulation is Mdm2, a protein encoded by a gene which is itself a transcriptional target of p53. Mdm2 acts as a ubiquitin ligase to direct p53 out of the nucleus to the proteasome, where it is degraded.

Various types of genotoxic and non-genotoxic stresses can lead to p53 activation, including agents that create single or double-strand breaks in DNA (irradiation, oxidative stress), mutagens— aflatoxins, benzo(a)pyrene, alkylating agents—and inhibitors of topoisomerases. Moreover, damage to the mitotic spindle, ribonucleotide depletion, hypoxia, heat shock, and exposure to nitric oxide can also induce p53. Induction follows a different time-course, depending upon the nature and intensity of the stress.

Induction in response to stress is a multi-step process. It involves phosphorylation of p53 in the N-terminus (e.g. by kinases activated after DNA-damage such as Atm or Chk-2), and dissociation of p53-Mdm2 interactions. Other changes in the protein include acetylation of the C-terminus (by acetyl-transferases of CBP/p300 family), conformational changes in the C-terminus leading to the unmasking of the DNA-binding domain, and changes in oxidation-reduction in the DNA-binding domain. All these changes turn the protein into an active form which binds DNA with high affinity.

Once activated, p53 can trigger several cellular events via two distinct and parallel pathways, transcription-dependent or transcription-independent (Figure 1). Examples of transcription-independent pathways include binding of p53 to components of the DNA replication/repair machinery such as the helicases ERCC2 and ERCC3, or the replication protein RPA. Genes transcriptionally regulated by p53 include cell cycle regulators in G1 and in G2 phases (p21/waf-1, 14-3-3s, GADD45), regulators of

apoptosis (BAX, CD95/FAS, KILLER/DR5, p53AIP1, PIG3, IGF-BP3), and genes involved in cellular responses to stress such as inducible forms of nitric oxide synthase (NOS₂) and cyclooxygenase (COX2), which are both repressed by p53. How p53 selects from the set of alternative responses (e.g. choosing between cell cycle arrest or apoptosis) depends upon the nature and the amplitude of the inducing signal, as well as of the cell and tissue type.

An important aspect of the role of p53 in cancer treatment is the fact that the function of p53 is crucial for the cytotoxic response of cancer cells to radio- or chemotherapy. There is evidence that many anti-cancer drugs induce apoptosis through a p53-dependent pathway. However, in clinical terms the presence of a wild-type TP53 gene is not always correlated with good response to treatment, as many other factors can also influence this response. On the other hand, in certain cell types activation of p53 by therapeutic agents may induce cell cycle arrest (and DNA repair) rather than apoptosis, thus resulting in a form of protection of cancer cells against the effects of therapy. Thus activation of p53 may be seen as both a chemo-sensitizer or a chemo-protective mechanism, depending on the cellular context. This is why current, experimental approaches that target p53 for cancer treatment include attempts to activate p53 (and thus induce apoptosis) as well as to inactivate p53 (and thus prevent destruction of normal cells by cytotoxic therapies).

4. Gene Therapy Using TP53

The capacity of wild-type TP53 to arrest the proliferation of cultured cells and induce apoptosis has raised an enormous interest in the possibility that restoring TP53 function in tumor cells may block tumor development. In addition, the finding that the p53 protein is a key factor in determining the response of cancer cells to therapy, has led to the concept that re-introduction of a normal protein may sensitize cells to cytotoxic killing and thus improve therapeutic response. Over the past ten years, several efforts have been made to translate these laboratory findings into clinical applications. One of the most popular approaches to achieve this goal is gene therapy. Below, we summarize the various modalities of TP53-based gene therapy that have been described in the recent literature.

4.1. Replacement Gene Therapy

The function of TP53 is lost in many cancers through mutation or loss of alleles. Therefore it seems reasonable to try to restore TP53 function by replacing the mutant gene with a functional, wild-type copy. The primary requirement to treat cancer with such replacement gene therapies is the necessity for highly efficient delivery of the wild-type TP53 into tumor cells *in vivo*. There must also be sufficient expression of functional p53 protein to mediate tumor suppression either through a direct mechanism involving cell death or growth arrest, or by increasing sensitivity to conventional anti-tumor agents. Other critical success factors include a low level of toxicity towards normal cells and the absence of a host immune response against the gene delivery system. The mechanisms of gene delivery can be subdivided in two broad categories: viral and non-viral.

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Bibliography

Abarzua P., Losardo E., Gubler M. L. and Nerla A. (1995). Microinjection of monoclonal antibody PAb421 into human SW480 colorectal carcinoma cells restores the transcription activation function to mutant p53. *Cancer Res.* **55**, 3490–3494. [Monoclonal antibody Pab421, when microinjected into human SW480 colorectal carcinoma cells, restores the transcriptional activity of the resident mutants p53 arg to his 273 and pro to ser 309.]

Agarwal M. L., Taylor W. R., Chernov M. V., Cjernova O. B. and Stark G. R. (1998). The p53 network. *J Biol Chem.* **273**, 1–4. [This paper focuses on the different aspects of the p53 network and regulation.]

Anker P., Mulcahy H., Chen X. Q. and Stroun M. (1999). Detection of circulating tumour DNA in the blood (plasma/serum) of cancer patients. *Cancer Metastasis Rev.* **18**, 65–73. [Small amounts of free DNA circulate in both healthy and diseased human plasma/serum, and increased concentrations of DNA are present in the plasma of cancer patients. The plasma DNA might be suitable for the development of non-invasive diagnosis, prognosis and follow up tests for cancer.]

Baas I. O., Hruban R. H. and Offerhaus G. J. (1999). Clinical applications of detecting dysfunctional p53 tumor suppressor protein. *Histol Histopathol.* **14**, 279–284. [This review summarizes the current understanding of the function of p53 as well as current methods to detect dysfunctional p53 and the clinical value of such analysis.]

Bergh J. (1999). Clinical studies of p53 in treatment and benefit of breast cancer patients. *Endocr Relat Cancer.* **6**, 51–59. [This article describes p53 as a prognostic and predictive factor, together with some information on how best to determine the p53 status.]

Bischoff J. R., Kirn D. H., Williams A., Heise C., Horn S., Muna M., Ng L., Nye J. A., Sampson-Johannes A., Fattaey A. and McCormick F. (1996). An adenovirus mutant that replicates selectively in p53-deficient human tumor cells. *Science* **274**, 373–376. [The authors show that a mutant adenovirus that does not express the 55-kilodalton viral protein encoded by E1B gene, can replicate in and lyse p53-deficient human tumor cells but not cells with functional p53.]

Bottger A., Bottger V., Sparks A., Liu W. L., Howard S. F. and Lane D. P. (1997). Design of a synthetic Mdm2-binding mini protein that activates the p53 response in vivo. *Curr Biol.* **7**, 860–869. [The authors designed a gene encoding a small protein that binds to the p53-binding pocket on the Mdm2 protein. Analysis of the results demonstrated that the p53 response is constitutively regulated in normal cells by Mdm2 and provides a powerful method to activate p53 without causing DNA damage.]

Cai D. W., Mukhopadhyay T., Liu Y., Fujiwara T. and Roth J. A. (1993). Stable expression of the wild-type p53 gene in human lung cancer cells after retrovirus-mediated gene transfer. *Hum Gene Ther.* **4**, 617–624. [A retroviral vector-mediated system was established to allow efficient transduction of the wild-type p53 gene into human lung cancer cell lines. Transduced cells could reduce the growth rate of non-transduced cells.]

Capizzi R. L. (1999). The preclinical basis for broad-spectrum selective cytoprotection of normal tissues from cytotoxic therapies by amifostine. *Semin Oncol.* **26**, 3–21. [The aminothiol amifostine, administered before cytotoxic chemotherapy or radiation, provides broad-spectrum cytoprotection of various normal tissues without attenuating antitumor response.]

- Capizzi R. L. (1999). Recent developments and emerging options: the role of amifostine as a broad-spectrum cytoprotective agent. *Semin Oncol.* **26**, 1–2. [The unique preclinical profile of amifostine can be the basis for the clinical development program for this new broad-spectrum cytoprotective agent.]
- Chen X., Bonnefoi H., Diebold-Berger S., Lyautey J., Lederrey C., Faltin-Traub E., Stroun M. and Anker P. (1999). Detecting tumor-related alterations in plasma or serum DNA of patients diagnosed with breast cancer. *Clin Cancer Res.* **5**, 2297–2303. [Small tumors of histoprognosis grade I of in situ carcinomas could present DNA alterations in the plasma/serum at an early stage. The wide increase of available microsatellite markers suggests that plasma or serum DNA may become a useful diagnostic tool for early and potentially curable breast cancer.]
- Cho Y., Gorina S., Jeffrey P. D. and Pavletich N. P. (1994). Crystal structure of a p53 tumor suppressor-DNA complex: understanding tumorigenic mutations. *Science* **265**, 346–355. [The crystal structure of a complex containing the core domain of human p53 and a DNA binding site has been determined.]
- Clayman G. L., El Naggar A. K., Lippman S. M., Henderson Y. C., Frederick M., Merritt J. A., Zumstein L. A., Timmons T. M., Liu T. J., Ginsberg L., Roth J. A., Hong W. K., Bruso P. and Goepfert H. (1998). Adenovirus-mediated p53 gene transfer in patients with advanced recurrent head and neck squamous cell carcinoma. *J Clin Oncol.* **16**, 2221–2232. [The authors evaluate the safety and the therapeutic potential of adenoviral (Ad-p53) in advanced head and neck squamous cell carcinoma (HNSSC).]
- Donehower L. A., Harvey M., Slagle B. L., McArthur M. J., Montgomery C. A., Jr., Butel J. S. and Bradley A. (1992). Mice deficient for p53 are developmentally normal but susceptible to spontaneous tumours. *Nature* **356**, 215–221. [The paper shows that a normal p53 gene is dispensable for embryonic development, that its absence predisposes the animal to neoplastic disease and that an oncogenic mutant form of p53 is not obligatory for the genesis of many types of tumors.]
- Dummer R., Bergh J., Karlsson Y., Horowitz J. A., Mulder N. H., Huinink D. T. B., Burg G., Hofbauer G. and Osanto S. (2000). Biological activity and safety of adenoviral vector-expressed wild-type p53 after intratumoral injection in melanoma and breast cancer patients with p53-overexpressing tumors. *Cancer Gene Ther.* **7**, 1069–1076. [A phase I dose escalation study of a replication-defective adenoviral expression vector containing wild-type p53 was carried out in patients with metastatic melanoma or breast cancer. The results demonstrate that p53 gene therapy is safe, feasible, and biologically effective in those patients.]
- Eastham J. A., Grafton W., Martin C. M. and Williams B. J. (2000). Suppression of primary tumor growth and the progression to metastasis with p53 adenovirus in human prostate cancer. *J Urol.* **164**, 814–819. [The authors investigate the ability of p53 adenovirus to suppress not only primary tumor growth but also the progression of metastatic disease. The results suggest that an adenovirus-p53 gene therapy strategy may be useful in the treatment of human prostate cancer.]
- Foster B. A., Coffey H. A., Morin M. J. and Rastinejad F. (1999). Pharmacological rescue of mutant p53 conformation and function. *Science* **286**, 2507–2510. [The authors identified compounds that stabilize the DNA binding domain of p53 in the active conformation. This class of compounds may be developed into anticancer drugs of broad utility.]
- Fujiwara T., Cai D. W., Georges R. N., Mukhopadhyay T., Grimm E. A. and Roth J. A. (1994). Therapeutic effect of a retroviral wild-type p53 expression vector in an orthotopic lung cancer model. *J Natl Cancer Inst.* **86**, 1458–1462. [The paper investigates the therapeutic efficacy of direct administration of a retroviral wild-type p53 expression vector in an orthotopic human lung cancer model.]
- Fujiwara T., Grimm E. A., Mukhopadhyay T., Cai D. W., Owen-Schaub L. B. and Roth J. A. (1993). A retroviral wild-type p53 expression vector penetrates human lung cancer spheroids and inhibits growth by inducing apoptosis. *Cancer Res.* **53**, 4129–4133. [The effects of retrovirus-mediated transduction of wild-type p53 were studied on multicellular tumor spheroids of human non-small cell lung cancer cell lines that have a mutant p53 protein. The results suggest that retroviral vectors can penetrate into multiple cell layers of three-dimensional tumor masses and induce potentially therapeutic effects.]
- Gaiddon C., Moorthy N. C. and Prives C. (1999). Ref-1 regulates the transactivation and pro-apoptotic functions of p53 in vivo. *EMBO J.* **18**, 5609–5621 [The paper addresses the physiological significance of the effects of the multifunctional protein Ref-1 on p53.]

Gallagher W. M. and Brown R. (1999). p53-oriented cancer therapies: current progress. *Ann Oncol.* **10**, 139–150. [This review focuses on the current progress in the development of p53-oriented cancer therapies: gene replacement therapy using wild type p53, restoration of p53 function by other means and targeting of the p53 dysfunction itself.]

Giuliano M., Catalano A., Strizzi L., Vianale G., Capogrossi M. and Procopio A. (2000). Adenovirus-mediated wild-type p53 overexpression reverts tumorigenicity of human mesothelioma cells. *Int J Mol Med.* **5**, 591–596. [The results reported in this paper indicate that p53-gene therapy should be strongly exploited for clinical trials in malignant mesothelioma patients.]

Greenblatt M. S., Bennett W. P., Hollstein M. and Harris C. C. (1994). Mutations in the p53 tumor suppressor gene: clues to cancer etiology and molecular pathogenesis. *Cancer Res.* **54**, 4855–4878. [This paper is an overview on the origin of p53 mutations, the mutational spectrum of p53 in human cancers and the hypotheses generated by the analysis of p53 mutations in premalignant and malignant cells.]

Habib N. A., Ding S. F., El Masry R., Mitry R. R., Honda K., Michail N. E., Dalla S. G., Izzi G., Greco L., Bassyouni M., El Toukhy M. and Abdel-Gaffar Y. (1996). Preliminary report: the short-term effects of direct p53 DNA injection in primary hepatocellular carcinomas. *Cancer Detect Prev.* **20**, 103–107. [This paper reports on a pilot study to assess the therapeutic potential of percutaneous injection of wild-type p53 in five patients with primary hepatocellular carcinoma.]

Hainaut P., Butcher S. and Milner J. (1995). Temperature sensitivity for conformation is an intrinsic property of wild-type p53. *Br J Cancer.* **71**, 227–231. [The results reported in this paper indicate that temperature sensitivity is an intrinsic property of wild-type p53 and suggests that small changes in temperature may directly affect p53 functions.]

Hainaut P. and Hollstein M. (2000). p53 and human cancer: the first ten thousand mutations. *Adv Cancer Res.* **77**, 81–137. [This review focuses on p53 biology and highlights the current understanding of the molecular biology and molecular epidemiology in human cancer.]

Hall A. R., Dix B. R., O'Carroll S. J. and Braithwaite A. W. (1998). p53-dependent cell death/apoptosis is required for a productive adenovirus infection. *Nat Med.* **4**, 1068–1072. [The authors show that cell expressing wild-type p53, as well as p53-deficient cells, allow adenovirus replication, but only cells expressing wild-type p53 show evidence of virus-induced cytopathic effect.]

Harris C. C. (1996). p53 tumor suppressor gene: from the basic research laboratory to the clinic--an abridged historical perspective. *Carcinogenesis* **17**, 1187–1198. [p53 is functionally inactivated by structural mutations, interaction with viral products, and endogenous cellular mechanisms in the majority of human cancers. Current research is defining the biochemical pathways through which p53 induces cell cycle arrest and apoptosis. Knowledge of these fundamental processes is leading to the identification of molecular targets toward which multimodality cancer therapies, using chemotherapeutic, immunotherapeutic, and gene-therapeutic strategies, can be based.]

Heise C., Sampson-Johannes A., Williams A., McCormick F., von Hoff D. D. and Kirn D. H. (1997). ONYX-015, an E1B gene-attenuated adenovirus, causes tumor-specific cytolysis and antitumoral efficacy that can be augmented by standard chemotherapeutic agents. *Nat Med.* **3**, 639–645. [This paper focuses on the effect of ONYX-015 (E1B gene-attenuated adenovirus) on human tumor cells. An antitumoral effect was documented following intratumoral or intravenous administration of ONYX-015 to nude mouse-human xenografts.]

Hernandez-Boussard T., Rodriguez-Tome P., Montesano R. and Hainaut P. (1999). IARC p53 mutation database: a relational database to compile and analyze p53 mutations in human tumors and cell lines. International Agency for Research on Cancer. *Hum Mutat.* **14**, 1–8. [The authors describe the structure and the format of the different databases on p53 mutations maintained at the International Agency for Research on Cancer (WHO) in Lyon.]

Hupp T. R. and Lane D. P. (1994). Allosteric activation of latent p53 tetramers. *Curr Biol.* **4**, 865–875. [P53 protein assembles naturally as a tetramer that can be converted between latent and activated forms by a concerted, allosteric transition. The system the authors have developed should facilitate the discovery of agents that can modulate the DNA-binding activity of p53 transition.]

Inoue A., Narumi K., Matsubara N., Sugawara S., Saijo Y., Satoh K. and Nukiwa T. (2000). Administration of wild-type p53 adenoviral vector synergistically enhances the cytotoxicity of anti-cancer

drugs in human lung cancer cells irrespective of the status of p53 gene. *Cancer Lett.* **157**, 105–112. [Recombinant adenovirus-mediated p53 gene transfer combined with anti-cancer drugs has clinical potential for gene therapy of lung cancer.]

Jayaraman J. and Prives C. (1995). Activation of p53 sequence-specific DNA binding by short single strands of DNA requires the p53 C-terminus. *Cell* **81**, 1021–1029. [The authors demonstrate that short single strand DNA can stimulate the ability of human and murine p53 protein to bind specifically to a p53 response element in supercoiled DNA. All the data taken together suggest a model in which the p53 C-terminus can recognize DNA structure resulting from damage-induced lesions.]

Kastan M. B., Onyekwere O., Sidransky D., Vogelstein B. and Craig R. W. (1991). Participation of p53 protein in the cellular response to DNA damage. *Cancer Res.* **51**, 6304–6311. [The paper suggests a role for the wild-type p53 protein in the inhibition of DNA synthesis that follows DNA damage and indicates a mechanism for the loss of wild-type p53 function that can contribute to tumorigenesis.]

Kastan M. B., Zhan Q., El-Deiry W. S., Carrier F., Jacks T., Walsh W. V., Plunkett B. S., Vogelstein B. and Fornace A. J. J. (1992). A mammalian cell cycle checkpoint pathway utilizing p53 and GADD45 is defective in ataxia-telangiectasia. *Cell* **71**, 587–597. [The authors identify three genes implicated in the signal transduction pathway that controls cell cycle arrest following DNA damage: AT gene, p53 and GADD45.]

Khuri F. R., Nemunaitis J., Ganly I., Arseneau J., Tannock I. F., Romel L., Gorem M., Ironside J., MacDougall R. H., Heise C., Randlev B., Gillenwater A. M., Brusio P., Kaye S. B., Hong W. K. and Kimm D. H. (2000). A controlled trial of intratumoral ONYX-015, a selectively-replicating adenovirus, in combination with cisplatin and 5-fluorouracil in patients with recurrent head and neck cancer. *Nat Med.* **6**, 879–885. [This paper reports a Phase II clinical trial of a combination of intratumoral injection of engineered adenovirus ONYX-015 with cisplatin and 5-fluorouracil in patients with recurrent squamous cell cancer of the head and neck. The results showed tumor-selective viral replication and necrosis induction.]

Kim A. L., Raffo A. J., Brandt-Rauf P. W., Pincus M. R., Monaco R., Abarzua P. and Fine R. L. (1999). Conformational and molecular basis for induction of apoptosis by a p53 C-terminal peptide in human cancer cells. *J Biol Chem.* **274**, 34924–34931. [A p53-derived C-terminal peptide induced rapid apoptosis in breast cancer cell lines carrying endogenous p53 mutations or over-expressing wild-type p53 but was not toxic to non-malignant human cell lines containing wild-type p53.]

Kirk G. D., Camus-Randon A. M., Mendy M., Goedert J. J., Merle P., Trepo C., Brechot C., Hainaut P. and Montesano R. (2000). Ser-249 p53 mutations in plasma DNA of patients with hepatocellular carcinoma from The Gambia. *J Natl Cancer Inst.* **92**, 148–153. [The ser-249 p53 mutation in plasma DNA is strongly associated with hepatocellular carcinoma in Gambian patients. Use of ser-249 p53 mutation should facilitate further molecular epidemiological studies on the development of hepatocellular carcinoma.]

Ko L. J. and Prives C. (1996). p53: puzzle and paradigm. *Genes Dev.* **10**, 1054–1072. [This review presents some of the recent advances in p53 research and discusses how they either shed light on or add to the complexities of p53.]

Komarov P. G., Komarova E. A., Kondratov R. V., Christov-Tselkov K., Coon J. S., Chernov M. V. and Gudkov A. V. (1999). A chemical inhibitor of p53 that protects mice from the side effects of cancer therapy. *Science* **285**, 1733–1737. [Pifithrin-alpha is a small molecule isolated for its ability to reversibly block p53-dependent transcriptional activation and apoptosis. Inhibitors of p53 can thus be useful drugs for reducing the side effects of cancer therapy and other types of stress associated with p53 induction.]

Komarova E. A. and Gudkov A. V. (2000). Suppression of p53: a new approach to overcome side effects of antitumor therapy. *Biochemistry (Mosc.)* **65**, 41–48. [The authors supposed that a temporary suppression of p53 can decrease the damage to sensitive tissues and accelerate their recovery after the antitumor radio- and chemotherapy. They isolated a chemical inhibitor of p53, called pifithrin-alpha, and determined its activity in vitro and in vivo. The results show that the suppression of p53 is a promising approach in the prevention of side effects of antitumor therapy.]

Kramer D. L., Vujcic S., Diegelman P., Alderfer J., Miller J. T., Black J. D., Bergeron R. J. and Porter C. W. (1999). Polyamine analogue induction of the p53-p21WAF1/CIP1-Rb pathway and G1 arrest in human melanoma cells. *Cancer Res* **59**, 1278–1286. [This paper provides the first indication of the cell

cycle regulatory pathways by which polyamine antagonists such as analogues might inhibit growth in cells containing wild-type p53 and further suggests a mechanistic basis for differential cellular responses to these agents.]

Kurbacher C. M. and Mallmann P. K. (1998). Chemoprotection in anticancer therapy: the emerging role of amifostine (WR-2721). *Anticancer Res.* **18**, 2203–2210. [The aminothiols amifostine offers a rational approach to protect patients against chemotherapy-specific and often dose-limiting effects and is thus likely to improve therapeutic outcome significantly.]

Laken S. J., Jackson P. E., Kinzler K. W., Vogelstein B., Strickland P. T., Groopman J. D. and Friesen M. D. (1998). Genotyping by mass spectrometric analysis of short DNA fragments. *Nat Biotechnol.* **16**, 1352–1356. [In this paper the authors describe a method developed to produce small DNA fragments from PCR products for analysis of defined DNA variations by mass spectrometry.]

Lane D. P. (1992). Cancer. p53, guardian of the genome. *Nature* **358**, 15–16. [This paper focuses on the role of p53 in the control of different aspects of cell life for the maintenance of the integrity of the genome.]

Lane D. P. (1999). Exploiting the p53 pathway for cancer diagnosis and therapy. *Br J Cancer*, **80** Suppl 1, 1–5. [This paper reviews the different pathways in which p53 is involved in order to suggest different therapeutic approaches for cancer diagnosis and treatments.]

Lane D. P. and Hall P. A. (1997). MDM2--arbiter of p53's destruction. *Trends Biochem Sci.* **22**, 372–374. [This review highlights the role of Mdm2 oncoprotein in the control of p53. Mdm2 is a potent inhibitor of p53 but is also induced by p53 in an autoregulatory feedback loop. The Mdm2-promoted degradation of p53 is a mechanism to ensure effective termination of the p53 signal.]

Lang F. F., Yung W. K., Sawaya R. and Tofilon P. J. (1999). Adenovirus-mediated p53 gene therapy for human gliomas. *Neurosurgery* **45**, 1093–1104. [Preclinical data reported in this paper strongly support the use of p53 gene transfer as a potential treatment for human gliomas.]

Lesoon-Wood L. A., Kim W. H., Kleinman H. K., Weintraub B. D. and Mixson A. J. (1995). Systemic gene therapy with p53 reduces growth and metastases of a malignant human breast cancer in nude mice. *Hum Gene Ther* **6**, 395–405 [The authors report an in vivo delivery system (liposome-p53 complex) that attenuates the growth, in nude mice, of a malignant human breast cancer cell line containing p53 mutation.]

Levine A. J. (1997). p53, the cellular gatekeeper for growth and division. *Cell* **88**, 323–331. [This review focuses on the cellular and molecular biology of p53 in cell growth and division.]

Li L., Li J., Rao J. N., Li M., Bass B. L. and Wang J. Y. (1999). Inhibition of polyamine synthesis induces p53 gene expression but not apoptosis. *Am J Physiol.* **276**, C946–C954. [The present study examines the effect of decreasing cellular polyamines on p53 expression and apoptosis in small intestinal epithelial cells (IEC-6). The results suggest that increased expression of the p53 gene may play an important role in growth inhibition caused by polyamine depletion.]

Lowe S. W. (1995). Cancer therapy and p53. *Curr Opin. Oncol.* **7**, 547–553. [This review focuses on the observations that inactivation of p53 promotes resistance to anticancer agents by attenuating apoptosis. This view identifies p53 as a potential drug target and suggests several strategies for therapeutic intervention.]

Lowe S. W., Schmitt E. M., Smith S. W., Osborne B. A. and Jacks T. (1993). p53 is required for radiation-induced apoptosis in mouse thymocytes. *Nature* **362**, 847–849. [This paper illustrates how p53 is required for radiation-induced cell death in the thymus but is not necessary for all forms of apoptosis.]

Maurici D., Monti P., Campomenosi P., North S., Frebourg T., Fronza G. and Hainaut P. (2001). Amifostine (WR2721) restores transcriptional activity of specific p53 mutant proteins in a yeast functional assay. *Oncogene*. [This paper illustrates the ability of the chemo/radio-protector Amifostine to restore the wild-type transcriptional activity in some p53 mutants in a yeast-based functional assay.]

Méplán C., Richard M. J. and Hainaut P. (2000). Redox signaling and transition metals in the control of the p53 pathway. *Biochem Pharmacol.* **59**, 25–33. [In this review the authors summarize the evidence for the involvement of reactive oxygen intermediates (ROI) in p53 pathway. Emphasis is placed on the role

of metals and ROI as potential regulators of p53 protein conformation and functions and on the putative toxicological consequences of such regulation.]

Milas M., Yu D., Lang A., Ge T., Feig B., El Naggar A. K. and Pollock R. E. (2000). Adenovirus-mediated p53 gene therapy inhibits human sarcoma tumorigenicity. *Cancer Gene Ther.* **7**, 422–429. [The authors constructed a replication deficient adenovirus expressing wild-type p53 (Ad5p53) with a Flag sequence tag and they transduced it into sarcoma cells containing mutant p53. The results indicate that wild-type p53 gene restoration in sarcoma retards tumor growth.]

Miyake H., Hara I., Haras S., Arakawa S. and Kamidono S. (2000). Synergistic chemosensitization and inhibition of tumor growth and metastasis by adenovirus-mediated P53 gene transfer in human bladder cancer model. *Urology* **56**, 332–336. [The paper shows that combined treatment with replication deficient adenovirus Ad5p53 and cisplatin could be an attractive strategy for inhibiting progression of bladder cancer through effective induction of apoptosis.]

North S., El Ghissassi F., Pluquet O., Verhaegh G. and Hainaut P. (2000). The cytoprotective aminothioliol WR1065 activates p21waf-1 and down regulates cell cycle progression through a p53-dependent pathway. *Oncogene* **19**, 1206–1214. [The data reported in this paper indicate that the active metabolite (WR1065) of the prodrug amifostine, a well-known cytoprotective agent, activates a cell cycle checkpoint involving p53.]

North S. and Hainaut P. (2000). p53 and cell cycle control: a finger in every pie. *Pathol Biol (Paris)* **48**, 255–270. [This review summarizes the data available on the direct involvement of p53 in the control of cell cycle progression in G1, S, G2 and M phases and discusses the way in which cell cycle regulation may contribute to control p53 protein levels and activity during the normal cell cycle.]

Oren M. (1992). p53: the ultimate tumor suppressor gene? *FASEB J* **6**, 3169–3176. [Despite the central role of p53 in the control of cell proliferation, cell survival and differentiation, p53 mice can develop apparently without any defect in the total absence of p53. The paper investigates the possibility that p53 may become critically limiting only when normal growth control is lost.]

Oren M. (1999). Regulation of the p53 tumor suppressor protein. *J Biol Chem.* **274**, 36031–36034. [This review focuses on the signals and the mechanisms that regulate p53 activity, maintaining it at low level under normal conditions and turning it on in cancer-prone cells.]

Osaki S., Nakanishi Y., Takayama K., Pei X. H., Ueno H. and Hara N. (2000). Alteration of drug chemosensitivity caused by the adenovirus-mediated transfer of the wild-type p53 gene in human lung cancer cells. *Cancer Gene Ther.* **7**, 300–307. [The authors investigate the effect of wild-type p53 transfer in combination with 12 anticancer agents on human pulmonary squamous cell carcinoma cell line. The results indicate that irinotecan (CPT-11) as well as cisplatin would be a candidate for the combination of chemotherapy and gene therapy for non-small cell lung cancer.]

Ramondetta L., Mills G. B., Burke T. W. and Wolf J. K. (2000). Adenovirus-mediated expression of p53 or p21 in a papillary serous endometrial carcinoma cell line (SPEC-2) results in both growth inhibition and apoptotic cell death: potential application of gene therapy to endometrial cancer. *Clin Cancer Res.* **6**, 278–284. [The authors compare the efficacy of adenoviral vector containing p53 (Adp53) or p21 in a papillary serous endometrial tumor cell line (SPEC-2) that contains mutated p53. These preclinical studies suggest that papillary serous endometrial carcinoma is a potent target for p53 or p21-mediated gene therapy.]

Ray R. M., Zimmerman B. J., McCormack S. A., Patel T. B. and Johnson L. R. (1999). Polyamine depletion arrests cell cycle and induces inhibitors p21(Waf1/Cip1), p27(Kip1), and p53 in IEC-6 cells. *Am J Physiol.* **276**, C684–C691. [The paper shows that polyamine depletion causes cell cycle arrest and upregulates cell cycle inhibitors, and suggests that MAPK and JNK kinases may be involved in the regulation of the activity of these molecules.]

Ries S. J., Brandts C. H., Chung A. S., Biederer C. H., Hann B. C., Lipner E. M., McCormick F. and Michael K. W. (2000). Loss of p14ARF in tumor cells facilitates replication of the adenovirus mutant dl1520 (ONYX-015). *Nat Med.* **6**, 1128–1133. [The authors identify loss of p14ARF as a mechanism that allows adenovirus ONYX-015 replication in tumor cells retaining wild-type p53. They demonstrate that the re-introduction of p14ARF into tumor cells with wild-type p53 suppress replication of ONYX-015 in a p53-dependent manner.]

Roth J. A., Nguyen D., Lawrence D. D., Kemp B. L., Carrasco C. H., Ferson D. Z., Hong W. K., Komaki R., Lee J. J., Nesbitt J. C., Pisters K. M., Putnam J. B., Schea R., Shin D. M., Walsh G. L., Dolormente M. M., Han C. I., Martin F. D., Yen N., Xu K., Stephens L. C., McDonnell T. J., Mukhopadhyay T. and Cai D. (1996). Retrovirus-mediated wild-type p53 gene transfer to tumors of patients with lung cancer. *Nat Med.* **2**, 985–991. [A retroviral vector containing the wild-type p53 gene under control of a beta actin promoter was produced to mediate transfer of wild-type p53 into human non-small cell lung cancers by direct injection.]

Roth J. A., Swisher S. G. and Meyn R. E. (1999). p53 tumor suppressor gene therapy for cancer. *Oncology (Huntingt.)* **13**, 148–154. [This review describes gene therapy strategies under investigation, summarizes preclinical data that provides a rationale for the gene replacement approach, and discusses the clinical trial data available to date.]

Runnebaum I. B., Wang S. and Kreienberg R. (1995). Retrovirally mediated wild-type p53 restores S-phase modulation without inducing WAF1 mRNA in breast carcinoma cells containing mutant p53. *J Cell Biochem.* **59**, 537–544. [The authors describe the use of retrovirally transduced wild-type p53 in breast cancer cell lines containing homozygously endogenous mutant p53.]

Selivanova G., Iotsova V., Okan I., Fritsche M., Strom M., Groner B., Grafstrom R.C. and Wiman K.G. (1997). Restoration of the growth suppression function of mutant p53 by a synthetic peptide derived from the p53 C-terminal domain. *Nat. Med.* **3**, 632–638. [The results reported in this paper show that a synthetic peptide, corresponding to the carboxy-terminal aminoacid residues 361–382 of p53 can activate specific DNA binding of wild-type p53 in vitro and can restore the transcriptional transactivating function of at least some mutant p53 proteins in living cells.]

Shinoura N., Yoshida Y., Asai A., Kirino T. and Hamada H. (2000). Adenovirus-mediated transfer of p53 and Fas ligand drastically enhances apoptosis in gliomas. *Cancer Gene Ther.* **7**, 732–738. [The authors attempt to develop a technique that overrides the resistance toward apoptosis in glioma cells. The results indicate that the coinfection approach with adenovirus for p53 and Fas ligand can be used as a modality for gene therapy of glioma.]

Soussi T. (2000). The p53 tumor suppressor gene: from molecular biology to clinical investigation. *Ann N Y Acad Sci.* **910**, 121–137. [This review focuses on the biological and molecular characteristic of p53 and reports the most important p53 mutations found in human tumors.]

Soussi T. (2000). p53 Antibodies in the sera of patients with various types of cancer: a review. *Cancer Res.* **60**, 1777–1788. [This review compiles more than 130 papers published in the specific field of p53 antibodies since 1992.]

Swiaher S. G., Roth J. A., Nemunaitis J., Lawrence D. D., Kemp B. L., Carrasco C. H., Connors D. G., El Naggar A. K., Fossella F., Glisson B. S., Hong W. K., Khuri F. R., Kurie J. M., Lee J. J., Lee J. S., Mack M., Merritt J. A., Nguyen D. M., Nesbitt J. C., Perez-Soler R., Pisters K. M., Putnam J. B., Jr., Richli W. R., Savin M. and Waugh M. K. (1999). Adenovirus-mediated p53 gene transfer in advanced non-small-cell lung cancer. *J Natl Cancer Inst.* **91**, 763–771. [This paper reports a phase I clinical trial with adenoviral vector containing wild-type p53 in patients with non-small-cell lung cancer that had progressed on conventional treatments.]

Tokino T. and Nakamura Y. (2000). The role of p53-target genes in human cancer. *Crit Rev Oncol Hematol.* **33**, 1–6. [This review focuses on the biological functions of the transcriptional targets of p53.]

Ueno M., Masutani H., Arai R. J., Yamauchi A., Hirota K., Sakai T., Inamoto T., Yamaoka Y., Yodoi J. and Nikaïdo T. (1999). Thioredoxin-dependent redox regulation of p53-mediated p21 activation. *J Biol Chem.* **274**, 35809–35815. [Thioredoxin (TRX) is an enzyme induced by various oxidative stresses. This paper investigates the role of TRX in the regulation of p53 activity.]

Winter S. F., Minna J. D., Johnson B. E., Takahashi T., Gazdar A. F. and Carbone D. P. (1992). Development of antibodies against p53 in lung cancer patients appears to be dependent on the type of p53 mutation. *Cancer Res.* **52**, 4168–4174. [The authors found that the ability of lung cancer patients to develop anti-p53 antibodies is correlated with the type of p53 mutation. Many patients have tumors with missense mutations and do not develop anti-p53 antibodies.]

Zalcman G., Schlichtholz B., Tredaniel J., Urban T., Lubin R., Dubois I., Milleron B., Hirsch A. and Soussi T. (1998). Monitoring of p53 autoantibodies in lung cancer during therapy: relationship to

response to treatment. *Clin Cancer Res.* **4**, 1359–1366. [The authors find a good correlation between the specific evolution of the p53 antibody titer and the response to therapy suggesting that p53 antibodies could represent a useful tool for checking the response to therapy and for monitoring some relapses before they are clinically detectable.]

Zeimet A. G., Riha K., Berger J., Widschwendter M., Hermann M., Daxenbichler G. and Marth C. (2000). New insights into p53 regulation and gene therapy for cancer. *Biochem Pharmacol.* **60**, 1153–1163. [This paper focuses on combinations of p53-based gene therapy with other components involving apoptosis or agents neutralizing tumor-promoting antiapoptotic signals. Those combinations should further improve the effectiveness of cancer treatment.]

Biographical Sketches

Daniela Maurici was born in Bologna, Italy, in 1965. She trained as a cell and molecular biologist under the supervision of Dr N. Baldini at the University of Bologna. After her PhD in 1997, she joined the Group of Molecular Carcinogenesis at the International Agency for Research on Cancer, where she worked on the development of model systems to analyze the effect of drugs modulating p53 protein conformation and activities. She is currently a research fellow at the Joint Research Center, European Union, Ispra, Italy.

Pierre Hainaut was born in Liège, Belgium, in 1958. He is a molecular biologist and is the leader of the Group of Molecular Carcinogenesis at the International Agency for Research on Cancer (WHO). He started to work on TP53 as a postdoctoral fellow in the group of Dr J. Milner, in Cambridge, in 1990. His research addresses the role of metals and redox factors in the control of p53 protein functions. This interest has led him to develop projects aimed at identifying agents that modulate p53 protein activities for therapeutic or preventive uses. Within his activities at WHO, he is also responsible for several molecular epidemiological studies on esophageal cancers and on liver cancers, in which TP53 is an important biomarker. He also supervises the IARC TP53 mutation database, a central resource compiling and annotating all TP53 mutations reported in the world literature.