

SURFACE PHENOMENA

Narsimhan, Ganesan

Biochemical and Food Process Engineering, Department of Agricultural and Biological Engineering, Purdue University, USA

Keywords: Protein adsorption, food emulsifier, food emulsions and foams, flocculation, coalescence, adsorption isotherm of proteins at fluid interfaces, kinetics of adsorption of proteins at fluid interfaces, interfacial rheology, competitive adsorption of emulsifier-protein mixtures, hydrophile-lipophile balance.

Contents

1. Introduction
 2. Colloidal Forces Between Particles
 - 2.1. Van der Waals Interaction
 - 2.2. Electrostatic Interactions
 - 2.3. Steric Interaction
 - 2.3.1. Forces Due to Adsorbed Macromolecules
 - 2.3.2. Forces Due to Free Macromolecules
 - 2.4. Combination of Forces
 3. Flocculation
 - 3.1. Brownian Flocculation
 - 3.2. Gravity-Induced Flocculation
 - 3.3. Coalescence
 4. Food Emulsifiers
 - 4.1. Small Molecular Weight Surfactants
 - 4.2. Hydrophile-Lipophile Balance
 - 4.3. Proteins
 - 4.4. Adsorption Isotherm
 - 4.5. Kinetics of Adsorption
 5. Competitive Adsorption in Mixed Emulsifier Systems
 6. Interfacial Rheological Properties of Adsorbed Layer
- Glossary
Bibliography
Biographical Sketch

Summary

Many food systems are dispersed systems in which one phase is dispersed into a second immiscible phase in the form of fine particles or droplets. When the two phases are liquids, the product is an emulsion. When one phase is air, the product is foam, in which the air bubbles are separated by thin aqueous films. Because of the large free energy associated with the large interfacial area, such systems are thermodynamically unstable. The shelf life of such products are prolonged by providing kinetic stability through the use of proteins and emulsifiers that modify the surface, as well as interaction forces between dispersed particles through the formation of an adsorbed interfacial layer. The evaluation of colloidal van der Waals, electrostatic, and steric interactions is discussed.

Calculation of the rate of flocculation of dispersed particles due to Brownian and gravitational motion is presented. Characterization of surface activity of small molecular weight surfactants via hydrophile-lipophile balance and the Gibbs adsorption equation is discussed. Lattice models for prediction of the adsorption isotherm of proteins at fluid-fluid interfaces are presented and compared with experimental data for some model protein systems. Energy barriers for adsorption of proteins at fluid-fluid interfaces are discussed. Models for the kinetics of protein adsorption accounting for these energy barriers in terms of molecular properties of proteins are presented and their predictions compared with experimental data. Finally, competitive adsorption of mixed emulsifier-protein systems at fluid-fluid interfaces and its effect on interfacial rheological properties are discussed.

1. Introduction

In many food products such as mayonnaise, whipped cream butter, and sauces, one phase (air, water, oil, or solid particles) is dispersed in the other phase. When the two phases are liquids, the product is called an emulsion, which is further sub-classified as oil-in-water or water-in-oil emulsion, depending on the nature of the dispersed phase (see *Food Emulsions*). Milk and mayonnaise are examples of oil-in-water emulsions, whereas butter is a water-in-oil emulsion. When the dispersed phase is air, the product is called foam. Ice cream and whipped cream are good examples of foam. When the dispersed phase is solid particles, the product is known as a dispersion (see *Food Suspensions*). Different types of sauces are examples of dispersions. Of course, many of these products are indeed combinations of emulsions, foams, or dispersions. Since one phase is dispersed in the form of very fine particles over a large interfacial area, such dispersions are thermodynamically unstable, i.e., eventually the system will phase separate. Phase separation occurs as a result of flocculation, followed by coalescence of colliding particles (see *Separation*). Particle collision in such colloidal systems is brought about by either thermal motion or gravitational motion caused by the density difference between the dispersed and continuous phases. The gravitational motion leads to creaming, thus resulting in the formation of a cream layer in the case of emulsion, which subsequently leads to coalescence and phase separation. In order to provide kinetic stability to these colloidal systems, with desirable shelf life, food emulsifiers are added. Food emulsifiers can be either small molecular weight surfactants such as monoglyceride or macromolecules such as proteins. By virtue of their surface activity, food emulsifiers adsorb at air-water, oil-water, or solid-water interfaces. Enhancement of shelf life is brought about by several mechanisms, namely, 1) the modification of interaction between dispersed particles by providing repulsive interactions, thereby preventing or slowing down flocculation or coalescence of colliding particles and 2) the modification of interfacial rheological properties in the case of emulsions and foams, thereby slowing down the drainage of the continuous phase between two colliding dispersed particles. In addition to providing shelf life, food emulsifiers also facilitate the formation of foams and emulsions of sufficiently small particles by reducing the surface/interfacial tensions due to their surface activity. Of course, the efficacy of food emulsifiers will depend on their surface activity. This chapter briefly describes the interaction between dispersed particles, the interfacial behavior of food emulsifiers and proteins, and their effect on the long-term stability of dispersed food systems.

2. Colloidal Forces Between Particles

As pointed out above, an important factor affecting the stability of a colloidal dispersion is the nature of the interparticle forces. The dispersed particles always experience van der Waals attractive forces. In addition, adsorption of proteins and ionic surfactant will impart charge to the colloidal particles, thereby introducing electrostatic repulsive forces between them. The adsorption of proteins will also introduce steric interactions. To better understand the role of food emulsifiers on colloidal stability, it is important to quantify these interactions.

2.1. Van der Waals Interaction

Van der Waals interaction is the result of permanent dipole-permanent dipole, permanent dipole-induced dipole, and induced dipole-induced dipole interactions between molecules. Of these, induced dipole-induced dipole interactions, known as dispersion forces, are experienced by all molecules and have their origin in the rapidly fluctuating dipole moment of a neutral molecule, which polarizes other neighboring molecules, thus giving rise to attractive interactions. Assuming pairwise additivity, the van der Waals interactions U_{vw} between two spherical particles for distances of separation much smaller than the particle radii is given by:

$$U_{vw} = -\frac{AR_1R_2}{6h(R_1 + R_2)} \quad R_1, R_2 \gg h \quad (1)$$

where R_1 and R_2 are the radii of both particles, A is the effective hamaker constant, which depends on the polarizabilities, and h is the surface to surface distance between the two particles. Van der Waals attraction is a fairly, long-range interaction, with the attractive force increasing significantly at shorter distances of separation. When the distance of separation between the particles is on the order of wavelength of electromagnetic radiation (typically in the order of 100 nm), dipoles shift during the propagation, causing retardation effects, which diminish the attraction and therefore cannot be neglected. Simplification of expressions for the retardation can be found elsewhere.

For particles (2) dispersed in a continuous medium (1), the effective Hamaker constant A_{212} is given by:

$$A_{212} = (A_{11}^{1/2} - A_{22}^{1/2})^2 \quad (2)$$

The effective hamaker constant is very sensitive to the polarizability of the medium. The net force is always attractive, becoming zero only when the dispersed and the continuous phases have the same polarizability. Generally, the adsorbed layer of proteins tends to decrease the net van der Waals attraction, the extent of reduction being dependent on the nature and the thickness of the adsorbed layer.

2.2. Electrostatic Interactions

Adsorption of ionic proteins and surfactants imparts charge to the colloidal particles. Electroneutrality requires that the net particle charge be counterbalanced by preferential accumulation of oppositely-charged counterions in the vicinity of the charged surface. Because of thermal motion, the neutralization occurs over a region known as the electrical double layer. The double layer consists of an inner compact stern layer and an outer diffuse layer. The electrostatic potential profile within an electrical double layer can be obtained by solving the Poisson-Boltzmann equation. The thickness of the electrical double layer depends on the ionic strength of the continuous medium, the valence number of the counterion, and the dielectric constant of the medium, given by:

$$\kappa = \left(\frac{2000e^2 N_A I}{\epsilon_0 \epsilon_r kT} \right)^{1/2} \quad (3)$$

where κ , the Debye-Huckel parameter, is inversely proportional to the double layer thickness, ϵ_0 is the permittivity of vacuum, ϵ_r the dielectric constant of the medium, e the elementary charge, N_A the Avagadro number, k the Boltzmann constant, T the temperature, and the ionic strength I is given by:

$$I = \frac{1}{2} \sum_{i=1,2} z_i^2 M_i \quad (4)$$

where z_i is the valence number of i^{th} type of ion and M_i is the molar concentration of i^{th} type of ion in the bulk. The double layer thickness decreases with increase in the ionic strength as well as with the valence number of the counterion.

When two colloidal particles approach each other, they experience a force of repulsion because of the overlap of double layers that surrounds each particle. This repulsion is due to the difference in the electrostatic potential profile in the overlap region compared to an isolated double layer, thus resulting in a net repulsive interaction. The interaction can be calculated from the electrostatic potential profile obtained from the solution of the Poisson Boltzmann equation in the region between the two particles. The relative motion between the two colliding particles is relatively slow, thus the surface potential of the approaching particles is usually assumed as constant. When the double layer around each particle is sufficiently thin, (i.e., κR is sufficiently large), Derjaguin's procedure can be employed to evaluate the interaction between two spherical particles using the results of interaction between two charged plates. For low surface potentials, the interaction potential between two equal-sized particles of radius R is given by:

$$U_{el} = 2\pi\epsilon_0\epsilon_r R\psi_0^2 \ln\{1 + \exp(-\kappa h)\} \quad (5)$$

where ψ_0 is the surface potential of the particle. When the surface potential is not low, the electrostatic interaction potential can be evaluated for negligible overlap of the two double layers, yielding:

$$U_{el} = \frac{64n_0kT\gamma_0^2}{\kappa} \exp(-\kappa h) \quad (6)$$

where n_0 is the bulk number concentration of electrolyte and γ_0 is related to the surface potential via

$$\gamma_0 = \frac{\exp\left(\frac{ze\psi_0}{2kT}\right) - 1}{\exp\left(\frac{ze\psi_0}{2kT}\right) + 1} \quad (7)$$

The surface potential ψ_0 is usually taken to be the zeta potential, the potential at the plane of shear.

2.3. Steric Interaction

2.3.1. Forces Due to Adsorbed Macromolecules

When proteins are employed to provide stability to emulsions, foams and colloidal dispersions, they adsorb at the interface and form a thick, adsorbed layer. The adsorption characteristics of proteins at the interfaces will be discussed in detail later in the following sections. When two colloidal particles with adsorbed protein layers approach each other, the overlap of the adsorbed layer usually results in a steric repulsive interaction between the two particles. If the surface-to-surface distance of separation between the two particles is greater than twice the thickness of the adsorbed layer, the two particles will not experience any steric interaction, since the adsorbed layers do not overlap. If the surface-to-surface distance of separation is between one and two spans of the adsorbed layer, i.e., $L_s \leq h \leq 2L_s$, L_s is the thickness of the adsorbed layer, the adsorbed segments will overlap but will not be compressed. In this regime, the steric interaction between the two particles is due to the difference in the free energy of mixing between the adsorbed segments and the solvent in the overlap region, because of the increase in the segment density. Whenever the interaction between the segments and the solvent is favorable, i.e., the solvent is good, the overlap of the adsorbed layers results in an increase of free energy, which manifests itself as a steric repulsion. Proteins generally have a favorable interaction with water (solvent). As a result, steric repulsion is usually experienced by the colloidal particles stabilized by proteins. For uniform segment density of proteins in the adsorbed layer, the steric interaction U_{steric} between two spherical particles of radii R is given by:

$$U_{steric} = 4\pi R\omega^2 N_A \left(\frac{\bar{v}_2^2}{\bar{v}_1}\right) \left(\frac{1}{2} - \chi\right) kT \left(1 - \frac{h}{2L_s}\right)^2 \quad (8)$$

where ω is the weight of the adsorbed segments per unit area, χ is the Flory Huggins parameter, \bar{v}_1 is the molar volume of the solvent, and \bar{v}_2 is the partial specific volume

of protein. Since the interaction between protein and the solvent is usually favorable, $\chi < \frac{1}{2}$. As a result, $U_{steric} > 0$, i.e., steric interaction is repulsive.

When the minimum surface-to-surface distance of separation between two particles is less than the thickness of the adsorbed layer, (i.e., $h \leq L_s$), loops and tails of adsorbed protein segments are compressed by the other particle. This compression always results in a repulsive interaction irrespective of the nature of the solvent, since loss of configurations due to compression leads to a decrease in entropy and therefore an increase in the free energy. For constant segment density in the adsorbed layer, the steric interaction is given by:

$$U_{steric} = 4\pi R\omega^2 N_A \left(\frac{\bar{v}_2^2}{\bar{v}_1} \right) \left(\frac{1}{2} - \chi \right) kT \left\{ \ln \left(\frac{L_s}{h} \right) - \frac{1}{4} + \frac{h}{2L_s} \right\} + 2\pi RkTvL_s \left(1 - \frac{h}{L_s} \right)^2 \quad (9)$$

where ν is the number of adsorbed segments per unit area. It should be noted that the second term is the elastic contribution to the interaction energy, which arises from the compression of the adsorbed segments.

2.3.2. Forces Due to Free Macromolecules

Water soluble polymers such as gums, pectins, and gelatin can cause flocculation when present at very low concentrations. When the distance of separation between two colloidal particles becomes comparable to the radius of gyration of the water soluble polymer, the free energy of the macromolecule in the region between the two particles will increase because of geometric constraints. As a result, the two particles experience a repulsive interaction. If the two particles are able to overcome this energy barrier and come closer, then the water soluble polymer is completely excluded in the region between the particles, since the distance of separation is now much smaller than the radius of gyration. Consequently, the osmotic pressure of the polymer solution does not act on the particles in the region between them, thus resulting in an attraction between particles. The attractive interaction between two particles is given by:

$$U_{steric} = -\pi_{osmotic} (\pi/12) \{ 2(2R + R_g)^3 - 3(2R + R_g)^2(2R + h) + (2R + h)^3 \} \quad (10)$$

where the osmotic pressure $\pi_{osmotic}$ can be related to the polymer number concentration n_{pol} via:

$$\pi_{osmotic} = kTn_{pol} \quad (11)$$

2.4. Combination of Forces

The combination of different interparticle forces discussed above determines the stability of colloidal dispersion to flocculation. The dispersion can be classified as 1) unstable, 2) metastable, and 3) stable. In an unstable dispersion, the net interaction is attractive except at very short distances of separation with the net interaction potential exhibiting a potential well where flocculation can occur. This happens when the repulsion is either small or short ranged. In a metastable dispersion, van der Waals attraction is dominant at very small and large distances, whereas repulsion is predominant at intermediate distances of separation, resulting in an energy barrier as shown in Figure 1. This is usually the case for electrostatically stabilized systems. In a stable system, repulsion predominates over van der Waals attraction for the entire range of separation distance. This is typical of a sterically stabilized system.

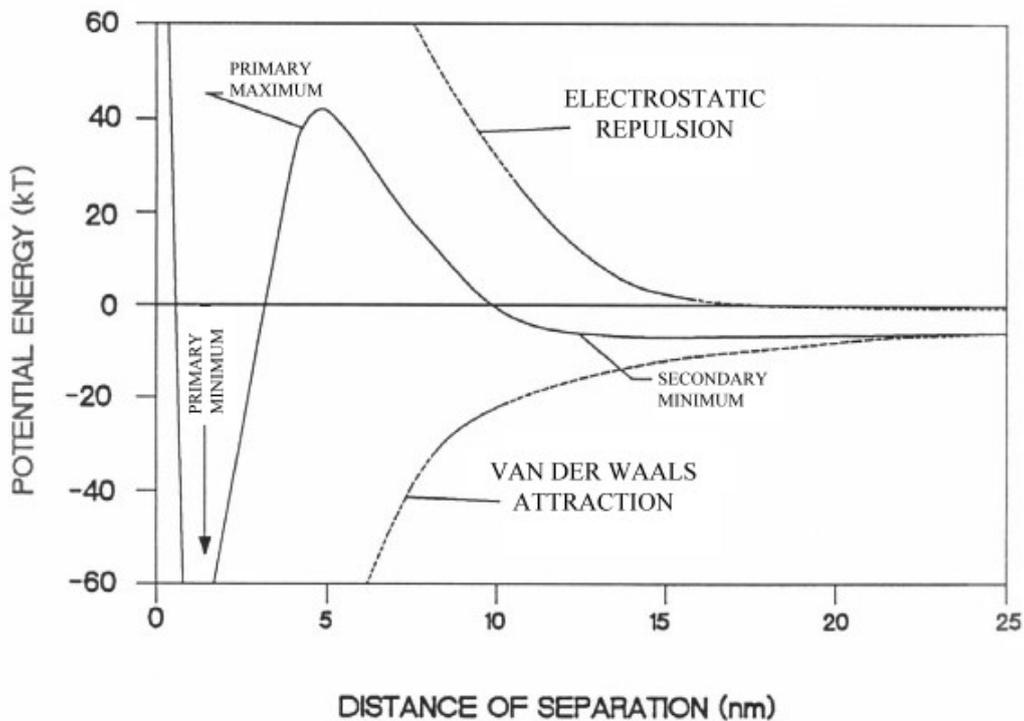


Figure 1. Potential energy versus distance for a pair of electrostatically stabilized particles. The separate contributions of van der Waals and electrostatic interactions are also shown.

[From: Narsimhan G. (1992). Emulsions, in *Physical Chemistry of Foods* (eds. H.G. Schwartzberg and R.W. Hartel). New York: Marcel Dekker].

3. Flocculation

In an unstable or a metastable system, van der Waals attraction leads to flocculation of a colliding pair of particles when the distance of separation between the two particles is small. Flocculation can be considered as a precursor to coalescence, and therefore it is important to know the rates of flocculation. The three mechanisms of particle loss are Brownian flocculation, gravitational flocculation, and creaming.

3.1. Brownian Flocculation

Brownian flocculation occurs because of a collision between the particles due to thermal motion. The effect of interparticle forces on Brownian flocculation is accounted for in the following expression for the rate of slow flocculation:

$$J_s = \frac{8\pi RDn_0}{W} \quad (12)$$

where D is the diffusion coefficient of the particle, n_0 is the particle number concentration, and the stability ratio W is given by:

$$W = 2R \int_{2R}^{\infty} \exp\left(\frac{U_{total}}{kT}\right) \frac{dr}{r^2} \quad (13)$$

In the above equation, U_{total} is the sum of different interaction potentials between the two colliding particles. In the absence of interparticle forces, the stability ratio is unity. Consequently, the numerator in Equation (12) gives the fast flocculation rate. Reerink and Overbeek (1954) have evaluated the following approximate expression for the stability ratio:

$$\log W = A - B \frac{R\gamma_0^2}{z^2} \log c \quad (14)$$

where A and B are constants, c is the electrolyte concentration, and z the valence number of the electrolyte. Equation (14) predicts that a log plot of W vs. c should yield a straight line in the slow flocculation regime. One can also identify a critical flocculation concentration of the electrolyte at which the stability ratio becomes unity. For electrolyte concentrations greater than the critical flocculation concentration, the stability ratio will still be unity because of the absence of an energy barrier. Consequently, a plot of $\log W$ vs. $\log c$ should consist of two straight lines intersecting at the critical flocculation concentration. The critical flocculation concentration has been shown to vary as the inverse sixth power (z^{-6}) of the valence number of the electrolyte for symmetrical electrolytes, such as NaCl, for example. In the case of non-symmetrical electrolytes, the valence number of the counterion has been found to influence the critical flocculation concentration.

-
-
-

TO ACCESS ALL THE 39 PAGES OF THIS CHAPTER,
Visit: <http://www.eolss.net/Eolss-sampleAllChapter.aspx>

Bibliography

Asakura S. and Oosawa F. (1954). On Interaction between Two Bodies Immersed in a Solution of Macromolecules. *J. Chem. Phys.* **22**, 1255. [Presents evaluation of interparticle forces due to depletion of free macromolecules as a result of osmotic pressure].

Bos M.A. and Nylander T. (1996). Interaction between β Lactoglobulin and Phospholipids at the Air/Water Interface. *Langmuir* **12**, 2791. [The surface pressure-area isotherms of mixed phospholipid- β Lactoglobulin monolayers indicated displacement of protein at sufficiently high phospholipid concentrations. The highest rate of adsorption of β Lactoglobulin was found to be on distearolphosphatidic acid monolayers. Both electrostatic and hydrophobic interactions have been shown to be important for the incorporation of β Lactoglobulin onto phospholipid monolayers].

Castle J., Dickinson E., Murray B.S., and Stainsby G. (1987). Mixed Protein Films Adsorbed at the Oil-Water Interface. *ACS Symp. Ser.* **343**, 118. [Experimental data on interfacial shear viscosity of mixed protein films at oil-water interface are given].

Chen J. and Dickinson E. (1995). Surface Shear Viscosity and Protein Surfactant Interactions in Mixed Protein Films Adsorbed at the Oil-Water Interface. *Food Hydrocolloids* **9**, 35. [The addition of Tween 20 to individual β Lactoglobulin or gelatin films brings the surface viscosity rapidly down to zero, but not so for the mixed β Lactoglobulin+gelatin film. The addition of sodium lauryl ether sulphate to the gelatin film produces a molecular rearrangement and a gradual decrease of the surface viscosity. A small amount of oil soluble surfactant diethylene glycol monododecyl ether induces a large increase in surface viscosity].

Chen J. and Dickinson E. (1995). Protein/Surfactant Interfacial Interactions. Competitive Adsorption of Protein plus Surfactant in Emulsions. *Colloids Surfaces A*: **101**, 77. [Results of competitive adsorption between proteins such as gelatin and β Lactoglobulin and nonionic (Tween 20), as well as anionic (Sodium lauryl ether sulphate) surfactants in various oil-water interfaces is presented].

Cho D., Narsimhan G., and Franses E.I. (1996). Adsorption Dynamics of Native and Alkylated Derivatives of Bovine Serum Albumin at Air-Water Interfaces. *J. Colloid Interface Sci.* **178**, 348. [The dynamics of adsorption of bovine serum albumin (BSA) and its derivatives at the air-water interface were investigated through the measurement of dynamic surface pressure. BSA was modified through reductive alkylation to yield methylated and pentylated BSA of higher surface hydrophobicities. The modified BSAs were found to occupy much larger areas per molecule than native BSA, possibly due to more unfolding].

Cho D., Narsimhan G., and Franses E.I. (1997). Adsorption Dynamics of Native and Pentylated Bovine Serum Albumin at Air-Water Interfaces: Surface Concentration/Surface Pressure Measurements. *J. Colloid Interface Sci.* **191**, 312. [¹⁴C radiolabelling was employed to measure the evolution of surface concentration during the dynamics of adsorption of native and pentylated BSA at air-water interface. The rate of adsorption as well as steady state value of surface concentration was smaller, whereas the rate of increase and steady state value of surface pressure were larger for p-BSA than native BSA. The area per adsorbed protein molecule during adsorption was smaller than that for a spread monolayer, indicating that the protein molecule partially unfolds during adsorption. Area per molecule for p-BSA was larger than that for BSA due to more unfolding of the p-BSA because of its lower conformational stability, as evidenced by the changes in CD spectra. The steady state surface pressure-surface concentration relationship at the air-water interface agreed well with spread monolayer isotherm, thus indicating that the adsorbed protein molecules completely unfolded after sufficiently long times. A previously developed mechanistic model for protein adsorption was modified by accounting for the adsorbed protein layer with different dielectric properties in the evaluation of electrostatic interactions].

Cho D., Narsimhan G., and Franses E.I. (1997). Interactions of Spread Lecithin Monolayers with Bovine Serum Albumin in Aqueous Solutions. *Langmuir* **13**, 4710. [The adsorption of BSA onto spread lecithin monolayers of 107 and 64 Å²/molecule was enhanced at short times, indicating synergism due to possible dissolution of BSA molecules into loosely or moderately packed lecithin monolayers. At long times,

monolayer composition was found, however, to be dominated by lecithin. BSA was expelled from the interface by close-packed lecithin monolayers, possibly because of surface pressure and steric exclusion effects].

Chen J. and Dickinson E. (1995). Protein/Surfactant Interfacial Interactions, 3. Competitive Adsorption of Protein plus Surfactant in Emulsions. *Colloids Surfaces A*: **101**, 77. [Results of competitive adsorption between proteins such as gelatin and β Lactoglobulin and nonionic (Tween 20) as well as anionic (Sodium lauryl ether sulphate) surfactants in various oil-water interfaces is presented].

Cornec M., Cho D., and Narsimhan G. (1999). Adsorption Dynamics of α Lactalbumin and β Lactoglobulin at Air-Water Interfaces. *J. Colloid Interface Sci.* **214**, 129. [Dynamics of adsorption of ^{14}C radiolabelled α Lactalbumin and β Lactoglobulin at the air-water interface was investigated through the measurement of surface pressure and surface concentration via whilhelmy plate and radiotracer technique. At low concentration, an apparent time lag was observed in the evolution of surface pressure for β lactoglobulin, but not for α lactalbumin, which was shown to be due to the non-linear nature of the surface pressure-surface concentration relationship for the former. The area per molecule of an adsorbed β lactoglobulin during the dynamics of adsorption was smaller than that for spread monolayer since it was not fully unfolded during adsorption. For α lactalbumin, however, no such differences were observed. A previously developed mechanistic model was improved to account for hydrophobic patches on the protein surface as well as adsorbed protein layer. The model predictions seem to indicate that α lactalbumin changes its orientation from side-on to end-on orientations at higher surface concentrations].

Cornec M., Mackie A.R., Wilde P.J., and Clark D.C. (1996). Competitive adsorption of β Lactoglobulin and β casein with Span 80 at the oil-water interface and the effects on emulsion behavior. *Colloids Surfaces A*: **114**, 237. [The interfacial tension, the emulsification capacity and interfacial concentration of β casein adsorbed layers was affected at lower Span 80 concentrations than β Lactoglobulin, which is believed to be due to *relative* surface activities of the proteins and the lower level of self-interaction between the β casein molecules. In contrast, the orthokinetic stability of β casein stabilized emulsions was greater than β Lactoglobulin stabilized emulsions in the presence of competing Span 80].

Cornec M., Wilde P.J., Gunning P.A., Mackie A.R., Husband F., Parker M.L., and Clark D.C. (1998). Emulsion Stability as Affected by Competitive adsorption Between and Oil-Soluble Emulsifier and Milk Proteins at the Interface. *J. Food Sci.* **63**, 39. [Depletion studies of emulsions stabilized by the mixture of proteins and emulsifier indicated that the emulsifier displaced β casein more readily than β lactoglobulin from the surface of the droplets, which correlated with emulsification and stability experiments].

Cornell D.G. and Patterson D.L. (1989). Interaction of Phospholipids in Monolayers with β Lactoglobulin Adsorbed from Solution. *J. Agric. Food Chem.* **37**, 1455. [The interaction of β lactoglobulin adsorbed from solution with phospholipids in monolayers has been studied via ultraviolet and circular dichroism spectroscopy of films transferred from the air/water interface of a Langmuir film balance].

Courthaudon J.L., Dickinson E., and Dalgleish D.G. (1991). Competitive Adsorption of β Casein and Nonionic Surfactants in Oil-in-Water Emulsions. *J. Colloid Interface Sci.* **145**, 390. [Competitive adsorption of β casein and three non-ionic surfactants has been investigated at hydrocarbon oil-in-water interface].

Courthaudon J.L., Dickinson E., Matsumara Y., and Clark D.C. (1991). Competitive adsorption of β lactoglobulin+Tween 20 at the Oil-Water Interface. *Colloids Surfaces* **56**, 293. [Up-to a surfactant to protein molar ratio of 1:1, addition of Tween 20 leads to a dramatic lowering of the surface viscosity of adsorbed β lactoglobulin at the n-tetradecane water interface without any significant change in the amount of protein adsorbed. At the surfactant to protein molar ratio of 8:1, there is complete displacement of protein from the emulsion drop surface].

Courthaudon J.L., Dickinson E., Matsumara Y., and Williams A. (1991). Influence of Emulsifier on the Competitive Adsorption of Whey Proteins in Emulsions. *Food Structure* **10**, 109. [The influence of Tween 20 on the composition of adsorbed protein films in hydrocarbon oil-in-water emulsion containing β lactoglobulin and α lactalbumin has been investigated at neutral pH].

Damodaran S. and Song K.B. (1988). Influence of Electrostatic Forces on the Adsorption of Succinylated β lactoglobulin at the Air-Water Interface. *Biochem. Biophys. Acta.* **954**, 253. [The rate of change of surface pressure as well as steady state surface pressure decreased progressively with increase in net charge (extent of succinylation) as a result of larger electrostatic energy barrier to adsorption. The results indicate that only a small fraction of charged groups are located at the interface].

Darling D.F. and Birkett R.J. (1986). Food Colloids in Practice. In *Food Emulsions and Foams* (ed. E. Dickinson), Special Publication No. 58. London: Royal Society of Chemistry, Burlington House. [A brief review of different factors involved in formation and stabilization of food colloids. The role of proteins and emulsifiers on emulsion stability is discussed. The effects of fat crystallization and particle adsorption at interfaces on emulsion stability are discussed. The effects of competitive adsorption and bridging flocculation on emulsion stability are also briefly pointed out].

Dickinson E. (1998). Proteins at Interfaces and in Emulsions: Stability, Rheology and Interactions. *J. Chem. Soc., Faraday Trans.* **94**, 1657. [This review discusses the relationship between the adsorbed structure and interfacial rheological properties of the adsorbed layer of protein at oil-water interfaces in emulsions].

Dickinson E., Owusu R.K., Tan S, and Williams A. (1993). Oil –Soluble Surfactants Have Little Effect on Competitive Adsorption of α Lactalbumin and β Lactoglobulin in Emulsions. *J. Food Sci.* **58**, 295. [Protein surface concentrations in emulsions stabilized by α lactalbumin, β lactoglobulin and their mixture were studied with various amounts of oil-soluble surfactant present during homogenization].

Dickinson E. and Iveson G. (1993). Adsorbed Films of β Lactoglobulin + Lecithin at the Hydrocarbon Water and Triglyceride Water Interfaces. *Food Hydrocolloids* **6**, 533. [The amount of protein adsorbed during formation of oil-in-water emulsions is substantially lower with soybean oil as the dispersed phase than with n-tetradecane. Limited protein displacement from the emulsion droplet surface occurs at high values of lecithin: protein molar ratios. The presence of lecithin at a molar ratio of 100 leads to a reduction in surface viscosity of the aged film by an order of magnitude].

Dickinson E. and Matsumura Y. (1991). Time Dependent Polymerization of β Lactoglobulin Through Disulphide Bonds at the Oil-Water Interface in Emulsions. *Int. J. Biol. Macromol.* **13**, 26. [Time dependent intermolecular sulphhydryl-disulphide interchange involving β lactoglobulin adsorbed at the oil-water interface in n-tetradecane in water emulsions has been investigated using gel electrophoresis].

Douillard R. and Lefebvre J. (1990). Adsorption of Proteins at the Gas-Liquid Interface: Models for Concentration and Pressure Isotherms. *J. Colloid Interface Sci.* **139**, 488. [A kinetic and statistical mechanical model of protein adsorption is proposed that assumes adsorption occurs in two layers. The surface pressure isotherm is obtained from the conformation of molecules in the first layer through the equation of state of an adsorbed polymer. Fitting of the model to experimental data suggests that casein can assume one conformation in the first layer, whereas globular proteins such as BSA and lysozyme assume two conformations].

Euston S.E., Singh H., Munro P.A., and Dalgleish D.G. (1995). Competitive Adsorption Between Sodium Caseinate and Oil-Soluble and Water-Soluble Surfactants in Oil-in-Water Emulsions. *J. Food Sci.* **60**, 1124. [Only partial displacement of protein was observed with Tween 60 of Span 60. The reduction in proteins surface concentration was greater in the presence of Tween 60. Interfacial protein composition was independent of surfactant type. In the absence of surfactant, preferential adsorption of β casein occurred in emulsions containing less than 1 wt percent proteins].

Frisch H.L. and Simha R. (1956). Monolayers of Linear Macromolecules. *J. Chem. Phys.* **24**, 652. [Extension of Singer's model for surface equation of state for macromolecules at interfaces. This model accounts for the formation of loops and trains at interfaces and considers only the entropic contribution of free energy].

Graham D.E. and Phillips M.C. (1979a). Proteins at Liquid Interfaces, I. Kinetics of Adsorption and Surface Denaturation. *J. Colloid Interface Sci.* **70**, 403. [The rates of change of surface pressure and

surface concentration of protein during the adsorption of β -casein, BSA and lysozyme at the air-water interface have been monitored by the Wilhelmy plate and surface radioactivity, respectively. The increases in surface pressure and surface concentration for the relatively flexible β -casein occur simultaneously with both attaining steady state around the same time. In contrast, for globular protein molecules, even though surface concentration increased steadily and reached a steady state faster, the surface pressure exhibited a t lag time and continued to increase slowly even after the surface concentration reached a steady state. The kinetics of adsorption was diffusion controlled at smaller times but at higher surface coverage an energy barrier to adsorption existed. Two timescales of adsorption were identified; the smaller timescale describes adsorption when both surface pressure and surface concentration were increasing, whereas the longer timescale refers to the change in the surface pressure after the surface concentration reached a steady state due to slow unfolding of globular protein].

Graham D.E. and Phillips M.C. (1979b). Proteins at Liquid Interfaces, II. Adsorption Isotherms. *J. Colloid Interface Sci.* **70**, 415. [Adsorption isotherms for the three proteins, β -casein, BSA and lysozyme, at the air-water and oil-water interfaces, have been determined using ellipsometry and surface radioactivity methods. Saturated monolayer coverage occurs via irreversible adsorption of 2 to 3 mg/m² of protein. Molecules adsorbed in the first layer dominate the surface pressure so that further adsorption onto multilayers does not affect the surface pressure. Experimental isotherm data and the Langmuir adsorption isotherm were in close agreement at low protein concentrations].

Guzman R.Z., Carbonell R.G., and Kilpatrick P.K. (1986). The Adsorption of Proteins to Gas-Liquid Interfaces. *J. Colloid Interface Sci.* **114**, 536. [A kinetic model for protein adsorption is proposed that accounts for a tightly adsorbed first layer and loose packing of a second layer. The model parameters were determined by fitting it to the adsorption isotherm data of Graham and Phillips to β -casein, BSA and lysozyme. The experimental adsorption rates were found to be faster than those predicted by diffusion alone. The rate of adsorption of protein was modeled in terms of a mass transfer coefficient].

Haynes C.A. and Norde W. (1994). Globular Proteins at Solid/Liquid Interfaces. *Colloids and Surfaces B: Biointerfaces* **2**, 517. [This review presents different factors that influence protein conformation in solution and protein adsorption at solid interfaces. Factors affecting protein folding and stability are discussed. Data on molecular dimensions and hydrophobicities of a variety of globular protein are given. The influences of protein hydrophobicity, charge, sorbent polarity, protein structural stability on adsorption isotherm of some globular proteins are presented. Hydrogen ion titration curves of some globular proteins are also presented. Thermodynamics of protein adsorption and principle forces involved in protein adsorption at solid interfaces are discussed].

Hiemenz P.C. and Rajagopalan R. (1997). *Principles of Colloid and Surface Chemistry*, Third Edition. New York: Marcel-Dekker. [This book on colloid and surface chemistry gives an excellent introduction to interparticle forces, electrical double layer, electrostatic and polymer-induced colloid stability, surface tension, Gibbs adsorption equation, osmotic and donnon equilibria, etc.].

Hunt J.A., Dickinson E., and Horne D.S. (1993). Competitive Displacement of Proteins in Oil-in-Water Emulsions Containing Calcium Ions. *Colloids Surf. A:* **71**, 197 [In the absence of calcium ions, addition of β casein to a phosvitin stabilized oil-in-water emulsion results in 70 percent of the originally adsorbed phosvitin becoming displaced within a few minutes, followed by the loss of a further 10 percent over a 48 hour period. In contrast, when calcium ions are present at a concentration sufficient to cause droplet aggregation in a mixed emulsion, no phosvitin is desorbed, despite substantial adsorption of the added protein. Displacement of phosvitin is facilitated when sufficient calcium ions are present to cause aggregation].

Hunter J.R., Kilpatrick P.K., and Carbonell R. (1990). Lysozyme Adsorption at the Air/Water Interface. *J. Colloid Interface Sci.* **137**, 462. [Adsorption isotherm measured using surface radioactivity indicated monolayer saturation at low concentrations. At intermediate concentrations, an abrupt increase in surface concentration with increasing bulk concentration indicates a change in orientation of the adsorbed protein molecule. Sequential adsorption experiments indicated that lysozyme adsorbed at concentrations below the multilayer region do not exchange significantly with the bulk].

Ibdah J.A. and Phillips M.C. (1988). Effects of Lipid-Composition and Packing on the Adsorption of Apolipoprotein-A-I to Lipid Monolayers. *Biochemistry* **27**, 7155. [The adsorption of human apolipoprotein A-I to phospholipid monolayers has been studied. The influence of lipid packing was investigated by spreading the monolayers at various initial surface pressures and by using various types of lipids].

Ivanov I.B. and Dimitrov D. (1974). Hydrodynamics of Thin Liquid Films: Effect of Surface Viscosity on Thinning and Rupture of Foam Films. *Colloid & Polymer Sci.* **252**, 982. [Discusses the analysis of liquid drainage due to capillary forces and disjoining pressure accounting for surface viscosity of foam films. The paper also discusses the growth of imposed thermal perturbations on a thinning film for the prediction of critical film thickness for rupture].

Kozorac Z., Dhathathregan A., and Möbius D. (1988). Adsorption of Cytochrome C to Phospholipid Monolayers Studied by Reflection Spectroscopy. *FEBS* **229**, 372.

MacRitchie F. (1986). Spread Monolayers of Proteins. *Adv. Colloid. Int. Sci.* **25**, 341. [A comprehensive review of different methods of spreading monolayers, different techniques for the measurement and interpretation of surface pressure-area isotherms, kinetic measurements of desorption of proteins using radiolabelling, reactions in monolayers and applications to biological membranes, foams and emulsions].

Murray B.S. and Dickinson E. (1996). Interfacial Rheology and the Dynamic Properties of Adsorbed Films of Food Proteins and Surfactants. *Food Sci. Technol. Int. (Jpn)* **2**, 131. [Factors affecting the interfacial rheology of films of food proteins and surfactants at air-water and oil-water interfaces are reviewed. A comparison is made between properties of adsorbed layers of small-molecule surfactants, proteins and mixed systems of protein+surfactant. The relationship between interfacial rheology and stability of emulsions is discussed].

Napper D.H. (1983). *Polymeric Stabilization of Colloidal Dispersions*. Academic Press, New York. [This book is a comprehensive treatment of different aspects of polymeric stabilization of colloidal dispersions. It introduces polymer solution thermodynamics and conformation of polymer molecules in solution and builds upon it to describe steric interaction between surfaces with attached polymers. It discusses theories of polymer stabilization and experimental data on colloid stability as well as distance dependence of steric interactions. Theoretical and experimental aspects of depletion flocculation and stabilization are also included].

Narsimhan G. and Uraizee F. (1992). Kinetics of Adsorption of Globular Protein at an Air-Water Interface. *Biotechnol. Prog.* **8**, 187. [Adsorption of globular proteins at an air-water interface from an infinite stagnant medium was modeled as one-dimensional diffusion in a potential field. The interaction potential experienced by an adsorbing protein molecule consisted of contributions made from electrostatic interactions, work done against the surface pressure to clear area at the interface in order to anchor the adsorbed segments, and the change in free energy due to exposure of penetrated surface hydrophobic functional groups to air. The energy barrier to adsorption, present at larger surface pressures, was found to be higher for smaller surface hydrophobicities, larger surface pressures, larger size molecules, and oblate orientation of an ellipsoidal molecule. More adsorption was shown to occur at larger surface hydrophobicities, smaller size molecules, and for prolate orientation of ellipsoidal molecules. The model predictions agreed with the experimental data for lysozyme].

Narsimhan G. (1992). Emulsions, in *Physical Chemistry of Foods* (eds. H.G. Schwartzberg, and R.W. Hartel). New York: Marcel Dekker. [This book chapter elucidates the role of proteins and emulsifiers in the stabilization of food emulsions. It describes the colloidal forces between particles. Flocculation of droplets due to creaming and Brownian motion are described. The role of thin film drainage and stability on drop coalescence is discussed. A brief discussion of drop break-up in a turbulent flow field during emulsion formation is also included].

Parker N.S. (1982). Properties and Functions of Stabilizing Agents in Food Emulsions. *CRC Crit. Reviews in Food Sci. & Nutrition* **25**, 285. [A comprehensive review describing the interparticle forces, flocculation, experimental methods of characterizing emulsion stability, factors that affect adsorption at

interfaces, competitive adsorption of mixed emulsifiers, and break-up of droplets during emulsion formation].

Reddy S.R. and Fogler H.S. (1981). Emulsion Stability: Delineation of Different Particle Loss Mechanisms. *J. Colloid Interface Sci.* **79**, 105. [Various particle loss mechanisms are delineated and the relative importance of Brownian flocculation and sedimentation flocculation compared. A general map presenting regimes of dominant particle loss mechanisms has been developed. The effects of particle size, surface potential, concentration, density difference, temperature and ionic strength on the relative importance of flocculation and creaming is analyzed].

Reerink H. and Overbeek J. Th. G. (1954). The Rate of Coagulation as a Measure of the Stability of Silver Iodide Sols. *Discuss. Faraday Soc.* **18**, 74-84. [Discusses the evaluation of stability ratio of colloids in terms of electrostatic energy barrier and the effect of ionic strength on colloid stability].

Schenkel J.H. and Kitchener J.A. (1960). A Test of the Derjaguin-Verwey-Overbeek Theory with a Colloidal Suspensions. *Trans. Faraday Soc.* **56**, 161. [Validation of DLVO theory of colloid stability from experimental data of evolution of particle number concentration from a coagulating colloidal suspension].

Singer S.J. (1948). Note on an Equation of State for Linear Macromolecules in Monolayers. *J. Chem. Phys.* **16**, 872. [An equation of state for monolayers of large threadlike molecules based on an idealized lattice model is proposed. This model accounts only for adsorption of macromolecules in the form of trains and considers only the entropic contribution in the evaluation of free energy of adsorbed macromolecule].

Uraizee F. and Narsimhan G. (1991). A Surface Equation of State for Globular Proteins at the Air-Water Interface. *J. Colloid Interface Sci.* **146**, 169. [A lattice model for surface equation of state of globular proteins at air-water interface was proposed accounting for the structure of the protein molecule, its degree of unfolding, segment-segment, segment-solvent, and electrostatic interactions. Fitting the equation to the experimental data indicates that the average degree of unfolding of BSA is greater than that for lysozyme. The number of adsorbed segments for BSA was found to vary linearly with the surface concentration, whereas the adsorbed segments for lysozyme were fairly independent of surface concentration. Due to exposure of unfolded hydrophobic functional groups, segment-solvent interactions were found to be unfavorable].

Wei A.P., Herron J.N., and Andrade J.D. (1986). The role of protein structure in surface tension kinetics. In *From Clone to Clinic* (eds. B.J.A. Crommelin and H. Schellekens), 305-313. Amsterdam: Kluwer Academic Publishers. [Investigates the surface tension kinetics and the lag period for adsorption of various proteins of different conformational stabilities at air/water interface].

Wüstneck R., Krägel J., Miller R., Fainerman V.B., Wilde P.J., Sarker D.K., and Clark D.C. (1996). Dynamic Surface Tension and Adsorption Properties of β casein and β Lactoglobulin. *Food Hydrocolloids* **10** (4), 395. [The adsorption kinetic behavior of β lactoglobulin and β casein solutions studied by dynamic surface tension measurements was interpreted by diffusion controlled models].

Biographical Sketch

Ganesan Narsimhan is a Professor of Agricultural & Biological Engineering at Purdue University. His monograph "*Surface Phenomena*" explains the role of proteins and emulsifiers in providing stability to colloidal dispersions, such as emulsions and foams, by modifying the interparticle colloidal forces as well as interfacial rheological properties. His research focuses on the role of proteins and other biomacromolecules on the structure, formation, stability and rheology of food emulsions, foams, and dispersions, with emphasis on (i) Adsorption of proteins at interfaces and (ii) Transport phenomena in dispersed systems. He teaches graduate level courses in *Colloidal Phenomena in Food & Biological Systems* and *Transport Phenomena in Food & Biological Engineering*, as well as undergraduate courses in *Food Process Engineering* and *Thermodynamics*.