

## VAGINAL SURGERY IN THE COW

**Laura Elvira-Partida,**

*TRIALVET S.L., C/ Encina, 22, 28721 Cabanillas de la Sierra, Madrid, Spain*

**Susana Astiz,**

*Department of Animal Reproduction, INIA, Avda Pta. de Hierro s/n, 28040 Madrid, Spain*

**Natividad Perez-Villalobos,**

*TRIALVET S.L., C/ Encina, 22, 28721 Cabanillas de la Sierra, Madrid, Spain*

**Juan Vicente Gonzalez-Martin**

*TRIALVET S.L., C/ Encina, 22, 28721 Cabanillas de la Sierra, Madrid, Spain  
and*

*Prof. Department of Animal Medicine and Surgery, Faculty of Veterinary Medicine, Universidad Complutense de Madrid (UCM), Avda Pta. de Hierro s/n, 28040 Madrid, Spain*

**Keywords:** vaginal surgery, anatomy of external cow genitalia, vaginal prolapse, pneumovagina, vaginal tears, rectovaginal fistula, urovagina, congenital abnormalities, vestibular gland cyst, Gartner duct cysts, vaginal tumors, perivaginal fat prolapse, presurgical considerations, surgical resolution

### Contents

1. Introduction
  2. Anatomy Fundamentals
  3. Genital Tract Exploration
  4. General Presurgical Considerations
  5. Vaginal Pathologies and Surgical Resolution
- Glossary  
Bibliography  
Biographical Sketch

### Summary

Farmers and veterinarians usually fail to appreciate the medical and economic benefits of vaginal surgical techniques. Some of these techniques are sufficient by themselves to solve fertility problems. They also solve problems such as urovagina, pneumovagina, vaginal tears and other minor problems of the reproductive tract. They offer many advantages: they can be scheduled and they are simple, inexpensive and fast. Moreover, these procedures often significantly improve the likelihood of pregnancy after artificial insemination. In this way, these techniques offer several economic benefits: they reduce the number of days from calving to conception (days open), the number of inseminations per pregnancy and the cost of maintaining herd size (replacement cost), while also enhancing genetic improvement. Moreover, these methods do not require any

pre- or postoperative antibiotic treatment, and no milk must be discharged. This chapter will describe the different vaginal pathologies and surgical techniques to resolve them.

## **1. Introduction**

At present bovine surgery is not a fashionable issue. Buiatric practice seems to focus on herd health, mastitis control, reproduction, indexes, computers, time, and money. Scientific articles and conferences deal mainly with herds rather than individuals. It is as if large farms were not formed by cows, but were a unit unto themselves--“no ants, only the mound.” The reality is quite different. Although very large cow farms do exist, the average farm is quite small with 30 cows per farm in the U.S. and even fewer in Spain. On all these farms, effective, fast and inexpensive surgical techniques are always very profitable for both the farmer and the veterinarian.

However, bovine surgery is not given much importance either in the world of animal surgery or in the curricula of veterinary schools. Most discussion is related with small animals' surgery, and large animal surgery is almost limited to equine procedures. This is not realistic, since bovine practitioners are routinely called upon to perform bovine surgery, and these cases cannot be submitted to specialized clinics as can be done with other species.

The most common surgeries in cattle are Cesarean section (C-section) in beef cows, and abomasum displacement in dairy cows. Other common surgeries are related to problems of the rumen, cecum, teats, feet, eyes or abdominal hernias. C-section is the most common bovine surgery for treating reproductive tract problems. Additionally, uterine and vaginal prolapse are urgent problems that require fast and effective surgical treatment, and thus such methods have also received attention.

Moreover, vaginal surgical techniques can be used to resolve fertility problems, but since these problems are not regarded as medical emergencies, farmers and veterinarians alike are usually unaware of their economic benefits. By resolving urovagina, pneumovagina, vaginal tears and other minor problems of the reproductive tract, these surgical procedures dramatically increase the probability of pregnancy after artificial insemination.

## **2. Anatomy Fundamentals**

The anatomy of the reproductive tract is one of the best-studied of all the organ systems in the cow. Indeed, the cow was the pioneer species in reproductive studies, and the first model for developing assisted-reproduction techniques such as artificial insemination and embryo transfer. Studies of the cow reproductive tract have been facilitated by the fact that it is easy to explore by direct methods (inspection and palpation), and the fact that large numbers of animals are available on farms and in slaughterhouses.

Nevertheless, certain anatomical characteristics, primarily pelvic conformation and body condition scores, have been described as predisposing factors of reproductive pathologies in the mare, and in a lesser extent in the cow.

This chapter will first provide an overview of the external reproductive anatomy of the cow, and then discuss the main anatomical features of reproductive pathologies.

## **2.1. Anatomy of Cow External Genitalia**

Knowledge of cow genital topography is necessary for some exploration methods, such as transrectal examination and ultrasonography, as well as for diagnosing pregnancy and identifying reproductive pathologies and their prognosis (Betteridge, 1970). The entire reproductive tract, except for the vulva, is located within the cow body, in the pelvic abdominal region. Its topography, which changes over the various stages of pregnancy, comprises the following structures.

### **2.1.1. Vulva**

The vulva is the external part of the reproductive tract and it is located ventrally to the anus. It is usually in a vertical position, with 20% located on the caudal edge of the pelvic floor and 80% below it (Grunert and Berchtold, 1988). It consists of two vulvar labia, the dorsal and ventral commissures, and the clitoris.

- The vulvar labia are covered externally by skin and internally by a stratified epithelium. They contain a circularly arranged muscle layer (vestibular constrictor muscle), which continues dorsally to the external sphincter of the anus. The main function of this muscle is to occlude the labia, thereby helping to isolate the cranial part of the genitals from the outside.
- The dorsal and ventral commissures are formed by the joining of the lips dorsally and ventrally, respectively.
- The clitoris is the homologue of the penis. It is 5-10 cm long and is located in a cranio-ventral position, usually covered by the lower third of the labia. Irrigation is supplied by urogenital and internal and external pudendal arteries. The innervation is the same as in the vagina. However, the vulva contains additional sensory fibers of pudendal and genital nerves (Sisson and Grossman, 1953).

### **2.1.2. Vagina**

The vagina is located in the pelvic cavity and it limits cranially to the cervix, caudally to the vulva, dorsally to the rectum and anus, and ventrally to the bladder, urethra and pelvic floor. It is the site of semen deposition during natural service. The vagina also serves as passage for the calf during calving. Another important function is to act as a line of defense, in which secretion of fluids by vaginal epithelium inhibits the growth of undesirable bacteria. Irrigation and innervation are provided by urogenital artery branches and the internal pudendal and pelvic autonomic nerve plexus. The vagina consists of the following three parts, in order from cranial to caudal: the vaginal fundus, the body of the vagina and the vaginal vestibule (Sloss and Dufty, 1980).

- The vaginal fundus is the most cranial part of the vagina, continuing with the cervix, which usually makes a short protrusion into the lumen at this portion. The vaginal fundus is broad and deep dorsally, but shallow and smooth ventrally.

- The body of the vagina is the longest portion of the vagina, extending for 25-30 cm. The line of demarcation between the body of the vagina and the vestibule is not obvious, except in the ventral part. Ventrally, a transverse fold, called the hymenal ring closure, covers the external opening of the urethra (Sloss and Dufty, 1980). This ring is formed by the medial part of the levator ani muscle, which runs along the pelvic diaphragm, and the bulbocavernosus muscle, also known as the vestibular constrictor muscle. The bulbocavernosus muscle originates on both sides of the ventral surface of the tendinous levator ani muscle, and it is reinforced by fibers of the levator ani muscle, giving rise to a thumb-sized muscle with spiral morphology. The union of these muscles constitutes the hymenal ring. These muscles run laterally, above the vestibular glands, and insert into the raphe on the ventral floor of the vagina at the suburethral diverticulum. In the young female, rudiments of the hymen usually remain as a ventral band. This structure is disrupted during natural service or artificial insemination, and less frequently during calving. However, the hymen can also remain throughout the life of the animal.

The vaginal vestibule is approximately 10-12 cm long, limiting caudally with the vulva.

This region contains the following structures:

- o Gartner ducts, which may end blind or open near the opening of the urethra;
- o Minor vestibular glands, located on the vestibular floor along its midline;
- o Greater vestibular glands or Bartholin's glands, which have a diameter of 1.5-3 cm and are located lateral to the glands above at the height of the urethra;
- o Suburethral diverticulum, which is 2.5-4 cm long and is located in a cranio-ventral position.

### **2.1.3. Cervix**

The cervix lies between the vagina and uterus. It is a powerful muscular tubular sphincter consisting of 3-5 musculo-fibrous bands with almost cartilaginous consistency, and it is designed to restrict access to the uterus. Although it is usually located in the pelvic cavity, it is frequently found on the cranial border of the pelvis or even in the abdominal cavity (Miller, 1989). The cervix wall is harder, thicker and stiffer than the uterine or vaginal wall, especially in multiparous animals, and although the entrance of the cervix does not project into the vagina, it may appear to do so in multiparous cows with prolapsed rings (Sloss and Dufty, 1980). The length of the cervix varies between 5 and 10 cm, and its diameter between 1.5 and 7 cm, and cervical size increases with age and parity. Within the cervix, three or four rings, called annular folds, can be distinguished. Although the cervix of the cow is difficult if not impossible to dilate manually, it expands naturally during estrus and delivery. Irrigation is provided by branches of the uterine artery and vaginal artery, while innervation comes from fibers of the pelvic autonomic nerve plexus (Arthur et al., 1991).

## **2.2. Closures or Seals of the Cow Genital Tract**

One of the main functions of the external genital tract is to keep the uterus isolated from the environment. This is accomplished through three well-defined seals that prevent the entry of environmental agents into the uterine lumen. These seals are the following, in order from caudal to cranial:

- o First seal: the vulva (vulvar constrictor muscle). This is the first external seal that isolates the vestibule from the environment.
- o Second seal: this is located between the vaginal vestibule and the vaginal body, and it comprises the vaginal levator muscle and the hymenal constriction. In addition to isolating the uterus from the outside, this structure prevents the reflux of urine into the cranial part of the vaginal body.
- o Third seal: the cervix. It separates the vagina and the uterus (Sloss and Dufty, 1980).

### **2.3. Relationships between External Genitalia and Other Anatomical Structures in the Cow**

Some anatomical characteristics related to vulvar conformation, pelvic bone conformation and body condition score (BCS) may predispose the animal to reproductive pathologies, and thus to a reduction in fertility. These relationships have been studied primarily in mares, where a proper vulvar seal is considered very important to prevent pneumovagina and achieve maximum reproductive efficiency. Pascoe (1979) developed a formula to predict a mare's predisposition to reproductive difficulties on the basis of conformation. He correlated vulvar length and angle of inclination of the vulva, deriving the "Caslick index," which is obtained by multiplying the angle of inclination of the vulva and the distance from the ischium to the dorsal commissure. This index can be used to classify mares into three types, allowing the prediction of reproductive problems. Mares with a Caslick index  $> 150$  were found to experience more numerous and severe reproductive problems, principally pneumovagina and urovagina. Pascoe (1979) concluded that mares with a Caslick index  $> 100$  should be treated surgically to avoid future complications.

This work in mares identified two key aspects of vulvar conformation: the angle between the vulva and the horizontal line and the length of the vulvar commissure. These anatomical relationships are poorly studied in the cow. Ayala et al. (2001) showed that, in the cow, the percentage of the vulvar commissure that is above the caudal edge of the pelvic floor is more important than its total length. A defective vulvar conformation favors the appearance of faults in the first and second vaginal seals, allowing the entrance of fecal material, air and other external agents. This increases the risk of pathologies such as urovagina, pneumovagina and metritis, causing reproductive problems and reducing fertility (St Jean et al., 1988; Gilbert et al., 1989; Celly, 1990).

In the cow, the physiological position of the vulva is vertical,  $90^\circ$  to the horizontal, with 80% of the vulvar commissure below the caudal edge of the pelvic floor (Grunert and Berchtold, 1988). A defective vulvar angle ( $>90^\circ$ ) has two main causes: an elevated percentage of the vulvar commissure located above the caudal edge of the pelvic floor, and poor body condition. Regarding the first cause, the vulvar angle depends directly on the percentage of the vulvar commissure located above the caudal edge of the pelvic floor. The higher the percentage of the vulvar commissure located above the caudal edge of the pelvic floor, the less likely it is to be dragged cranially. Ayala et al. (2001) observed a greater incidence of urovagina and pneumovagina in cows in which the vulvar commissure was located more dorsally, on the floor of the pelvis. Regarding the second cause, a lower BCS means fewer fatty deposits in the perivaginal area, which allows the upper third of the vulva to shift to a cranio-ventral position. The BCS is

particularly important in cows with a vulva positioned dorsally, since a loss of only 2 BCS points (on a scale of 1 to 5) is sufficient to increase the vulvar angle. This problem can also occur in animals that have the optimal vulvar conformation but that suffer from cachexia. Indeed, low BCS is considered a cause of urovagina and neumovagina (Celly et al., 1990).

Another structural aspect of the cow reproductive tract that can affect fertility is pelvic conformation. The most important issue is the hip angle, which is the angle formed between the ischia and coxal tuberosity. This relationship directly affects reproductive indices, facilitating or preventing proper drainage of the reproductive tract (Watson, 1984). The hip angle also significantly affects vulvar conformation. It has been shown that the caudal reproductive tract (vulva and vagina) occupies a more cranio-ventral position in animals with ischial tuberosities at the same level as, or above, the coxal tuberosity (Figure 1), which is known as splacnoptosis vulvar (Figure 2). In addition, animals that have a sloped hip angle tend to have a smaller percentage of vulvar commissure below the pelvis edge. Variation in the vulvar angle is less frequent in cows with a desirable hip angle; in these animals, the vulvar commissure is normally found in a physiological position (Ayala et al., 2001).



Figure 1. Sloped hip angle



Figure 2. Vulvar splacnoptosis

### **3. Genital Tract Exploration**

#### **3.1. External Exploration**

External examination can be performed by inspection and palpation, and no special containment methods are required. It can be performed during routine reproduction visits. First, the anatomical integrity of the entire area should be checked, paying attention to the presence of second- and third-degree perineal injuries. Second-degree lacerations affect the vulva and perineum; third-degree lacerations affect the vulva, perineum and rectum. If injuries are present, the stage of healing should be determined in order to identify the best treatment method and timing.

The position and anatomical relationships of the various organs, especially the vulva and the pelvis, as well as the BCS and the degree of depletion of the ischiorectal fat should also be evaluated.

Finally, the efficiency of the vulvar and hymenal constriction closure should be checked, which can be done by opening the labia with two fingers and observing whether air enters the vagina when the diaphragm creates negative pressure during expiration. It should be noted that mild vaginal closure failure is observed only during estrus due to the relaxing effect of estrogens on connective tissue.

##### **3.1.1. Rectal Palpation**

Rectal palpation is the standard examination method at reproduction visits. It normally involves locating the cervix, usually at the midline anterior border of the pelvic floor, followed by exploring the uterus and ovaries.

The vagina is usually unnoticed, except when pneumovagina is present. Rectal palpation can also allow the diagnosis of perivaginal masses as being, for example, fatty nodules or abscesses. A complete reproductive diagnosis is always necessary before vaginal surgery can be undertaken, since it is contraindicated when injuries incompatible with pregnancy are present, such as adhesions.

##### **3.1.2. Vaginal Examination**

This exam is not routinely performed. It requires more extensive animal containment measures than are normally necessary, as well as previous cleaning and disinfection. It is not essential for diagnosing the most important vaginal diseases, but it is necessary when exploring the vestibule and when identifying cases of urovagina involving small amounts of urine. In addition, inflammation of the vagina (vaginitis) or uterine wall (metritis) can be diagnosed only by ultrasonography or vaginal examination, either manually or with a speculum.

## 4. General Presurgical Considerations

### 4.1. Immobilization and Anesthesia

#### 4.1.1. Immobilization

Vaginal surgery is always performed with the animal standing up, except for cervix fixation to the prepubial tendon. Therefore, the animal should be immobilized through a halter that is tied as short as possible to a fixed location. The tail should be tied with a rope to one anterior extremity and then tightened to expose as large a field as possible (Figure 3).

If the animal's head is properly immobilized and the animal is adequately anesthetized, the surgery can usually be carried out with two assistants, one on each side of the animal.

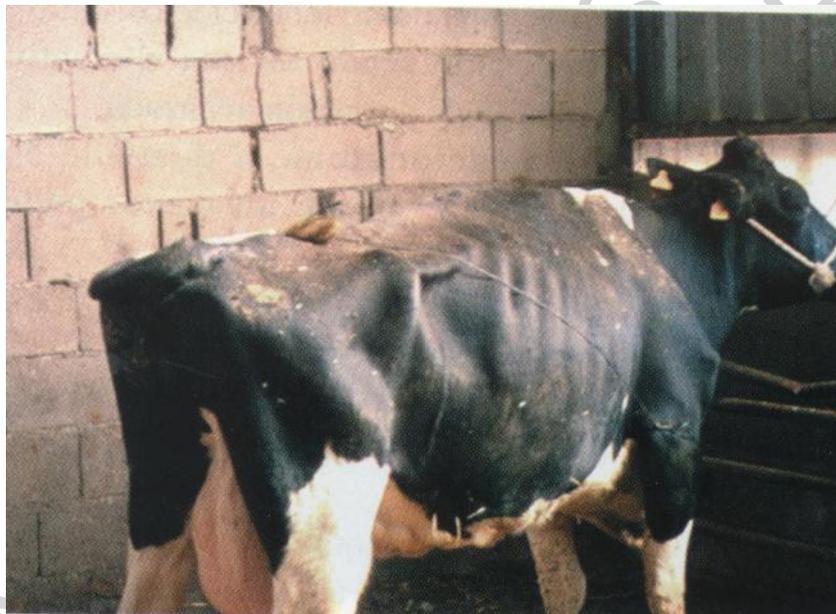


Figure 3. Proper immobilization of a Holstein cow

#### 4.1.2. Anesthesia

Most examinations and surgeries of the vaginal area can be performed only with restraint and regional anesthesia. However, sedation or tranquilization is indicated when adequate facilities are unavailable or if the animal is excessively nervous. In fact, tranquilization can be helpful even when surgeries are performed under optimal conditions, since it will leave us less anxious and better able to focus on the surgery. However, as surgeries are performed with the cow standing up, overdosing should be avoided in order to prevent the animal from lying down.

Most tranquilizers used in cattle are  $\alpha$ -2-agonists. The most common cow tranquilizer has traditionally been xylazine (Caron and Leblanc, 1989), and several drugs used in horses have recently been extended to cows, such as detomidine, medetomidine and

-  
-  
-

TO ACCESS ALL THE 41 PAGES OF THIS CHAPTER,  
Visit: <http://www.eolss.net/Eolss-sampleAllChapter.aspx>

### Bibliography

Ayala LE, Astiz S, González HV. (2001) Relaciones topográficas de la pelvis y el aparato reproductor externo de la vaca. *Producción Animal* Nov 2001; 69: 4-21. [Thesis studying the reproductive caudal tract topography and its relation with pathology].

Berchtold M and Rüsç P (1978): Die operative Behandlung der Uro-vagina beim Rind. *Berl Münch Tierärztl Wochenschr*; 91: 318-321. [Describes different surgical techniques to solve urovagina].

Betteridge KJ. (1970) The normal genital organs (In) Laing JA (eds): *Fertility and infertility in the domestic animals*. London. Bailliere, Tindall y Cassel, 26-53. [Anatomy of the caudal reproductive tract].

Caron, JP and LeBlanc PH. (1989) Caudal epidural analgesia in cattle using xylazine. *Can J Vet Res* 53: 486-489. [Review of caudal epidural analgesia with xylazine in cattle].

Celly CS, Dhoble RL, Singh GR. (1990) Modified Caslick's operation for bovine pneumovagina a case report. *Indian Vet J*; 67: 655-656. [Describes a Caslick modified technique to solve pneumovagina in the cow].

Dhillon KS, Singh BB, Kumar H, Bal MS, Singh J. (2006) Treatment of vaginal prolapse in cows and buffaloes. *Vet Res*. 2006 158(9): 312. [Resolution of vaginal prolapse].

Divers, T and Peek, S. (2008) *Rebhhun's diseases of dairy cattle*. 2nd edition. Philadelphia. W. B. Saunders Company [Reference book about dairy cattle internal medicine].

Dreyfuss DJ, Tulleners EP, Donawick WJ, Ducharme NG. (1990) Third-degree perineal lacerations and rectovestibular fistulae in cattle: 20 cases (1981-1988). *J Am Vet Med Assoc.*; 196(5):768-70. [Description of different clinical cases of 3rd degree vaginal lacerations and rectovestibular fistulae].

Erben J. (1975) Pneumo-vagina des Rindes. *Tierärztl Umsch*; 30: 286-289. [Treatment of pneumovagina in cattle].

Farhoodi M, Nowrouzian I, Hovareshti P, Bolourchi M, Nadalian MG. (2000) Factors associated with rectovaginal injuries in Holstein dairy cows in a herd in Tehran, Iran. *Prev Vet Med.*; 20; 46(2):143-148. [Retrospective study of 147 with rectovaginal injuries to determine risk factors].

Fathalla M, Abdou, MSS and Fahmi H. (1978) Bartholin Gland Cyst in the Cow. *Can. vet. J.*; 19: 340. [Case report about Bartholin Gland Cyst in the cow].

Fierheller, EE, Caulkett NA and Bailey JV. (2003) A romifidine and morphine combination for epidural analgesia of the flank in cattle. *Vet Clin North Am Food Anim Pract*; 19 (3): 679-693. [Describes epidural analgesia with romifidine and morphine combination in cattle].

Gautam G, Nakao T. (2009) Prevalence of urovagina and its effects on reproductive performance in Holstein cows. *Theriogenology*; 71(9):1451-61. [Study of urovagina prevalence, classification, predisposing factors and effect over reproductive pathologies and fertility].

Gilbert O, Wilson D, Levine S, Bosu W. (1989) Surgical management of urovagina and associated infertility in a cow. *JAVMA*; 194: 931-942. [Urethral extension surgery resulted in resolution of the urovagina and endometrial recovery].

Gilbert R and Fubini SL. (2005) Cirugía del sistema reproductor y vías urinarias en bovinos. En Fubini, S. L.; Ducharme, N.G. *Cirugía en animales de granja*. Ed. Intermédica S.A.I.C.I. Buenos Aires. pp. 367-396. [Reference book about farm animal surgery].

González JV (2000): Cerclaje de la constricción himeneal, nueva técnica quirúrgica para el tratamiento de la urovagina en la vaca. Congreso Mundial de Buiatría. Punta del Este. Uruguay. Proc, 2000. [Description of the hymenal cerclaje technique to solve urovagina].

González-Martín, JV, Astiz, S, Elvira, L, López-Gatius F. (2008) New surgical technique to correct urovagina improves the fertility of dairy cows. *Theriogenology*; 69: 360-365. [Description of the hymenal cerclaje technique to solve urovagina and the effects over fertility].

Grunert E y Berchtold M. (1988) *Infertilidad en la vaca*. 1ra ed. Hemisferio sur S.A. 267. [Book describing the infertility in the cow].

Hendrickson DA. (2007) *Techniques in large animal surgery*, Ed. Blackwell Publishing. U.S.A [Book about surgery in large animals].

Hofmeyr, CFB. (1987) *Ruminant urogenital surgery*. Iowa State University Press Ames, Iowa. [Book about urogenital surgery in cattle].

Hudson RS. (1980) Surgical procedures of the reproductive system of the cow. In: Morrow DA (eds): *Current therapy in theriogenology*. Philadelphia. W. B. Saunders Company, pp.: 257-271. [Chapter about surgeries of the reproductive tract in the cow].

Hudson, RS. (1986) Genital surgery of the cow. In: Morrow, DA. *Current therapy in theriogenology 2*. Philadelphia W. B. Saunders Company, pp.: 341-352. [Chapter about surgeries of the reproductive tract in the cow].

Hull BL. (1995) Female reproductive surgery. *Vet Clin North Am Food Anim Pract.*; 11(1):37-53. [Review of miscellaneous reproductive surgeries in the cow].

Miesner MD, Anderson DE. (2008) Management of uterine and vaginal prolapse in the bovine. *Vet Clin North Am Food Anim Pract*; 24(2): 409-419. [Review of vaginal and uterine prolapse resolution in the cow].

Miller HV. (1989) Endometritis of dairy cattle: Diagnosis, treatment and fertility. *Bov Pract*; 15:13-23. [Description of endometritis in the dairy cow].

Monin T. Vaginoplasty (1972): a surgical treatment for urine pooling in the mare. *Proc Am Assoc Equine Pract*; 18: 99-102. [Surgical treatment of urovagina in the mare through translocation of the urethral fold].

Noakes DE. (1996) Infectious forms of infertility in cattle. In: Arthur GH, Noakes DE, Pearson H, Parkinson TJ. *Veterinary reproduction and obstetrics*. 7th ed. London: W. B. Saunders Co., pp.: 389-422. [Chapter about infertility of infectious origin in cattle].

Pascoe RR. (1979) Observations on the length and angle of declination of the vulva and its relation to fertility in the mare. *J Reprod Fert Suppl*; 27: 299-305. [Anatomy of external genitalia in the mare and its relation with fertility].

Pouret, EJM. (1982) Surgical technique for the correction of pneumo and urovagina. *Equine Vet J*; 14: 249-250. [Description of the Pouret technique to solve pneumo- and urovagina].

Prado TM, Schumacher J, Hayden SS, Donnell RR, Rohrbach BW. (2007) Evaluation of a modified surgical technique to correct urine pooling in cows. *Theriogenology*; 67:1512–1517. [Evaluation of a modified urethral extension technique for urovagina to prevent the formation of a fistula].

Prado, ME, Streeter, RN, Mandsager, RE, Shawley, RV, Claypool, PL. (1999) Pharmacologic effects of epidural versus intramuscular administration of detomidine in cattle. *Am J Vet Res*; 60 (10): 1242-1247. [Detomidine use in cattle: intramuscular versus epidural analgesic and systemic effects].

Sisson S y Grossman JD. (1953) *The anatomy of the domestic animals*. 4th ed. Philadelphia. W. B. Saunders Company. [Reference book about anatomy in domestic animals].

Sloss V y Dufty JH. Gross anatomy (In) Sloss V y Dufty JH (eds) (1980): *Handbook of bovine obstetrics*. London. Baltimore. Williams y Wilkins, 17-27. [First book on veterinary obstetrics exclusively in the cow].

St Jean, GS, Hull, BL, Robertson, JT. (1988) Urethral extension for correction of urovagina in cattle: a review of 14 cases. *Vet. Surgery*; 17(5): 258-262. [Review of 14 cases of urovagina solved by urethral extension technique, its effect over fertility and the most frequent complications founded].

Trotter GW. (1992) Surgical diseases of the caudal reproductive tract (In) Auer JA (eds): *Equine surgery*. Philadelphia. W. B. Saunders Company, pp.: 730-750. [Surgeries of the caudal reproductive tract in the mare].

Watson P. (1984) La debilidad en el lomo causa problemas de fertilidad. *Frisona Española*; 21: 63-66. [Relationships between pelvic conformation and fertility].

Wolfe, DF and Baird, AN. (1993) Female urogenital surgery in cattle. *Vet Clin of North Am. Food Anim Prac* 9 (2): 369-388. [Review of urogenital surgery in the cow].

Youngquist, RS. (1997) Surgical correction of abnormalities of genital organs of cow. In: Youngquist, RS ed. *Current therapy in large animal. Theriogenology*. Philadelphia: W.B. Saunders Co. pp.: 429-440. [Chapter about urogenital surgery in the cow].

### **Biographical Sketches**

**Laura Elvira Partida** is practitioner veterinarian, specialized in mastitis, milk quality, and calf rearing. She has consultancy experience, being member of Spanish and European Mastitis Groups, and has published articles, chapters and books in this area. She collaborates in practical teaching for the Animal Medicine and Surgery Department of the Veterinary Faculty of Madrid, Spain (U.C.M.). She is also co-editor of the Spanish Association of Bovine Practitioners (A.N.E.M.B.E.) bulletin.

**Susana Astiz** is a recognized bovine veterinarian scientist specialized in bovine medicine and reproduction. She studied Veterinary Medicine in Madrid, became Master in Health Sciences in Barcelona, Spain and obtained the PhD degree from the Tierärztliche Hochschule in Hannover, Germany. She worked for several years as practitioner in Germany and Spain (with specialization in bovine medicine and reproduction), and as technical manager of ruminants for pharmaceutical companies (Schering Plough and Pfizer AH). She is Diplomat of the European Colleague of Herd Health Management (E.C.B.H.M.). In the present she is researcher at the Department of Animal Reproduction of the National Institute of Agricultural and Food Research (INIA) in Madrid, Spain.

**Natividad Pérez Villalobos** is veterinarian, specialized in bovine medicine and reproduction. She has consultancy experience, and is veterinary collaborator in practical teaching for the Animal Medicine and Surgery Department of the Veterinary Faculty of Madrid, Spain (U.C.M.). She has published articles, chapters and books related to bovine medicine. She also collaborates in the bulletin of the Spanish Association of Bovine Practitioners (A.N.E.M.B.E.).

**Juan Vicente González Martín**, a veterinary with a large experience in internal medicine and herd health in bovine. He has dilated clinical and consultancy experience, as well as in bovine research. He is professor of the Department for Animal Medicine and Surgery of the Veterinary Faculty of Madrid, Spain (U.C.M.) for more than 20 years. Additionally, he is member of the board of the Spanish Association of Bovine Practitioners (A.N.E.M.B.E.) and Diplomat of the European Colleague of Herd Health Management (E.C.B.H.M.). He has published numerous articles, chapters, and books related to bovine medicine and management, and is frequently invited as speaker to national and international buiatric events. He is also co-editor of the Spanish A.N.E.M.B.E. bulletin.

UNESCO-EOLSS  
SAMPLE CHAPTERS