

## GRAPEVINE BREEDING AND GENETICS

**P. Tantasawat, O. Poolsawat and W. Chaowiset**

*School of Crop Production Technology, Suranaree University of Technology, Nakhon Ratchasima 30000, Thailand*

**Keywords:** biotechnology, botany, conventional breeding, DNA fingerprinting, embryo rescue, genetic engineering, genetic map, genetic resource, genomic, in vitro selection, micropropagation, marker-assisted selection, molecular marker, mutation, polyploidy, quality, resistance, tissue culture, tolerance, yield.

### Contents

1. Introduction
  2. Botany and Taxonomy of Grapevines
  3. Grapevine Genetic Resources
  4. Inheritance and Genetic Analyses of Grapevines
  5. Grapevine Improvement
  6. Conclusions
- Acknowledgments  
Glossary  
Bibliography  
Biographical Sketches

### Summary

The grapevine is one of the most important economic fruit crops that are widely grown in almost all continents. The fruits produced worldwide are mainly processed into wine. In addition, significant portions have also been used for fresh consumption, dried as raisins or processed into juice. The objectives of grapevine breeding vary according to its usage and often are region-specific. However, most breeding programs aim at combining high yield and high fruit quality with improved resistance to multiple diseases and pests, and/or increased adaptation to adverse environments. These desirable traits can be exploited from the vast genetic resources of the *Vitis* genus. Different methods have been used to incorporate useful traits including conventional breeding, mutation and polyploid breeding, and biotechnological approaches. Integration of these tools will allow breeders to meet with the increasing demands for novel grapevine varieties with improved yield and quality in an era of limited resources, increasing health and ecological awareness, as well as increasing environmental pressures.

### 1. Introduction

The grapevine is an economically important woody perennial fruit crop, cultivated in most of the continents of the world. Approximately 71% of the world's harvest is processed into wine, 27% is consumed fresh and 2% is dried for raisins. In 2009, grapevine cultivation covered approximately 8 million hectares with ca. 67 million metric tons of fruit produced annually. Italy, China, United States, France and Spain

are the leading grapevine producers in the world. Other than these five countries, Turkey, Iran, Argentina, Chile, India, Australia and South Africa also produce grape in large quantities.

*V. vinifera* L. is the most widely cultivated *Vitis* species due to its adaptability to a wide range of temperate to subtropical or tropical conditions. It probably originated in the Mediterranean Basin and the Middle East, where it was domesticated 5,000 years ago. *V. vinifera* has given rise to over 14,000 cultivars grown in most of the cultivated areas worldwide. It is estimated that more than 90% of the world's grapevines are either *V. vinifera* or *V. vinifera* hybrids. *V. vinifera* cultivars are usually of outstanding quality but they are susceptible to a variety of biotic and abiotic stresses. Therefore, grapevine improvement is necessary to achieve specific goals and local adaptation. Numerous breeding programs have been conducted in different parts of the world using various breeding methods that will be mentioned in this chapter. The advantages, specific problems and challenges, as well as the achievements associated with each method will also be discussed. Continued advances in technology at the genomic level will increase the understanding of grapevine genetics and genes which underlie its important traits. Therefore, integration of these new and classical technologies into grapevine breeding programs will further ensure outstanding breeding successes in the future.

## 2. Botany and Taxonomy of Grapevines

The grape family, *Vitaceae* consists of 17 genera and more than 1,000 species. *Vitis* species are widely distributed in the tropics and subtropics, and have been extended into the temperate zones of the world. Variation in chromosome numbers is observed among different species/genus of *Vitaceae*. Some genus like *Ampelocissus*, *Parthenocissus* and *Ampelopsis* have a chromosome number of  $2n = 40$ . Species of the genus *Cissus* have a chromosome number of  $2n = 24$ . However, most genera of *Vitaceae* have a chromosome number of  $2n = 38$ . *Vitis* is the only food-producing genus in the family and is divided into 2 subgenera based on morphological criteria and chromosome number: *Muscadinia* Planch (muscadine grape),  $2n = 40$ , and *Euvitis* Planch (bunch grape),  $2n = 38$ . *Muscadinia* consists of three species including *V. munsoniana* and *V. rotundifolia* which are native to southeastern America and *V. popenoei*, native to Central America. The species of *Muscadinia* have also been considered within a separate genus, and their placement is still one of many ongoing debates. *Euvitis* species are interfertile and separated by geographic, phonologic and ecologic barriers, composing the American group (ca. 30 species), the Eurasia group (1 species) and the Asiatic group (more than 30 species). Most American species including *V. aestivalis*, *V. berlandieri*, *V. longii*, *V. cinerea*, *V. labrusca*, *V. monticola*, *V. riparia*, *V. rupestris*, *V. smalliana* and *V. shuttleworthii* have been used as parents in the breeding programs and/or rootstocks that exhibit resistance to several diseases and other pests, and tolerance to various soil and climatic conditions (Table 1). Native species in eastern Asia, China, Japan and south into Java constitute the Asiatic group. In China, 40 species, 1 subspecies and 13 varieties of Chinese wild grapes were found. Among these, *V. amurensis* is perhaps most commonly known. It has been frequently used to transfer cold hardiness and disease resistance (Table 1). In some areas of northeastern China and Japan, the edible fruits of this species are used as fresh fruit, juice, wine and jelly. Although, only one species (*V. vinifera*) was found in the Eurasia group, this species

and its hybrids constitute over 90 percent of cultivated grapes. *V. vinifera* consists of three subspecies, a cultivated subspecies, ssp. *sativa* D.C. (ssp. *vinifera*), and two wild subspecies, ssp. *sylvestris* Gmel. and ssp. *caucasica* Vav.

Species	Resistance to biotic stresses						Tolerance to abiotic stresses						Growth			Used for breeding			Other characteristics	Geographic location		
	Downy mildew	Powdery mildew	Black rot	Anthraxnose	Pierce's disease	Root-knot nematode	Phylloxera	Drought	Lime	Cold	Hot	Salinity	Iron	Grafting	Rooting	Vigor	New cultivars	Rootstocks			Interspecific hybrids	
<i>V. rotundifolia</i>	+	+	+	+	+	+	+				+						++	+	+			se USA
<i>V. rupestris</i>	+	+	+				+	+	-					+	+	+	++	++	+++			Originally c TX, AR, MS, TN, KY, WV nw MD, sw PA; now rare mostly s MO, n AR
<i>V. riparia</i>	+	+	+				+	-	+		+			+	+		++	++	+++	Flower & ripen early, high sugar & acid in fruit		All ne USA into Canada, south to n LA, VA
<i>V. monticola</i>							+	+												Poor growth & wood production		sc TX
<i>V. vulpina</i>	+		+		+		+		-	+						+						se USA
<i>V. aestivalis</i>	+	+			+		-			+					-				++	Desirable fruit characteristics		New England to GA, west to AR & Mississippi river
<i>V. lincedumii</i>	+			+			+	+			+											AR, LA, OK, TX
<i>V. bicolor</i>										+												ne North America from Canada to n GA
<i>V. candicans</i>	+	+	+		+	+	+	-		+	+			-	-		+					AL, AK, LA, OK, TX
<i>V. cinerea</i>	+	+	+				+		-					-	-		+	++				e & se USA
<i>V. berlandieri</i>	+	+					+	+	+		+	+		-	-		+	++				Texas, n Mexico
<i>V. labrusca</i>	+	+	+/-	+	-		+			+							++	++	+++	Large berries, strong distinctive flavor		East coast from ME to SC, west to OH, MI south to LA, AL
<i>V. acerifolia</i>	+	+					-	+		+		+	-		+		+	++		Mild flavor, seeds germinate at once		n NM & CO, KS, OK, n TX
<i>V. × champinii</i>					+	+	+	+			+	-	-				+	+		Natural hybrid ( <i>V.</i> <i>candicans</i> × <i>V.</i> <i>rupestris</i> )		sc TX
<i>V. amurensis</i>	+	+		+				-	+								+	++				China, Japan, Korea, Russia

AL, Alabama; AR, Arkansas; CO, Colorado; GA, Georgia; KS, Kansas; KY, Kentucky; LA, Louisiana; MO, Missouri; MS, Mississippi; OH, Ohio; OK, Oklahoma; MD, Maryland; ME, Maine; MI, Michigan; PA, Pennsylvania; SC, South Carolina; TN, Tennessee; TX, Texas; VA, Virginia; WV, West Virginia; c, central; e, east; n, north; s, south; w, west.

Table 1. Characteristics and usage of some *Vitis* species.

The unique characteristics of *Muscadinia* are non-shedding bark, prominent lenticels, short small clusters, thick-skin berries that detach one by one as they mature, and simple tendrils with no fork. In addition, they have a unique fruity aroma. Their seeds are oblong without beaks. It was found that a bunch of *V. munsoniana* and *V. rotundifolia* consists of 6-24 berries, which do not ripen at the same time. By contrast, *Euvitis* has shedding bark, forked tendrils, mostly elongated flower clusters and berries adhering to the stem at maturity. Their seeds are pyriform, with long and short beaks.

*Muscadinia* requires approximately 100 days on the vine for the fruits to mature and a long growing season. They have higher resistance levels to most of the diseases than *Euvitis*. In addition, they have been cultivated commercially since the middle of the 18<sup>th</sup> century. Most grape products like wine, juice and jam have been made from *V. rotundifolia* because of its unique fruity aroma. The interspecific crosses between *Euvitis* and *Muscadinia* are difficult to achieve due to chromosome number and genomic differences. Most of F<sub>1</sub> hybrids are completely or highly sterile, but some hybrid combinations can survive. However, the species of *Euvitis* are easily intercrossed and produce vigorous and fertile F<sub>1</sub> progeny.

Flowering of grape requires two consecutive growing seasons. The latent primary buds will be induced during the summer. However, initiation and development of the flower will not take place until the following spring. In addition, flowering in grapes is controlled by the gibberellin: cytokinin balance. Moreover, the external factors such as high light intensity, short term exposure to high temperature and optimum levels of soil moisture and macronutrients, are also factors which promote their flowering. There are three types of flower depending on the characteristics of the species and cultivars (Figure 1).

Perfect or hermaphroditic flowers have both functional pistils and stamens. The female flower has functional pistils. However, it has stamens that produce pollen, but is generally sterile. The male flower has an undeveloped pistil, but contains only a small rudimentary ovary which cannot be fertilized. These flower characteristics can be used to differentiate the wild and cultivated forms of grapevine. Male and female flowers are found on separate vines in the wild forms that are dioecious (most of them have male flowers), while the cultivated forms usually have hermaphroditic flowers.

In Chinese wild grapes, it was found that most types are male or female. Hermaphroditic flowers are found in only a few species of *V. quinquangularis*, *V. amurensis* and *V. davidii*. In perfect flowers, the pistil is normally surrounded by five stamens, although the number may vary from five to more on individual flowers of the same inflorescence. The other distinct characteristic between wild and cultivated forms is their seeds. Seeds obtained from wild grapes are round in shape with short beaks, while those from cultivated forms have long beaks.

In dioecious grapes, fertilization is most likely to occur by means of wind or insect pollination, whereas, in the hermaphroditic flowers, fertilization happens via self-pollination. The ovary and seeds of fertilized grapevine flowers develop into a fleshy fruit called a berry, typically known as a grape. Berries of *Euvitis* develop in clusters of up to 100 or more, but in *Muscadinia* small clusters of three to five berries are usually

formed. Each berry contains one to four seeds. Grape berries are non-climacteric fruits whose ripening is associated with an accumulation of soluble solids and berry pigmentation (anthocyanins). During ripening berry softness increases and hexoses accumulate while malic and tartaric acids decline.

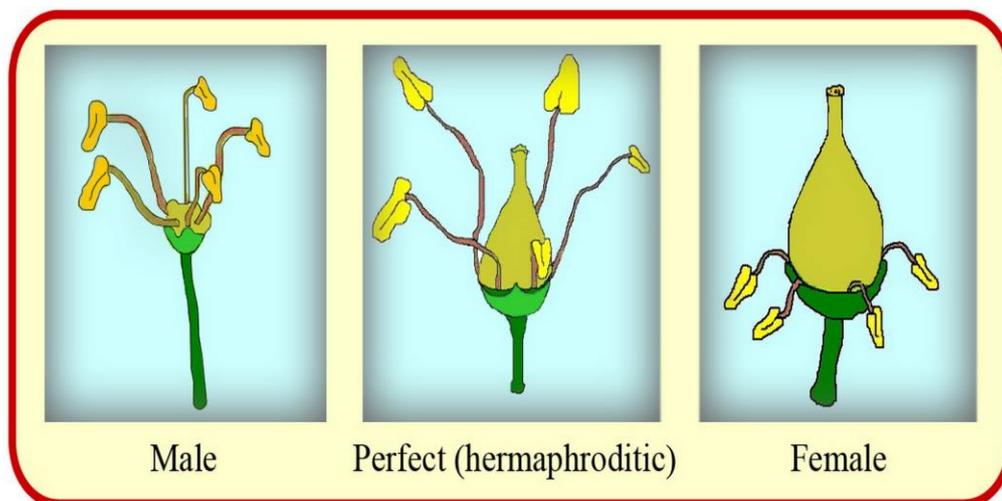


Figure 1. Three types of grapevine flowers.

### 3. Grapevine Genetic Resources

There are several species of wild and cultivated grapevines in the world, and each species consists of numerous varieties. In different countries more than 10,000 named varieties of grapevine were found. Moreover, the list of new varieties is continually being added to every year. Grapevine germplasm collections are crucial for maintaining the genetic resources of *Vitis*. Genetic resources can be divided into 4 types; wild species, old traditional cultivars, new cultivars and breeding lines. The wild species are potential sources for various traits (Table 2).

Both *Euvitis* and *Muscadinia* species are good sources of resistance. Several American species are resistant to phylloxera (*Dactylosphaera vitifoliae*; mainly *V. riparia*, *V. rupestris* and *V. berlandieri*) and are used extensively in breeding programs. Among these three species, only *V. berlandieri* is adapted to highly calcareous soils, and it is used to develop root stocks resistant to phylloxera and lime-induced chlorosis.

Wild Chinese species including *V. bryoniifolia*, *V. davidii* and *V. piasezkii*, and Asiatic species *V. amurensis* are good sources of powdery mildew (*Uncinula necator*), and/or downy mildew (*Plasmopara viticola*) resistance. *Muscadinia* is a useful source of resistance to phylloxera, nematodes, Pierce's disease (PD; *Xylella fastidiosa*) and fungal diseases including powdery mildew and downy mildew. Several sources of tolerance to abiotic stresses (cold, drought, salinity, lime etc.) are also observed in different *Vitis* spp. (Table 2), for example, cold tolerance is found in the northern species *V. riparia*, *V. labrusca* and *V. amurensis*.

While southern species, *V. lincecumii*, *V. bourquiniana* and *V. rotundifolia*, provide tolerance to hot conditions. However, these wild species have small berries with excessive seeds and strong pungent flavors. Therefore, extensive backcrosses are often needed to eliminate some unfavorable characteristics of the wild species. In addition to the wild *Vitis* spp., many traditional or new cultivars are also good sources of desirable traits (Table 2).

Desired properties	Disease/ nematode/ insect/ stress	<i>Vitis</i> species/ varieties
Disease resistance		
Fungal resistance	Powdery mildew ( <i>Uncinula necator</i> )	<i>V. aestivalis</i> , <i>V. amurensis</i> , <i>V. cinerea</i> , <i>V. berlandieri</i> , <i>V. labrusca</i> , <i>V. riparia</i> , <i>V. rotundifolia</i> , <i>V. rupestris</i> , <i>V. davidii</i> , <i>V. piasezkii</i> , <i>V. quinquangularis</i> , <i>V. romanetii</i> , ‘Nistru’
	Downy mildew ( <i>Plasmopara viticola</i> )	<i>V. aestivalis</i> , <i>V. amurensis</i> , <i>V. bryoniifolia</i> , <i>V. labrusca</i> , <i>V. lincecumii</i> , <i>V. flexuosa</i> , <i>V. piasezkii</i> , <i>V. pseudoreticulata</i> , <i>V. riparia</i> , <i>V. rupestris</i> , <i>V. rotundifolia</i> , <i>V. romanetii</i> , <i>V. yenshanensis</i>
	Black rot ( <i>Guignardia bidwellii</i> )	<i>V. candicans</i> , <i>V. cinerea</i> , <i>V. riparia</i> , <i>V. rupestris</i> , <i>V. rotundifolia</i> , ‘Chancellor’, ‘Léon Millot’, ‘Seyval’
	Anthraxnose ( <i>Elsinoe ampelina</i> )	<i>V. amurensis</i> , <i>V. munsoniana</i> , <i>V. labrusca</i> , <i>V. rotundifolia</i> , <i>V. smalliana</i> , <i>V. simpsoni</i> , <i>V. shuttleworthii</i> , <i>V. davidii</i> , <i>V. piasezkii</i> , <i>V. pseudoreticulata</i> , <i>V. quinquangularis</i> , <i>V. romanetii</i> , ‘Muscat Hamburg’, ‘Muscat Onitskanskkii’, ‘Suruchenskii’, ‘Bangalore Blue’, ‘Gros Colman’
	Botrytis bunch rot ( <i>Botrytis cinerea</i> )	<i>V. riparia</i> , <i>V. rupestris</i> , <i>V. vinifera</i>
	Rust ( <i>Physopella ampelopsidis</i> )	<i>V. caribaea</i> , <i>V. rotundifolia</i> , <i>V. simpsoni</i> , <i>V. shuttleworthii</i>
	Rotbrenner ( <i>Pseudopezicula tracheiphila</i> )	<i>V. cinerea</i> , <i>V. vinifera</i>
Bacterial resistance	Pierce’s disease ( <i>Xylella fastidiosa</i> )	<i>V. aestivalis</i> , <i>V. candicans</i> , <i>V. rotundifolia</i> , <i>V. smalliana</i> , <i>V. simpsoni</i> , <i>V. shuttleworthii</i> , <i>V. × champinii</i> , <i>V. vulpina</i> , ‘Norris’, ‘Lake Emerald’, ‘Blue Lake’
	Crown gall ( <i>Agrobacterium tumefaciens</i> )	<i>V. amurensis</i> , <i>V. labrusca</i> , ‘Chardonnay’, ‘Fateasca Regala’, ‘Muscat Ottonel’, ‘Rhine Riesling’, ‘Crimean Comichon’
Viral resistance	Grapevine fanleaf virus (GFLV)	<i>V. arizonica</i> , <i>V. candicans</i> , <i>V. riparia</i> , <i>V. rotundifolia</i> , <i>V. rufotomentosa</i> , <i>V. slavinii</i> , <i>V. vinifera</i>
Nematode resistance	Root knot nematode ( <i>Meloidogyne</i> spp.)	<i>V. candicans</i> , <i>V. × champinii</i> , <i>V. rotundifolia</i>
	Dagger nematode ( <i>Xiphinema index</i> )	<i>V. cinerea</i> , <i>V. rotundifolia</i> , <i>V. rufotomentosa</i>
Insect resistance	Phylloxera ( <i>Dactylosphaera vitifoliae</i> )	<i>V. berlandieri</i> , <i>V. cinerea</i> , <i>V. × champinii</i> , <i>V. rotundifolia</i> , <i>V. riparia</i> , <i>V. rupestris</i>
	Aphid ( <i>Aphis illinoisensis</i> )	<i>V. cinerea</i>
Abiotic stress tolerance	Cold	<i>V. adstricta</i> , <i>V. amurensis</i> , <i>V. acerifolia</i> , <i>V. labrusca</i> , <i>V. riparia</i> , <i>V. vinifera</i> ssp <i>sylvestris</i> var <i>tipica</i> , var <i>balcanica</i> , var <i>aberrans</i> , <i>V. vulpina</i> , <i>V. yenshanensis</i> , ‘Italian Riesling’
	Hot	<i>V. candicans</i> , <i>V. lincecumii</i> , <i>V. bourquiniana</i> , <i>V. rotundifolia</i>

Drought	<i>V. acerifolia</i> , <i>V. arizonica</i> , <i>V. × champinii</i> , <i>V. monticola</i> , <i>V. berlandieri</i> , <i>V. rupestris</i> , <i>V. vinifera</i>
Salinity	<i>V. acerifolia</i> , <i>V. berlandieri</i> , <i>V. riparia</i> , <i>V. candicans</i> , <i>V. × champinii</i>
Iron	<i>V. berlandieri</i> , <i>V. vinifera</i>
Lime	<i>V. monticola</i>

Table 2. Sources of resistance/tolerance and other desirable traits.

Germplasm collections have stored at least 10,000 grapevine varieties. However, because synonyms (many names for the same varieties) and homonyms (convergence of name for different varieties) occur in grapevine, a more accurate estimate of the number of varieties might be closer to 5,000. The true number of varieties and the relationships between them remain to be determined possibly by extensive DNA profiling of the grape varieties in different collections and the development of a common database. Germplasms are being maintained in the field as well as in tissue culture or cryopreservation (maintained at -196°C). It was found that selection of the superior accessions and evaluation of the germplasms are probably one of the first steps for grapevine breeding.

During the past ten years, there has been increasing interest in grape germplasm resources and genetic diversity analysis. It helps to protect some varieties of grapes, especially wild grapes like *V. vinifera* ssp. *sylvestris*, which is an ancestor of cultivated varieties, and facilitates their utilization in grapevine breeding programs. *Sylvestris* grapes can climb forest trees at about 20-30 m of height and produce small bunches of fruits. In addition, they resist/tolerate both biotic and abiotic stress factors including drought, lime, pests and diseases. Nowadays, information on grape database and germplasm collections can be found on the website (Table 3).

Databases	Source
Bulgarian <i>Vitis</i>	<a href="http://bulgenom.abi.bg/Genotyping%20and%20Mapping%20Crops%20Selected.htm/">http://bulgenom.abi.bg/Genotyping%20and%20Mapping%20Crops%20Selected.htm/</a>
European <i>Vitis</i>	<a href="http://www.eu-vitis.de/">http://www.eu-vitis.de/</a>
Greek <i>Vitis</i>	<a href="http://gvd.biology.uoc.gr/gvd/index.htm/">http://gvd.biology.uoc.gr/gvd/index.htm/</a>
Italian <i>Vitis</i>	<a href="http://www.vitisdb.it/">http://www.vitisdb.it/</a>
Swiss <i>Vitis</i>	<a href="http://www1.unine.ch/svmd/">http://www1.unine.ch/svmd/</a>
<i>Vitis</i> International Variety Catalogue	<a href="http://www.vivc.de/">http://www.vivc.de/</a>
Germplasm collections	Source
<a href="http://www.bdn.ch/culture/reben_startseite/">Conservation des ressources phyto-génétiques</a>	<a href="http://www.bdn.ch/culture/reben_startseite/">http://www.bdn.ch/culture/reben_startseite/</a>
<a href="http://www.deutsche-genbank-reben.jki.bund.de/">Deutsche Genbank Reben</a>	<a href="http://www.deutsche-genbank-reben.jki.bund.de/">http://www.deutsche-genbank-reben.jki.bund.de/</a>
INRA Daomaine de Vassal - Centre de ressources génétiques de la vigne	<a href="http://www1.montpellier.inra.fr/vassal/">http://www1.montpellier.inra.fr/vassal/</a>
The International Grape Genome Program (IGGP)	<a href="http://www.vitaceae.org/">http://www.vitaceae.org/</a>
The NCBI taxonomy for <i>V. vinifera</i>	<a href="http://www.ncbi.nlm.nih.gov/Taxonomy/Browser/wwwtax.cgi?mode=Info&amp;id29760&amp;lvl=3&amp;keep=1&amp;srchmode=1&amp;unlock">http://www.ncbi.nlm.nih.gov/Taxonomy/Browser/wwwtax.cgi?mode=Info&amp;id29760&amp;lvl=3&amp;keep=1&amp;srchmode=1&amp;unlock</a>
The Institute for Genomic Research (TIGR)	<a href="http://www.tigr.org/tigr-scripts/tgi/T_index.cgi?species=grape">http://www.tigr.org/tigr-scripts/tgi/T_index.cgi?species=grape</a>

The Genomic-Info Research Unit (URGI) of INRA	<a href="http://urgi.infobiogen.fr/GnpMap2/mapping/searchMap.do">http://urgi.infobiogen.fr/GnpMap2/mapping/searchMap.do</a>
The French National Resources Centre for Plant Genome (CNRGV)	<a href="http://cnrgv.toulouse.inra.fr/ENG/">http://cnrgv.toulouse.inra.fr/ENG/</a>
<a href="#">USDA, ARS Cold Hardy</a> Grape Collection	<a href="http://www.ars.usda.gov/Aboutus/docs.htm?docid=6245/">http://www.ars.usda.gov/Aboutus/docs.htm?docid=6245/</a>
<a href="#">USDA, ARS National</a> Germplasm Repository at Davis	<a href="http://www.ars.usda.gov/main/site_main.htm?modecode=53-06-20-00/">http://www.ars.usda.gov/main/site_main.htm?modecode=53-06-20-00/</a>

Table 3. Grape database and germplasm collections.

#### 4. Inheritance and Genetic Analyses of Grapevines

In grapevines, inheritance and genetic analyses have been carried out only for a limited number of traits because it has a long life cycle, a large number of chromosomes, partial sterility of ovules, and low seed germination. The following are examples of the traits that have been studied. Downy mildew resistance in *Vitis* is based on two genetic systems: (1) a single gene for the hypersensitive reaction of stomatal tissues, for which resistant species are homozygous dominant, and the susceptible *V. vinifera* is homozygous recessive, (2) polygenes for inhibition of the mycelium growth in plant tissues. Broad sense heritability for downy mildew resistance was estimated at 0.83 to 0.94, suggesting minimum environmental effects, while narrow sense heritability was 0.26 to 0.39. For powdery mildew resistance, a polygenic system has been suggested. The narrow sense heritability estimate for this trait ranged from 0.31-0.51, suggesting quite a high contribution of additive genetic variance. *Botrytis* resistance is associated with the synthesis of phytoalexin stilbene. This resistance had a narrow sense heritability and broad sense heritability of 0.23 to 0.26 and 0.82 to 0.92, respectively. It was found that resistance to anthracnose (*Elsinoe ampelina*) is controlled by two dominant genes for susceptibility ( $An_1$  and  $An_2$ ) and a single dominant resistance gene ( $An_3$ ).

The narrow sense heritability for anthracnose resistance was measured as 0.79, indicating that anthracnose resistance gene(s) were highly heritable. The resistance to black rot (*Guignardia bidwellii*) was reported to be either controlled by 2 dominant genes or quantitatively controlled. Previous research suggested that Pierce's disease resistance required 3 dominant genes,  $Pd_1$ ,  $Pd_2$  and  $Pd_3$ . Resistance to grapevine fanleaf virus can be obtained by using host plants resistant to the virus or plants resistant to the dagger nematode vector, *Xiphinema index*. The resistance to nematode was controlled by either a single dominant gene or by two genes, one dominant and one recessive, while the resistance to the virus was a recessive trait controlled by at least 2 genes. Resistance to phylloxera, an insect pest, is controlled by multiple genes. Those from *V. rotundifolia* appear to be dominant. *V. × champinii* and *V. candicans* confer resistance to the root knot nematode (*Meloidogyne* spp.), for which the resistance is mediated by a dominant gene. The heritability of resistance to this pest was estimated at 0.39, suggesting that additive effects were important.

Two types of seedlessness are found in grapevine: (1) stenospermocarpic, the seeds abort while still small and soft, (2) parthenocarpic, the seeds do not develop at all. The former is the one utilized in table grape improvement. Seedlessness is related to three independent recessive genes regulated by a dominant inhibitor locus, *SdI* (Seed Development Inhibitor). Two pairs of genes affect fruit color with epistatic action: *B*, a

dominant gene for black fruit, and *R*, a dominant gene for red fruit. The white-fruited grapes are recessive for both genes. Similarly, the composition of fruit anthocyanins is controlled by 2 genes: *G* for diglucosides or *g* for monoglucosides, and *O* for triphenols or *o* for diphenols. For fruit aroma, muscat flavor is controlled by 5 dominant complementary genes, methyl anthranilate is controlled by 3 dominant complementary genes, while volatile ester levels are determined by 2 genes. Several quality traits have generally high heritabilities: cluster compactness (0.55), berry weight (0.49), skin texture (0.75) and pulp texture (1.0).

-  
-  
-

TO ACCESS ALL THE 50 PAGES OF THIS CHAPTER,  
Visit: <http://www.eolss.net/Eolss-sampleAllChapter.aspx>

## Bibliography

- Allen, A. (2004) Vineyard and vintage view. *Mid-America Viticulture and Enology Center* 19(2), 1-17. [This article outlines grape maturity and sampling for growers.]
- Anonymous. (2003) Field trial of GM grapevines - evaluation of berry color, sugar composition, flower and fruit development and pollen flow study. *CSIRO Plant Industry, South Australia*, pp. 1-97. [This article evaluates the expression of genes related to fruit quality in transgenic grapevines.]
- Anonymous. (2011) Grape. *In Wikipedia, the free encyclopedia, developed under the auspices of the Wikimedia Foundation, Inc.*, [<http://en.wikipedia.org>]
- Atanassova, R., M. Leterrier, C. Gaillard, A. Agasse, E. Sagot, P. Coutos-Thévenot, and S. Delrot. (2003) Sugar-regulated expression of a putative hexose transport gene in grape. *Plant Physiology* 131(1), 326-334. [This article shows how to regulate expression of a putative hexose transport gene in grape.]
- Avramov, M.L., D.M. Milutinović, V.B. Sivcev, N.N. Gašić, M.D. Žunić, T.D. Nolić, and S.D. Vujović. (1996) Heredity of some berry characteristics in F<sub>1</sub> cross-breeding of grapevine variety Zupa Teinturier × Belgrade Seedless. *Journal of Wine Research* 7(3), 197-200. [This article describes grapevine breeding for seedlessness.]
- Babu, R., S.K. Nair, B.M. Prasanna, and H.S. Gupta. (2004) Integrating marker-assisted selection in crop breeding – prospects and challenges. *Current Science* 87(5), 607-619. [This review covers applications of marker-assisted selection in crop improvement.]
- Barritt, B.H., and J. Einset. (1969) The inheritance of three major fruit colors in grapes. *Journal of the American Society for Horticultural Science* 94, 87-89. [This article describes the inheritance of genes related to three major fruit colors in grapes.]
- Bellin, D., E. Peressotti, D. Merdinoglu, S. Wiedemann-Merdinoglu, A-F. Adam-Blondon, G. Cipriani, M. Morgante, R. Testolin, and G. Di Gasparo. (2009) Resistance to *Plasmopara viticola* in grapevine “Bianca” is controlled by a major dominant gene causing localized necrosis at the infection site. *Theoretical and Applied Genetics* 120(1), 163-176. [This article explains mapping of qualitative traits and QTL analysis.]
- Bogs, J., F.W. Jaffé, A.M. Takos, A.R. Walkers, and S.P. Robinson. (2007) The grapevine transcription factor VvMYBPA1 regulates proanthocyanidin synthesis during fruit development. *Plant Physiology* 143, 1347-1361. [This article is one of the few dealing with regulated proanthocyanidin synthesis during fruit development.]

- Boso, S., M.C. Martínez, S. Unger, and H.-H. Kassemeyer. (2006) Evaluation of foliar resistance to downy mildew in different cv. Albariño clones. *Vitis* 45(1), 23-27. [This article describes evaluation of downy mildew resistance in grapevine leaves.]
- Bouquet, A., and Y. Danglot. (1996) Inheritance of seedlessness in grapevine (*Vitis vinifera* L.). *Vitis* 35(1), 35-42. [This article explains the inheritance of seedlessness in grapevine.]
- Brînduse, E., M. Ionescu, and M. Tomescu. (2005) *Vinifera* genotype breeding for resistance to downy mildew by inter-specific hybridization using irradiated pollen. *Romanian Agricultural Research*, 33-40. [This article deals with conventional grapevine breeding to achieve downy mildew resistance.]
- Brown, D.C.W., and T.A. Thorpe. (1995) Crop improvement through tissue culture. *World Journal of Microbiology and Biotechnology* 11, 409-415. [A comprehensive review of several methods of plant tissue culture used for plant breeding.]
- Brown, M.V., J.N. Moore, P. Fenn, and R.W. McNew. (1999) Evaluation of grape germplasm for downy mildew resistance. *Fruit Varieties Journal* 53, 22-29. [This article identifies potential source of downy mildew resistance in grape germplasm.]
- Brumin, M., S. Stukalov, S. Haviv, M. Muruganatham, Y. Moskovitz, O. Batuman, A. Fenigstein, and M. Mawassi. (2009) Post-transcriptional gene silencing and virus resistance in *Nicotiana benthamiana* expressing a *Grapevine Virus A* minireplicon. *Transgenic Research* 18, 331-345. [This article shows an attempt to develop GVA resistant transgenic tobacco.]
- Burger, P., A. Bouquet, and M.J. Striem. (2009) Grape Breeding. *In* *Breeding Plantation Tree Crops: Tropical Species*. (eds. S.M. Jain and P.M. Priyadarshan) New York: Springer Science+Business Media. [This chapter reviews several aspects of grapevine breeding.]
- Carmona, M.J., P. Cubas, M. Calonje, and J.M. Martínez-Zapater. (2007) Flowering transition in grapevine (*Vitis vinifera* L.). *Canadian Journal of Botany* 85, 701-711. [This article explains the flowering process in the grapevine.]
- Chandra, R., M. Kamle, A. Bajpai, M. Muthukumar, and S. Kalim. (2010) *In vitro* selection: A candidate approach for disease resistance breeding in fruit crops. *Asian Journal of Plant Sciences* 9(8), 437-446. [This article provides information on the utilization of in vitro selection approach to increase disease resistance in fruit crops.]
- Chen, I., and S.R. Manchester. (2007) Seed morphology of modern and fossil *Ampelocissus* (*Vitaceae*) and implications for phylogeography. *American Journal of Botany* 94(9), 1534-1553. [This paper describes the taxonomy of grapevines.]
- Chervin, C., A. Tira-Umphon, P. Chatelet, A. Jauneau, P.K. Boss, and C. Tesniere. (2009) Ethylene and other stimuli affect expression of the UDP glucose: flavonoid 3-*o*-glucosyltransferase in a non-climacteric fruit. *Vitis* 48(1), 11-16. [This is an article dealing with the ethylene and expression of *ugt* in a non-climacteric fruit.]
- Clarke, E.E., Z. Ren, and J. Lu. (2003) Evaluation of grape germplasm for resistance to Pierce's disease and glassy-winged sharpshooter. *Proceedings of the Florida State Horticultural Society* 116, 32-35. [This article identifies accessions with Pierce's disease and glassy-winged sharpshooter resistance in grape germplasm.]
- Colova-Tsolova, V., A. Perl, S. Krastanova, S. Samuelian, and A. Atanassov. (2009) Progress in genetic engineering of grapevine for disease and stress tolerance. *In* *Grapevine Molecular Physiology & Biotechnology*, 2<sup>nd</sup> edn. (ed. K.A. Roubelakis-Angelakis). New York: Springer Science+ Business Media. [This chapter describes recent development in genetic engineering of the grapevine.]
- Common Wealth Scientific and Industrial Research Organisation. (2003) Field trial of GM grapevines – evaluation of berry colour, sugar composition, flower and fruit development and pollen flow study. Risk assessment and risk management plan. Developed under the auspices of the the Office of the Gene Technology Regulator, Australia, [<http://www.oagr.gov.au>] [This manuscript describes several transgenic grapevines that are on field trial in Australia.]
- Conner, P.J. (2009) A century of Muscadine grapes (*Vitis rotundifolia* Michx.) breeding at the university of Georgia. *Acta Horticulturae* 827, 481-484. [This article summarizes the progress on breeding of Muscadine grapes.]

- Costantini, L., F.M. Moreira, E. Zyprian, J.M. Martínez-Zapater, and M.S. Grando. (2009) Molecular maps, QTL mapping & association mapping in grapevine. *In Grapevine Molecular Physiology & Biotechnology*, 2<sup>nd</sup> edn. (ed. K.A. Roubelakis-Angelakis). New York: Springer Science+ Business Media. [This chapter describes recent applications of molecular markers and mapping in grapevine breeding.]
- Covert, C. (2008) Cold climate grape varieties from eastern U.S. breeding programs. *FPS Grape Program Newsletter*, pp. 10-12. [This article describes grape breeding programs for cold tolerance.]
- Cristinzio, G., C. Iannini, G. Scaglione, and M. Boselli. (2001) Effect of rootstocks on *Botrytis cinerea* susceptibility of *Vitis vinifera* cv. Falanghina. *Advances in Horticultural Science* 14, 83-86. [This article is about the effect of rootstocks on *Botrytis cinerea*.]
- Cutanda-Perez, M.-C., A. Ageorges, C. Gomez, S. Vialet, N. Terrier, C. Romieu, and L. Torregrosa. (2009) Ectopic expression of *VlmybA1* in grapevine activates a narrow set of genes involved in anthocyanin synthesis and transport. *Plant Molecular Biology* 69, 633-648. [This article shows regulation of anthocyanin biosynthetic genes by a transcriptional factor.]
- Davies, C., T. Wolf, and S.P. Robinson. (1999) Three putative sucrose transporters are differentially expressed in grapevine tissues. *Plant Science* 147, 93-100. [This is one of the articles on altered fruit sucrose content in grapevine tissues.]
- Di Gaspero, G., and F. Cattonaro. (2010) Application of genomics to grapevine improvement. *Australian Journal of Grape and Wine Research* 16, 122-130. [This article provides information on application of recent genomic technology to grapevine breeding.]
- Du, Y.P., H. Zhai, Q.H. Sun, and Z.S. Wang. (2009) Susceptibility of Chinese grapes to grape phylloxera. *Vitis* 48(1), 57-58. [This article is about phylloxera susceptibility in various Chinese grapes.]
- Fan, C., N. Pu, X. Wang, Y. Wang, L. Fang, W. Xu, and J. Zhang. (2008) *Agrobacterium*-mediated genetic transformation of grapevine (*Vitis vinifera* L.) with a novel stilbene synthase gene from Chinese wild *Vitis pseudoreticulata*. *Plant Cell, Tissue and Organ Culture* 92, 197-206. [This manuscript describes transformation of the grapevine to produce stilbene using *Agrobacterium*.]
- Fennell, J.L. (1948) Inheritance studies with the tropical grape. *Journal of Heredity* 39, 54-64. [This manuscript is about inheritance analysis of some agronomical traits in tropical grapes.]
- Gambino, G., I. Gribaudo, S. Leopold, A. Scharl, and M. Laimer. (2005) Molecular characterization of grapevine plants transformed with GFLV resistance genes: I. *Plant Cell Report* 24, 655-662. [Transgenic grapevine plants with GFLV resistance genes are described in this manuscript.]
- Gray, D.J., S. Jayasankar, and Z. Li. (2005) *Vitis* spp. Grape. *In Biotechnology of Fruit and Nut Crops*. (ed. R.E. Litz) New York: CABI publishing. [This chapter reviews several approaches of grapevine breeding.]
- Grzegorzczak, W., and M.A. Walker. (1998) Evaluating resistance to grape phylloxera in *Vitis* species with an *in vitro* dual culture assay. *American Journal of Enology and Viticulture* 49(1), 17-22. [This article describes in vitro evaluation of grape phylloxera resistance in grapes.]
- Gölles, R., R. Moser, H. Pühringer, H. Katinger, M.L. da Câmara Machado, A. Minafra, V. Savino, P. Saldarelli, G.P. Martelli, and A. da Câmara Machado. (2000) Transgenic grapevines expressing coat protein gene sequences of grapevine fanleaf virus, arabis mosaic virus, grapevine virus A and grapevine virus B. *Acta Horticulturae* 528, 305-311. [This article introduces transgenic grapevine expressing coat protein gene sequences of several plant viruses.]
- Hajdu, E. (2007) Breeding of table grape varieties in Hungary and beyond our national borders. *Hungarian Agricultural Research*, 4-9. [This manuscript covers table grape improvement using several approaches in Hungary.]
- Hemstad, P.R., and J.J. Luby. (1997) Utilization of *Vitis riparia* for the development of new wine cultivars with resistance to disease and extreme cold. *Annual Report of the Minnesota Grape Growers Association*, 7-10. [This manuscript shows the development of new wine cultivars to extreme cold using *V. riparia*.]

Hoffmann, S., P. Cindric, and P. Kozmajr. (2007) Breeding resistance cultivars to downy and powdery mildew. *In* 30<sup>th</sup> World Congress of Vine and Wine. 10-16 June, 2007. Kongresszusa, Budapest. [This article outlines improvement of grapevine for increased resistance to downy and powdery mildew.]

Huang, Y., D.F. Karnosky, and C.G. Tauer. (1993) Applications of biotechnology and molecular genetics to tree improvement. *Journal of Arboriculture* 19(2), 84-98. [This article reviews the application of tissue culture, genetic engineering and molecular markers in tree improvement.]

Hvarleva, Tz., A. Bakalova, K. Rusanov, G. Diakova, I. Ilieva, A. Atanassov, and I. Atanassov. (2009) Toward marker assisted selection for fungal disease resistance in grapevine. *Biotechnology & Biotechnology Equipment* 23, 1431-1435. [This review covers detailed application of marker assisted selection for fungal resistance in grapevine.]

International Service for the Acquisition of Agri-biotech Applications. (2011) Pocket K No. 15: ‘Omics’ sciences: Genomics, proteomics, and metabolomics. *In* Global Knowledge Center on Crop Biotechnology, developed under the auspices of the International Service for the Acquisition of Agri-biotech Applications (ISAAA), Metro Manila, Philippines, [<http://www.isaaa.org>]

Jackson, D. (1999) Grapes. (1999) *In* Temperate and Subtropical Fruit Production 2<sup>nd</sup> edn. (eds. D.I. Jackson and N.E. Looney) New York: CABI publishing. [This chapter describes genetic resources and breeding of grapevines.]

Jindal, P.C., and B. Shankar. (2002) Screening of grape germplasm against anthracnose (*Sphaceloma ampelinum* de Bary.). *Indian Journal of Agricultural Research* 36(2), 145-148. [The work in this manuscript discusses the screening of grape germplasm for accessions resistant to anthracnose.]

Korkutal, I. (2005) Embryo abortion in some new seedless table grape (*Vitis vinifera* L.) varieties. *International Journal of Botany* 1(1), 1-4. [This article describes the development and abortion of embryo in some new seedless table grapes.]

Krastanova, S., K.S. Ling, H.Y. Zhu, B. Xue, T.J. Burr, and D. Gonsalves. (2000) Development of transgenic grapevine rootstocks with genes from grapevine fanleaf virus and grapevine leafroll associated closteroviruses 2 and 3. *Acta Horticulturae* 528, 367-372. [This article shows development of transgenic grapevine rootstocks with genes from GFLV, GLRaV-2 and GLRaV-3.]

Laimer, M., D. Mendonca, F. Maghuly, G. Marzban, S. Leopold, M. Khan, I. Balla, and H. Katinger. (2005) Biotechnology of temperate fruit trees and grapevines. *Acta Biochimica Polonica* 52(3), 673-678. [This review describes application of molecular markers in grapevine and other fruit trees.]

Lestari, E.G. (2006) In vitro selection and somaclonal variation for biotic and abiotic stress tolerance. *Biodiversitas* 7(3), 297-301. [This review summarizes the basis and application of in vitro selection for biotic and abiotic stress tolerance.]

Li, D., Y. Wan, Y. Wang, and P. He. (2008) Relatedness of resistance to anthracnose and to white rot in Chinese wild grapes. *Vitis* 47(4), 213-215. [This manuscript explores Chinese wild grapes for resistance to anthracnose and white rot.]

Liang, Z., C. Yang, J. Yang, B. Wu, L. Wang, J. Cheng, and S. Li. (2009) Inheritance of anthocyanins in berries of *Vitis vinifera* grapes. *Euphytica* 167, 113-125. [Inheritance of anthocyanins in grape berries is described in this article.]

Loomis, N.H., and J.H. Weinberger. (1979) Inheritance studies of seedlessness in grape. *Journal of American Society for Horticultural Science* 104, 181-184. [This manuscript shows analysis of inheritance of seedlessness in grape.]

Mehta-Bhatt, P. (2007) Utilization of genetic engineering in commercial crops: A global overview. Developed under the auspices of the Arab Authority for Agricultural Investment and Development, Abu Dhabi, United Arab Emirates, [<http://www.aaaid.org>] [This article provides an overview of plant genetic engineering.]

Meredith, C.P. (2001) Grapevine genetics: Probing the past and facing the future. *Agriculturae Conspectus Scientificus* 66(1), 21-25. [This review describes the applications of DNA markers in the grapevine.]

Mezzett, B., T. Pandolfini, O. Navacchi, and L. Landi. (2002) Genetic transformation of *Vitis vinifera* via organogenesis. *BMC Biotechnology* 2(18), 1-10. [Role of *DefH9-iaaM* gene in young flower buds is described in this article.]

Mikio, S., Y. Masahiko, M. Nobuhito, U. Toshihito, N. Ryoji, and N. Masaaki. (2006) Rapid screening assay for ripe rot resistance in grape cultivars. *Journal of the Japanese Society for Horticultural Science* 75(3), 264-266. [This manuscript describes development of rapid screening assay for ripe rot resistance in grape cultivars.]

Nikolić, D. (2006) Components of variance and heritability of resistance to important fungal diseases agents in grapevine. *Journal of Agricultural Science* 51(1), 47-54. [This article indicates heritability of fungal resistance in grapevine.]

Ocete, R., M.Á. López, A. Gallardo, and C. Arnold. (2008) Comparative analysis of wild and cultivated grapevine (*Vitis vinifera*) in the Basque Region of Spain and France. *Agriculture, Ecosystems & Environment* 123, 95-98. [This manuscript shows the pest infestation levels of wild and cultivated grapevine.]

Olmo, H.P. (1951) Breeding tetraploid grapes. *In Proceedings of Investigations in Polyploidy of Fruits Symposium*. 12 Sept., 1951. Montpellier, France. [This article presents the characterization of tetraploid grapes.]

Owens, C.L. (2008) Grapes. *In Temperate Fruit Crop Breeding: Germplasm to Genomics*. (ed. J.F. Hancock). New York: Springer Science+Business Media. [This chapter emphasizes on application of genomic analysis and molecular markers in grapevine breeding.]

Pavloušek, P. (2007) Evaluation of resistance to powdery mildew in grapevine genetic resources. *Journal of Central European Agriculture* 8(1), 105-114. [This article is about screening grapevine genetic resources for powdery mildew resistance.]

Pazzi, F. (2008) Genetically modified grapevine: State of research, possible risks and future scenario. *In* Published on-line: March 2008, developed under the auspices of the Fondazione Diritti Genetici, Roma, Italy, [<http://www.fondazioneirittigenetici.org>] [This article presents recent development of transgenic grapevine.]

Peressotti, E., S. Wiedemann-Merdinoglu, F. Delmotte, D. Bellin, G.D. Gaspero, R. Testolin, D. Merdinoglu, and P. Mestre. (2010) Breakdown of resistance to grapevine downy mildew upon limited deployment of a resistant variety. *BMC Plant Biology* 10(147), 1-11. [This research covers the breakdown of downy mildew resistance in grapevine.]

Perl, A., N. Sahar, P. Spiegel-Roy, S. Gavish, R. Elyasi, E. Orr, and H. Bazak. (2000) Conventional and biotechnological approaches in breeding seedless table grapes. *Acta Horticulturae* 528, 607-612. [This article deals with conventional and biotechnological approaches for seedless table grape improvement.]

Pernes, G. (2004) New resistant table grape cultivars bred in Hungary. *Acta Horticulturae* 652, 321-327. [This manuscript aims to develop new table grape cultivars resistant to downy mildew, powdery mildew and grey rot, with high cold hardiness.]

Péros, J.P., T.H. Nguyen, C. Troulet, C. Michel-Romiti, and J.L. Notteghem. (2006) Assessment of powdery mildew resistance of grape and *Erysiphe necator* pathogenicity using a laboratory assay. *Vitis* 45(1), 29-36. [This article deals with a laboratory assay to assess powdery mildew resistance.]

Pierquet, P., and C. Stushnoff. (1980) Relationship of low temperature exotherms to cold injury in *Vitis riparia* Michx. *American Journal of Enology and Viticulture* 31(1), 1-6. [This article covers cold tolerance in *V. riparia* Michx.]

Punja, Z.K. (2001) Genetic engineering of plants to enhance resistance to fungal pathogens – a review of progress and future prospects. *Canadian Journal of Plant Pathology* 23, 216-235. [This review explains mechanisms of fungal resistance in transgenic plants.]

Ramming, D.W., M.A. Walker, A. Tenschler, and A.F. Krivanek. (2009) Breeding table and raisin grapes with increased fruit quality while retaining Pierce's disease resistance. *Acta Horticulturae* 827, 445-450. [This article deals with breeding for increased fruit quality in table and raisin grapes resistant to Pierce's disease.]

- Ray, P.K. (2002) Grape. *In* Breeding Tropical and Subtropical Fruits. New Delhi: Narosa publishing house. [This chapter summarizes different methods related to grapevine breeding, especially in India.]
- Reisch, B.I. (1998) Molecular markers – the foundation for grapevine genetic mapping, DNA fingerprinting and genomics. *In* Proceedings of the 7<sup>th</sup> International Symposium on Grapevine Genetics and Breeding. 6-10 July, 1998. Montpellier, France. [This report explains types of molecular markers and their applications in plant breeding.]
- Reisch, B.I., and C. Pratt. (1996) Grapes. *In* Fruit Breeding, Vol. II: Vine and Small Fruit Crops. (eds. J. Janick and J.N. Moore) New York: John Wiley & Sons, Inc. [This chapter reviews several aspects of grapevine breeding.]
- Ren, Z., and J. Lu. (1999) Inheritance of berry size, color and flower sex in muscadine grapes. *Proceedings of the Florida State Horticultural Society* 112, 167-168. [This manuscript describes the inheritance of berry size, color and flower sex in muscadine grapes.]
- Roh, J.H., K.K. Yun, K.S. Park, C.H. Lee, and S.B. Jeong. (2003) *In vivo* evaluation of resistance of grape varieties to crown gall disease. *Journal of Plant Pathology* 19(5), 235-238. [This manuscript describes the evaluation of crown gall resistance in grape varieties.]
- Rosenfield, C.L., S. Samuelian, J.R. Vidal, and B.I. Reisch. (2010) Transgenic disease resistance in *Vitis vinifera*: Potential use and screening of antimicrobial peptides. *American Journal of Enology and Viticulture* 61(3), 348-357. [This article is about transgenic grapevine with increased disease resistance.]
- Roytchev, R. (1998) Inheritance of grape seedlessness in seeded and seedless hybrid combinations of grape cultivars with complex genealogy. *American Journal of Enology and Viticulture* 49 (3), 302-305. [This article describes the inheritance of grape seedlessness.]
- Savin, G. (2010) Conservation and using of grapevine genetic resources in the Republic of Moldova for pre-breeding stage. *Romanian Biotechnology Letters* 15(2), 113-116. [This manuscript discusses the maintenance and usage of grapevine genetic resources.]
- Scorza, R., W.V. Kearneysville, J.M. Cordts, D.J. Gray, D. Gonsalves, R.L. Emershad, and D.W. Ramming. (1996) Producing transgenic 'Thompson Seedless' grape (*Vitis vinifera* L.) plants. *Journal of the American Society for Horticultural Science* 121, 616-619. [This article reports the transformation of grapevine with the TomRSV coat protein gene using *A. tumefaciens*.]
- Semagn, K., Å. Bjørnstad, and M.N. Ndjiondjop. (2006) Principles, requirements and prospects of genetic mapping in plants. *African Journal of Biotechnology* 5(25), 2569-2587. [This review explains principles and application of genetic mapping for crop improvement.]
- Sestras, R., S.D. Moldovan, and C.F. Popescu. (2008) Variability and heritability of several important traits for grape production and breeding. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca* 36(1), 88-97. [This manuscript reports measurement of variability and heritability of many traits including downy mildew resistance.]
- Shiraishi, M., M. Koide, H. Itamura, M. Yamada, N. Mitani, T. Ueno, R. Nakaune, and M. Nakano. (2007) Screening for resistance to ripe rot caused by *Colletotrichum acutatum* in grape germplasm. *Vitis* 46(4), 196-200. [This article shows screening for ripe rot resistance in grapevine germplasm.]
- Shiraishi, M., H. Fujishima, and H. Chijiwa. (2010) Evaluation of table grape genetic resources for sugar, organic acid and amino acid composition of berries. *Euphytica* 174, 1-13. [Evaluation of table grape genetic resources for fruit quality is shown in this article.]
- Sim, S.T. (2006) Virus elimination from grape selections using tissue culture. *FPS Grape Program Newsletter*, pp. 30-31. [This article shows the utilization of tissue culture to obtain virus-free plants.]
- Slimane, M.H.B., H. Snoussi, R. Bouhlal, and H. Nahdi. (2010) Ampelometry to test for genetic diversity in Tunisian *Vitis sylvestris*. *The African Journal of Plant Science and Biotechnology* 4(2), 17-22. [This manuscript describes the genetic diversity analysis of wild grape.]
- Snyder, E., and F.N. Harmon. (1936) Hastening the production of fruit in grape hybridizing work. *Journal of the American Society for Horticultural Science* 34, 426-427. [This article shows how to shorten the juvenile period of grapevine.]

Srinivasan, C., and M.G. Mullins. (1981) Physiology of flowering in the grapevine – a review. *American Journal of Enology and Viticulture* 32(1), 47-63. [Flowering process of the grapevine is reviewed in this article.]

Stafne, E. (2011) Interspecific hybrid grapes. *In* Published on-line: May 2011, developed under the auspices of the eXtension, United States, [<http://www.extension.org>] [This article presents information on interspecific hybrid grapevines.]

Stout, A.B. (1939) Progress in breeding for seedless grapes. *Proceedings of the American Society for Horticultural Science* 37, 627-629. [Progress in breeding for seedless grapes is discussed in this article.]

Suslow, T.V., B.R. Thomas, and K.J. Bradford. (2002) Biotechnology provides new tools for plant breeding. *Agricultural Biotechnology in California Series Publication* 8043. 19 p. [This article provides detailed information on recombinant DNA techniques and genetic engineering.]

Süle, S., and T.J. Burr. (1998) The effect of resistance of rootstocks to crown gall (*Agrobacterium* spp.) on the susceptibility of scions in grapevine cultivars. *Plant Pathology* 47, 84-88. [This manuscript discusses the use of resistant rootstocks to protect susceptible scions from crown gall.]

Swenson, E.P. (1985) Wild *Vitis riparia* from north U.S. and Canada: Breeding source for winter hardiness in cultivated grapes – a background of the Swenson hybrids. *MGGA. 1985 Annual Report*, pp. 5-11. [This article is about breeding source for cold tolerance in cultivated grapes.]

Tesniere, C., L. Torregrosa, M. Pradal, J.-M. Souquet, C. Gilles, K.D. Santos, P. Chatelet, and Z. Gunata. (2006) Effects of genetic manipulation of alcohol dehydrogenase levels on the response to stress and the synthesis of secondary metabolites in grapevine leaves. *Journal of Experimental Botany* 57(1), 91-99. [This manuscript describes the effects of genetic manipulation of alcohol dehydrogenase levels.]

This, P., T. Lacombe, and M.R. Thomas. (2006) Historical origins and genetic diversity of wine grapes. *Trends in Genetics* 22(9), 511-519. [This is an article dealing with the wine grape germplasm resources.]

Tian, L., Y. Wang, L. Niu, and D. Tang. (2008) Breeding of disease-resistant seedless grapes using Chinese wild *Vitis* spp. I. *In vitro* embryo rescue and plant development. *Scientia Horticulturae* 117, 136-141. [This article presents efforts to breed for seedless grapes with the help of embryo rescue technique.]

Töpfer, R. (2010) Grape germplasm resources. *In* *Grape Genetic Resources*, developed under the auspices of the International Grape Genome Program, [<http://www.vitaceae.org>] [This website shows grape germplasm resources.]

Troggio, M., G. Malacarne, G. Coppola, C. Segala, D.A. Cartwright, M. Pindo, M. Stefanini, R. Mank, M. Moroldo, M. Morgante, M.S. Grando, and R. Velasco. (2007) A dense single-nucleotide polymorphism-based genetic linkage map of grapevine (*Vitis vinifera* L.) anchoring Pinot Noir bacterial artificial chromosome contigs. *Genetics* 176: 2637-2650. [This article reports the construction of high resolution linkage map using SNP, SSR and AFLP.]

Tsuchihira, A., T. Yuko, Y.T. Hanba, N. Kato, T. Doi, T. Kawazu, and M. Maeshima. (2010) Effect of overexpression of radish plasma membrane aquaporins on water-use efficiency, photosynthesis and growth of *Eucalyptus* trees. *Tree Physiology* 30(3), 417-430. [This article describes the expression of *PIP* gene in transgenic plants to improve drought tolerance.]

Uzun, I.H., and A. Bayir. (2010) Distribution of wild and cultivated grapes in Turkey. *Notulae Scientia Biologicae* 2(4), 83-87. [This manuscript aims to describe the distribution of wild and cultivated grapes in Turkey.]

Velasco, R., A. Zharkikh, M. Troggio, D.A. Cartwright, A. Cestaro, D. Pruss, M. Pindo, L.M. FitzGerald, S. Vezzulli, J. Reid, G. Malacarne, D. Iliev, G. Coppola, B. Wardell, D. Micheletti, T. Macalma, M. Facci, J.T. Mitchell, M. Perazzolli, G. Eldredge, P. Gatto, R. Oyzerski, M. Moretto, N. Gutin, M. Stefanini, Y. Chen, C. Segala, C. Davenport, L. Demattè, A. Mraz, J. Battilana, K. Stormo, F. Costa, Q. Tao, A. Si-Ammour, T. Harkins, A. Lackey, C. Perbost, B. Taillon, A. Stella, V. Solovyev, J.A. Fawcett, L. Sterck, K. Vandepoele, S.M. Grando, S. Toppo, C. Moser, J. Lanchbury, R. Bogden, M. Skolnick, V. Sgaramella, S.K. Bhatnagar, P. Fontana, A. Gutin, Y. Van de Peer, F. Salamini, and R. Viola. (2007) A high quality draft consensus sequence of the genome of a heterozygous grapevine variety. *PLoS ONE* 12, 1-18. [This manuscript describes a comprehensive sequencing and annotation of genes in grapevine genome.]

Viegas, P.M., and N.K. Notani. (1993) Heritable transformation of tobacco by *Agrobacterium* -mediated transfer of the *Streptomyces*-derived herbicide resistance gene *bar*. *Journal of Genetics* 72(1), 35-42. [This manuscript describes usage of *Agrobacterium*-mediated transformation to transfer the *bar* gene to tobacco.]

Vivier, M.A., and I.S. Pretorius. (2000) Genetic improvement of grapevine: Tailoring grape varieties for the third millennium – a review. *South African Journal for Enology and Viticulture* 21, 5-25. [This review discusses the applications of biotechnology in grapevine improvement.]

Vivier, M.A., and I.S. Pretorius. (2002) Genetically tailored grapevines for the wine industry. *Trends in Biotechnology* 20(11), 472-478. [This review describes the use of genetic engineering for wine grape improvement.]

Wakana, A., M. Hiramatsu, S.M. Park, N. Hanada, I. Fukudome, and K. Yasukochi. (2003) Seed abortion in crosses between diploid and tetraploid grapes (*Vitis vinifera* and *V. complex*) and recovery of triploid plants through embryo culture. *Journal of the Faculty of Agriculture Kyushu University* 48(1-2), 39-50. [This article shows the use of embryo culture to obtain seedless triploid grapes.]

Walker, G.E., and G.R. Stirling. (2008) Plant-parasitic nematodes in Australian viticulture: Key pests, current management practices and opportunities for future improvements. *Australasian Plant Pathology* 37(3), 268-278. [This article provides information on management of plant-parasitic nematodes in Australian viticulture.]

Wan, Y., P. He, and Y. Wang. (2007a) Inheritance of downy mildew resistance in two interspecific crosses between Chinese wild grapes and European grapes. *Vitis* 46(3), 156-157. [This article is about interspecific crosses between Chinese wild grapes and European grapes to transfer downy mildew resistance.]

Wan, Y., H. Schwaninger, P. He, and Y. Wang. (2007b) Comparison of resistance to powdery mildew and downy mildew in Chinese wild grapes. *Vitis* 46(3), 132-136. [This manuscript shows powdery mildew and downy mildew resistance in various Chinese wild grapes.]

Wan, Y., H. Schwaninger, D. Li, C.J. Simon, Y. Wang, and C. Zhang. (2008a) A review of taxonomy research on Chinese wild grapes. *Vitis* 47(2), 81-88. [Taxonomy on Chinese wild grapes is reported in this paper.]

Wan, Y., Y. Wang, D. Li, and P. He. (2008b) Evaluation of agronomic traits in Chinese wild grapes and screening superior accessions for use in a breeding program. *Vitis* 47(3), 153-158. [This article deals with the evaluation of agronomic traits in Chinese wild grapes.]

Welter, L.J., N. Gökürk-Baydar, M. Akkurt, E. Maul, R. Eibach, R. Töpfer, and E.M. Zyprian. (2007) Genetic mapping and localization of quantitative trait loci affecting fungal disease resistance and leaf morphology in grapevine (*Vitis vinifera* L.). *Molecular Breeding* 20, 359-374. [This article describes QTLs for downy mildew resistance in 'Regent'.]

Wikipedia: The Free Encyclopedia. Developed under the auspices of the Wikimedia Foundation, Inc., [<http://en.wikipedia.org>] [This website provides comprehensive definitions of technical words.]

Wolpert, J.A., and G.S. Howell. (1985) Cold acclimation of concord grapevines. I. Variation in cold hardiness within the canopy. *American Journal of Enology and Viticulture* 36(3), 185-188. [This article describes cold acclimation of concord grapevines.]

Yasmin, T. (2009) Cloning and over-expression of a germin-like protein gene for its functional analysis. Rawalpindi, Pakistan Arid Agriculture University, Ph.D. Thesis. [This work shows cloning and over-expression of a germin-like protein gene.]

Zhangsun, D., S. Luo, R. Chen, and K. Tang. (2007) Improved *Agrobacterium*-mediated genetic transformation of GNA transgenic sugarcane. *Cell and Molecular Biology* 62(4), 386-393. [This is a manuscript on improvement of insect resistance using *Agrobacterium*-mediated genetic transformation.]

Zok, A., A. Pedryc, and R. Olah. (2007) Genetic transformation experiments applying useful gene constructions for grapevine growing. In 30<sup>th</sup> World Congress of Vine and Wine. 10-16 June, 2007. Kongresszusa, Budapest. [This article describes experiments conducted to improve abiotic stress tolerance of the grapevine by genetic engineering.]

## Biographical Sketches

**Piyada Tantasawat** was born in Bangkok, Thailand, and obtained her B.Sc. in Agriculture (Horticulture) at Kasetsart University with First Class Honors. She worked as a research assistant at Kasetsart University for nearly a year before receiving a Fulbright Scholarship to pursue her M.Sc. at Cornell University where she obtained her Ph.D. in Plant Breeding in 1997. After three years of postdoctoral research in plant genetic engineering and biochemistry at Cornell University, she returned to Thailand and was appointed to the staff of the School of Crop Production Technology, Institute of Agricultural Technology, Suranaree University of Technology (SUT), Thailand. Her major scientific interests are breeding for plant pest resistance utilizing conventional, mutation, and biotechnological approaches, plant tissue culture, and plant resistance mechanisms. She has worked on grapevine, mungbean, sunflower, cucumber, tomato and orchid improvement, and published over 40 papers that are related to these interests.

**Oythip Poolsawat** was born in Nakhon Nayok, Thailand. She has a B.Sc. (Second Class Honors) and a Ph.D. in Crop Production Technology from SUT. She is currently a post-doctoral fellow at SUT. She is interested in the resistance mechanisms and breeding of grapevines for resistance to diseases, especially anthracnose.

**Wirot Chaowiset** was born in Suphanburi, Thailand, and has a B.Sc. and an M.Sc. in Crop Production Technology from SUT, Thailand. His thesis study was on physiology of *Pueraria mirifica* and the effect of the active ingredients in *P. mirifica* on the relaxation of blood vessels in rats. After graduation (2007), he worked as a research assistant in the sunflower breeding project. Currently, he is a post-graduate researcher at SUT. He has performed research on many crops, including tomatoes, cucumbers, grapevines and orchids etc. He is interested in plant physiology, mechanisms of resistance, and crop improvement via tissue culture and other techniques.